



## Review

Fusidic acid for the treatment of bone and joint infections caused by methicillin-resistant *Staphylococcus aureus*

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## ABSTRACT

There is a lack of surveillance data on resistance to fusidic acid (FA) in Asia, and no reviews of FA usage for the treatment of orthopaedic infections have been conducted since the year 2000. In this study, we present a systemic literature review of FA resistance in Asia and the clinical use of FA for the treatment of bone and joint infections (BJIs). The in vitro activity of FA against methicillin-resistant *Staphylococcus aureus* (MRSA) isolates remains good, with low (<10%) resistance rates in most Asian countries. FA in Asia appears to be a better oral anti-MRSA agent than trimethoprim/sulfamethoxazole and clindamycin. More than 80 cases of FA use for BJI have been reported since 2000 and the recurrence or failure rate is <10%. There is much evidence supporting the use of FA in combination with other antibiotics (e.g. rifampicin) as an oral treatment following intravenous glycopeptide treatment for BJIs.

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## 1. Introduction

There is a need for safe, oral antibiotics against methicillin-resistant *Staphylococcus aureus* (MRSA) for effective step-down therapy after hospitalisation or for initial therapy for community-acquired infections [1]. Fusidic acid (FA), manufactured by Leo Pharmaceuticals, has been widely used in Europe, Canada, Australia and some Asian countries for decades. Although FA has not been approved for use in the USA by the US Food and Drug Administration (FDA), Cempra Pharmaceuticals has designed a dosing regimen

for the use of FA as monotherapy if and when it is approved for use in the USA [1,2].

A number of review papers on FA were published in the *International Journal of Antimicrobial Agents* in the 1990s [3–7] and, in 2010, Schöfer and Simonsen reviewed the clinical efficacy of FA for skin and soft-tissue infections (SSTIs) [8]. The evidence provided in those review papers showed that FA is an effective treatment for SSTIs, acute osteomyelitis, chronic osteomyelitis, vertebral infection, septic arthritis, and prosthetic and other device-related infections due to methicillin-susceptible *S. aureus* (MSSA). There are few oral antibiotics listed in the treatment guidelines for treatment of MRSA infections [3–8]. The therapeutic guidelines in Australia list the combination of rifampicin (RIF) and FA as a treatment option for recurrent staphylococcal skin infections (including

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MRSA-positive infections) and MRSA osteomyelitis involving the bone or joint prostheses, both in adult and paediatric patients [9].

Treatment guidelines in the UK recommend FA and RIF for bone and joint infections (BJIs) [10]. In several reviews on BJIs, FA is listed as the treatment of choice [11–15].

Here we provide a review of FA resistance patterns in Asia and focus on the use of FA for the treatment of BJIs due to MRSA after the year 2000. Comprehensive searches were conducted of Medline, PubMed, Google Scholar, CINAHL, EMBASE and the CNKI (Chinese National Knowledge Infrastructure) database as well as reference lists of retrieved articles. Search terms were ‘fusidic acid’, ‘*Staphylococcus aureus*’, ‘MRSA’, ‘osteomyelitis’, ‘septic arthritis’ and ‘bone and joint infection’, with each of the target country names in turn. Studies with a percentage of FA resistance in MRSA in Chinese articles are included.

## 2. Pharmacology

FA inhibits polypeptide chain elongation by binding to the ribosome elongation factor G (EF-G)–GDP complex. FA has good oral bioavailability and is metabolised and excreted by the liver [16]. This bacteriostatic agent is highly protein-bound and has been shown to have good concentrations in soft tissue, bone and synovial fluid [3,17,18]. FA is active against several *Staphylococcus* spp., including MSSA and MRSA, but has poor activity against *Streptococcus pyogenes* [4]. Rates of resistance to FA are higher among coagulase-negative staphylococci (CoNS) than among *S. aureus* [4,19–21]. The recommended dose of FA is 250–500 mg every 8–12 h; however, new pharmacokinetic data show that front-loaded dosing [ $\geq 1200$  mg every 12 h (q12h)  $\times$  2 doses followed by  $\geq 600$  mg q12h] has better activity against MRSA and *S. pyogenes* than non-front-loaded dosing (600 mg q12h) [2].

## 3. In vitro activity of fusidic acid in combination with other antibiotics

Most in vitro and in vivo studies on FA combinations have concentrated on the effects of FA with RIF. Other common oral combinations with FA include minocycline, linezolid and fosfomycin. Checkerboard dilution and time–kill methods have revealed that FA combined with RIF had partial synergistic effects [22–26]. A combination of FA and minocycline has been reported for the treatment of vancomycin-intermediate *S. aureus* (VISA) infection, but there is a lack of in vitro data [27]. FA combined with linezolid has been successful in the treatment of VISA infection and endocarditis [28–30]. When combined with FA, linezolid was shown to prevent selection of resistant mutants but showed no synergy in one study [31]. Saginur et al. found that RIF and FA, plus either ciprofloxacin or vancomycin, were consistently bactericidal against MRSA biofilms [32]. Another study in China showed 88% synergism when FA was combined with fosfomycin [33]. Tang et al. found that the in vitro killing effects of fosfomycin combined with FA were better than the killing effects of RIF combined with FA [34].

## 4. Fusidic acid resistance in North America and Europe

The prevalence of resistance to FA in *S. aureus* remained low well into the 1990s. In staphylococci, high-level FA resistance is usually caused by mutations in *fusA*, the gene encoding EF-G, and low-level resistance is generally caused by the horizontally transferable genes *fusB* and *fusC* [35–37]. FusB-type proteins bind to EF-G on the ribosome, which allows the ribosomes to resume translation. There is also a concern that recent exposure to topical FA is correlated with the presence of FA-resistant *S. aureus* [38–40].

In addition, there is also a significant trend towards increased FA resistance among *S. aureus* with increased duration of use [40,41].

Updated surveillance data on FA resistance in MRSA have been reported in studies from North America, Australia and Europe [20,21,41,42,36]. *Staphylococcus aureus* resistance rates are very low in the USA (0.3%) and are relatively high in Canada and Australia (ca. 7.0%) [20,21]. In Canada, the occurrence of *fusB* and *fusC* is similar among *S. aureus*, whereas in Australia *S. aureus* isolates tend to be *fusC*-positive [20,21].

The increase in FA resistance in Europe appears to be due to clonal expansion of a strain called the epidemic European FA-resistant impetigo clone. This clone has been reported in the UK, France and other European countries [43–45]. In an updated surveillance report on FA resistance in 13 European countries, 10.7% of *S. aureus* isolates displayed FA minimum inhibitory concentrations (MICs)  $\geq 2$  mg/L [41]. Israel, Italy, Poland, Spain and Sweden had low rates (1.4–3.1%) of FA resistance [41]. Greece (62.4%) and Ireland (19.9%) had the highest resistance rates. Many of the FA-resistant *S. aureus* isolates in Greece and Ireland were spread clonally and showed high fusidic MIC values ( $\geq 512$  mg/L), especially among isolates that carried the *fusA* L461K mutation [41]. It is unclear whether the emerging FA resistance patterns seen in some European countries are also present in Asia [37,41,42], mainly because there is an overall lack of surveillance data in Asian countries.

## 5. Fusidic acid resistance in Asia

Resistance rates to FA in Asian countries are shown in Table 1 (references S1–S31 in Supplementary data). Resistance rates from 2000 to the present are available for 13 Asian countries, namely China, Japan, South Korea, Taiwan, Hong Kong, Singapore, Malaysia, Thailand, Cambodia, Kuwait, Iran, Saudi Arabia and Pakistan. Resistance was determined by the disk diffusion method in most of those studies. In most Asian countries, with the exception of Kuwait, Pakistan and South Korea, resistance rates are relatively low (<10%). In general, clindamycin resistance is high in Asia (>60%). Although most isolates are more susceptible to trimethoprim than to clindamycin, the rates of resistance to these two antimicrobial agents are higher than those to FA. Few studies have provided data on resistance to doxycycline and minocycline (data not shown).

In Kuwait, most MRSA isolates have high FA MICs (>256 mg/L), possibly because of the circulation of epidemic clones. Transmission of these clones and their maintenance in different hospitals may explain their high prevalence in Kuwaiti hospitals. The circulating strains with sequence type (ST) 80 commonly isolated from patients with community-acquired (CA) MRSA in Kuwait are not seen in other Asian countries. In contrast, ST59 is the most common CA-MRSA strain, and ST239 is the most common nosocomial MRSA strain in most Asian countries. In contrast to other CA-MRSA clones, the ST80 clone commonly carries the *fusB* gene, which appears to confer reduced susceptibility to FA. Studies from Greece and Kuwait have reported a high incidence of ST80 among CA-MRSA isolates and high rates of FA resistance [46,47]. According to recent data gathered by the Asian Network for Surveillance of Resistant Pathogens (ANSORP), which is a prospective, multinational surveillance study, ST80 has not been isolated in any Asian country [48]. ST80 is common in the Mediterranean and Balkan regions as well as in the Middle East. The reason for the relatively high prevalence of FA resistance in Pakistan and South Korea is not known. FA resistance among MSSA is also high in South Korea [49,50].

In Kuwait, the rate of FA resistance increased dramatically from 22% in 1994 to 92% in 2004 [51]. In Malaysia, the rate of resistance to FA among MRSA isolates increased from 3–5% during the

**Table 1**  
Resistance profiles of fusidic acid (FA) in Asian countries.

Country	Reference <sup>a</sup>	Study year	Susceptibility testing method	No. of isolates tested	% of resistant isolates			
					FA	CLI	SXT	RIF
<b>High FA resistance</b>								
South Korea	Lee et al. [S1]	1996	DDM	90	12.2	86.7	N/A	N/A
South Korea	Kim et al. [S2]	1999–2001	DDM	439	14.1	84.3	8.9	18.0
South Korea	Kim et al. [S3]	2008	ADM	40	52.5	50	10	17.5
Kuwait	Udo et al. [S4]	2004	DDM	930	92	N/A	27	4.7
Kuwait	Udo et al. [S5]	2005	DDM	588	84	N/A	81	1
Pakistan	Idrees et al. [S6]	2005–2007	DDM	501	9	79	59	50
Pakistan	Zafar et al. [S7]	2006–2007	DDM	126	18	N/A	43	N/A
Pakistan	Shabir et al. [S8]	2010	DDM	60	20	N/A	N/A	52
<b>Low FA resistance</b>								
Cambodia	Chheng et al. [S9]	2006–2007	DDM	17	0	77	88	N/A
China <sup>b</sup>	Chen et al. [S10]	2001–2003	DDM	50	2.0	N/A	36	14
China <sup>b</sup>	Ni [S11]	2002	DDM	57	1.7	95.9	23.5	22.7
China <sup>b</sup>	Chang [S12]	2002–2003	DDM	280	1.7	95.9	23.5	22.7
China <sup>b</sup>	Pan [S13]	2002–2003	DDM	50	12	100	40	10
China	Liu et al. [S14]	2003–2007	ADM	11	0	90.9	N/A	N/A
China <sup>b</sup>	Hu et al. [S15]	2006–2007	DDM	56	1.8	92.8	25.0	N/A
China	Liu et al. [S16]	2008–2009	DDM	66	3	N/A	N/A	N/A
Hong Kong	Ip et al. [S17]	2000–2001	DDM	200	1	67.5	6	N/A
Iran	Askarian et al. [S18]	2006	DDM	32	0	69	N/A	3
Iran	Japoni et al. [S19]	2008–2009	Etest	156	0	76	69	11
Japan	Takizawa et al. [S20]	2003	ADM	54	0	N/A	N/A	0
Japan	Nakaminami et al. [S21]	2006	DDM	76	1.3	35.5	N/A	N/A
Malaysia	Norazah et al. [S22]	1997–1999	DDM	640	5	N/A	N/A	5
Malaysia	Thong et al. [S23]	2003–2007	DDM	66	11	19	N/A	12
Taiwan	Chen et al. [S24]	1995–2006	ADM	257	0	88	75.8	N/A
Taiwan	Lin et al. [S25]	2003–2007	DDM	94	1.1	85.6	4.8	N/A
Taiwan	Lo et al. [S26]	2004–2006	DDM	131	0	88.5	0	0
Taiwan	Lo et al. [S26]	2007–2009	DDM	240	0	89.6	2.9	1.3
Thailand	Hortiwakul et al. [S27]	2000–2001	Etest	100	0	N/A	N/A	N/A
Thailand	Mekviwattanawong et al. [S28]	2005	DDM	184	6.1	91.4	85.9	53.8
Thailand	Nickerson et al. [S29]	2006–2007	DDM	23	13	65	96	35
Saudi Arabia	Baddour et al. [S30]	2004–2005	Etest	512	4.3	N/A	33.8	N/A
Singapore	Hsu et al. [S31]	2005	DDM	197	<5	N/A	N/A	<5

CLI, clindamycin; SXT, trimethoprim/sulfamethoxazole; RIF, rifampicin; DDM, disk diffusion method; N/A, not available; ADM, agar dilution method.

<sup>a</sup> References are given in the [Supplementary data](#).

<sup>b</sup> Article in Chinese.

period 1992–1996 to 11% in 2009 [52]. In Singapore, the rate of FA resistance was <5% during 1997–2004 [53]. Data from one medical centre in Taiwan revealed that FA usage remained stable from 2002 to 2009 and that the prevalence of FA-resistant Gram-positive bacteria causing healthcare-associated infections also remained stable (ca. 3–7%) [54].

Only three papers have been published on FA resistance gene determinants in Asia. In one study from central Taiwan, the most common FA resistance determinant in 34 isolates was *fusC* (74%) [55]. In northern Taiwan, Chen et al. found that 84% of 45 FA-resistant MRSA isolates had *fusA* mutations [56]. A small study in China showed that *fusB* and *fusC* were the main resistance determinants in four FA-resistant clinical isolates [57].

## 6. Fusidic acid in bone and joint infections caused by methicillin-resistant *Staphylococcus aureus*

No randomised controlled trials of FA as treatment for BJIs due to MRSA have been conducted. Trampuz and Zimmerli recommended a 2-week regimen of vancomycin and RIF followed by FA for treatment of prosthetic joint infections due to MRSA [13]. Oral antimicrobial therapy for osteomyelitis is important given the associated costs and potential morbidity of prolonged courses of intravenous (i.v.) therapy [14]. Unfortunately, the only oral antibiotics that are active against MRSA include pristinamycin, linezolid, trimethoprim/sulfamethoxazole (SXT), doxycycline and FA.

Table 2 presents the results of the literature search for reviews of treatment of osteomyelitis and septic arthritis [30,58–70]. The

three large series of FA use for BJIs involved prosthetic joint infections with or without implant removal [63,67,69]. In most cases, FA was administered along with RIF following administration of i.v. glycopeptides. Other oral antibiotics that are used in combination with FA for the treatment of osteomyelitis and septic arthritis with or without removal of the prosthesis include chloramphenicol, doxycycline, linezolid and pristinamycin. In general, oral FA is appropriate for long-term use. In more than 80 reported cases of infections treated with FA, only 7 patients (<10%) had treatment failure or recurrence. In one study of a two-stage revision in total knee arthroplasty infected with MRSA (89%) or methicillin-resistant CoNS (11%) in Taiwan [69], i.v. vancomycin or teicoplanin was used for  $\geq 2$  weeks or until clinical infection control was observed and C-reactive protein (CRP) had decreased to <2.0 mg/dL. Oral sodium fusidate tablets were then administered for  $\geq 4$  weeks until the CRP value returned to normal. The criteria for re-implantation were CRP < 1.0 mg/dL and no clinical signs of infection 2 weeks after discontinuing the oral regimen. In that series, there were only three cases of recurrent infection [69]. In a retrospective cohort study of patients with MRSA orthopaedic device-related infections between 2000 and 2008 at Geneva University Hospital (Switzerland), none of the patients who received RIF plus FA ( $n=12$ ) experienced treatment failure [67]. In a large series from Australia involving patients with MRSA prosthetic joint infections treated with surgical debridement and prosthesis retention, RIF combined with FA was administered for a mean duration of 12 months (range 6–33 months) and only 1 of 11 MRSA infections recurred [63]. Other case reports of FA use for the treatment of BJIs caused by MRSA after 2000 are listed in Table 2 [30,58–70].

**Table 2**  
Literature review of fusidic acid use for the management of bone and joint infections caused by methicillin-resistant *Staphylococcus aureus*.

Reference	Publication year	Country	No. of patients	Prosthetic device	Duration of i.v. antibiotic treatment	Oral antibiotic in combination	No. of patients with treatment failure or recurrence
Howden et al. [30]	2004	Australia	4	Yes	N/R	RIF, CHL	1
Ng and Gosbell [58]	2005	Australia	4	No	N/R	PRI	0
Nather et al. [59]	2005	Singapore	3	No	6–8 weeks	CLI	0
Chiang et al. [60]	2005	Taiwan	1	No	4 weeks	None	0
Donaldson et al. [61]	2006	Australia	2	No	7 weeks	RIF	2
Inverarity et al. [62]	2006	UK	1	No	6 weeks	DOX	0
Aboltins et al. [63]	2007	Australia	11	Yes	1–4 weeks	RIF	1
Apisarnthanarak and Mundy [64]	2007	Thailand	1	No	2 weeks	RIF	0
Murray et al. [65]	2008	Australia	7	No	N/R	RIF	0
Ahamed Puthiyaveetil [66]	2009	Australia	1	No	N/R	RIF	0
Ferry et al. [67]	2010	Switzerland	12	Yes	N/R	RIF	0
O'Neill et al. [68]	2011	Ireland	1	No	2 weeks	LZD	0
Chiang et al. [69]	2011	Taiwan	39	Yes	2–4 weeks	N/R	3
Wolfe [70]	2011	USA	1	No	10 weeks	None	0

i.v., intravenous; N/R, not reported; RIF, rifampicin; CHL, chloramphenicol; PRI, pristinamycin; CLI, clindamycin; DOX, doxycycline; LZD, linezolid.

## 7. Conclusions

The in vitro activity of FA in Asia is good and most countries, with the exception of Kuwait, South Korea and Pakistan, have low resistance rates (<10%). FA appears to be a better oral anti-MRSA agent than SXT, doxycycline/minocycline and linezolid. For the treatment of MRSA BJIs, most studies support the use of FA in combination with other antibiotics (e.g. RIF) as step-down oral antibiotic regimens following administration of i.v. glycopeptides.

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## Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.ijantimicag.2012.03.010>.

## References

- Fernandes P, Pereira D. Efforts to support the development of fusidic acid in the United States. *Clin Infect Dis* 2011;52(Suppl. 7):S542–6.
- Tsuji BT, Okusanya OO, Bulitta JB, Forrest A, Bhavnani SM, Fernandez PB, et al. Application of pharmacokinetic–pharmacodynamic modeling and the justification of a novel fusidic acid dosing regimen: raising Lazarus from the dead. *Clin Infect Dis* 2011;52(Suppl. 7):S513–19.
- Turnidge J. Fusidic acid pharmacology, pharmacokinetics and pharmacodynamics. *Int J Antimicrob Agents* 1999;12(Suppl. 2):S23–34.
- Collignon P, Turnidge J. Fusidic acid in vitro activity. *Int J Antimicrob Agents* 1999;12(Suppl. 2):S45–58.
- Spelman D. Fusidic acid in skin and soft tissue infections. *Int J Antimicrob Agents* 1999;12(Suppl. 2):S59–66.
- Atkins B, Gottlieb T. Fusidic acid in bone and joint infections. *Int J Antimicrob Agents* 1999;12(Suppl. 2):S79–93.
- Whitby M. Fusidic acid in the treatment of methicillin-resistant *Staphylococcus aureus*. *Int J Antimicrob Agents* 1999;12(Suppl. 2):S67–71.
- Schöfer H, Simonsen L. Fusidic acid in dermatology: an updated review. *Eur J Dermatol* 2010;20:6–15.
- Australian Prescriber Rifampicin for MRSA. <http://www.australianprescriber.com/magazine/33/5/145/6/> [accessed 02.04.12].
- Gould FK, Brindle R, Chadwick PR, Fraise AP, Hill S, Nathwani D, et al. Guidelines (2008) for the prophylaxis and treatment of methicillin-resistant *Staphylococcus aureus* (MRSA) infections in the United Kingdom. *J Antimicrob Chemother* 2009;63:849–61.
- Darley ES, MacGowan AP. Antibiotic treatment of Gram-positive bone and joint infections. *J Antimicrob Chemother* 2004;53:928–35.
- Zimmerli W, Trampuz A, Ochsner PE. Prosthetic-joint infections. *N Engl J Med* 2004;351:1645–54.
- Trampuz A, Zimmerli W. Prosthetic joint infections: update in diagnosis and treatment. *Swiss Med Wkly* 2005;135:243–51.
- Davis JS. Management of bone and joint infections due to *Staphylococcus aureus*. *Intern Med J* 2005;35(Suppl. 2):S79–96.
- Matthews PC, Berendt AR, McNally MA, Byren I. Diagnosis and management of prosthetic joint infection. *BMJ* 2009;338:b1773.
- Brown NM, Reeves DS, McMullin CM. The pharmacokinetics and protein-binding of fusidic acid in patients with severe renal failure requiring either haemodialysis or continuous ambulatory peritoneal dialysis. *J Antimicrob Chemother* 1997;39:803–9.
- Somekh E, Golan T, Tanay A, Poch F, Dan M. Concentration and bactericidal activity of fusidic acid and cloxacillin in serum and synovial fluid. *J Antimicrob Chemother* 1999;43:593–6.
- Sattar MA, Barrett SP, Cawley MI. Concentrations of some antibiotics in synovial fluid after oral administration, with special reference to anti-staphylococcal activity. *Ann Rheum Dis* 1983;42:67–74.
- Turnidge J, Collignon P. Resistance to fusidic acid. *Int J Antimicrob Agents* 1999;12(Suppl. 2):S35–44.
- Castanheira M, Watters AA, Bell JM, Turnidge JD, Jones RN. Fusidic acid resistance rates and prevalence of resistance mechanisms among *Staphylococcus* spp. isolated in North America and Australia, 2007–2008. *Antimicrob Agents Chemother* 2010;54:3614–7.
- Pfaller MA, Castanheira M, Sader HS, Jones RN. Evaluation of the activity of fusidic acid tested against contemporary Gram-positive clinical isolates from the USA and Canada. *Int J Antimicrob Agents* 2010;35:282–7.
- Foldes M, Munro R, Sorrell TC, Shanker S, Toohey M. In vitro effects of vancomycin, rifampicin, and fusidic acid, alone and in combination, against methicillin-resistant *Staphylococcus aureus*. *J Antimicrob Chemother* 1983;11:21–6.
- Farber BF, Yee YC, Karchmer AW. Interaction between rifampin and fusidic acid against methicillin-resistant coagulase-positive and -negative staphylococci. *Antimicrob Agents Chemother* 1986;30:174–5.
- Zinner SH, Lagast H, Klastersky J. Antistaphylococcal activity of rifampin with other antibiotics. *J Infect Dis* 1981;144:365–71.
- Valardo PE, Debbia E, Schito GC. In vitro activities of rifapentine and rifampin, alone and in combination with six other antibiotics, against methicillin-susceptible and methicillin-resistant staphylococci of different species. *Antimicrob Agents Chemother* 1985;27:615–18.
- Biedenbach DJ, Rhomberg PR, Mendes RE, Jones RN. Spectrum of activity, mutation rates, synergistic interactions, and the effects of pH and serum proteins for fusidic acid (CEM-102). *Diagn Microbiol Infect Dis* 2010;66:301–7.
- Denis O, Nonhoff C, Byl B, Knoop C, Bobin-Dubreux S, Struelens MJ. Emergence of vancomycin-intermediate *Staphylococcus aureus* in a Belgian hospital: microbiological and clinical features. *J Antimicrob Chemother* 2002;50:383–91.
- Balkhair A, Al Muharrmi Z, Darwish L, Farhan H, Sallam M. Treatment of vancomycin-intermediate *Staphylococcus aureus* (VISA) endocarditis with linezolid. *Int J Infect Dis* 2010;14(Suppl. 3):e227–9.
- Falagas ME, Manta KG, Ntziora F, Vardakas KZ. Linezolid for the treatment of patients with endocarditis: a systematic review of the published evidence. *J Antimicrob Chemother* 2006;58:273–80.
- Howden BP, Ward PB, Charles PG, Korman TM, Fuller A, du Cros P, et al. Treatment outcomes for serious infections caused by methicillin-resistant *Staphylococcus aureus* with reduced vancomycin susceptibility. *Clin Infect Dis* 2004;38:521–8.
- Grohs P, Kitzis MD, Gutmann L. In vitro bactericidal activities of linezolid in combination with vancomycin, gentamicin, ciprofloxacin, fusidic acid, and rifampin against *Staphylococcus aureus*. *Antimicrob Agents Chemother* 2003;47:418–20.
- Sagunur R, StDenis M, Ferris W, Aaron SD, Chan F, Lee C, et al. Multiple combination bactericidal testing of staphylococcal biofilms from implant-associated infections. *Antimicrob Agents Chemother* 2006;50:55–61.
- Yu XH, Song XJ, Cai Y, Liang BB, Lin DF, Wang R. In vitro activity of two old antibiotics against clinical isolates of methicillin-resistant *Staphylococcus aureus*. *J Antibiot (Tokyo)* 2010;63:657–9.

- [34] Tang HJ, Chen CC, Cheng KC, Toh HS, Su BA, Chiang SR, et al. In vitro efficacy of fosfomicin-containing regimens against methicillin-resistant *Staphylococcus aureus* in biofilms. *J Antimicrob Chemother* 2012;67:944–50.
- [35] Cox G, Thompson GS, Jenkins HT, Peske F, Savelsbergh A, Rodnina MV, et al. Ribosome clearance by FusB-type proteins mediates resistance to the antibiotic fusidic acid. *Proc Natl Acad Sci USA* 2012;109:2102–7.
- [36] Farrell DJ, Castanheira M, Chopra I. Characterization of global patterns and the genetics of fusidic acid resistance. *Clin Infect Dis* 2011;52(Suppl. 7):S487–92.
- [37] McLaws FB, Larsen AR, Skov RL, Chopra I, O'Neill AJ. Distribution of fusidic acid resistance determinants in methicillin-resistant *Staphylococcus aureus*. *Antimicrob Agents Chemother* 2011;55:1173–6.
- [38] Sule O, Brown NM, Willocks LJ, Day J, Shankar S, Palmer CR, et al. Fusidic acid-resistant *Staphylococcus aureus* (FRSA) carriage in patients with atopic eczema and pattern of prior topical fusidic acid use. *Int J Antimicrob Agents* 2007;30:78–82.
- [39] Mason BW, Howard AJ. Fusidic acid resistance in community isolates of methicillin susceptible *Staphylococcus aureus* and the use of topical fusidic acid: a retrospective case–control study. *Int J Antimicrob Agents* 2004;23:300–3.
- [40] Faber M, Rosdahl VT. Susceptibility to fusidic acid among Danish *Staphylococcus aureus* strains and fusidic acid consumption. *J Antimicrob Chemother* 1990;25(Suppl. B):7–14.
- [41] Castanheira M, Watters AA, Mendes RE, Farrell DJ, Jones RN. Occurrence and molecular characterization of fusidic acid resistance mechanisms among *Staphylococcus* spp. from European countries (2008). *J Antimicrob Chemother* 2010;65:1353–8.
- [42] O'Neill AJ, McLaws F, Kahlmeter G, Henriksen AS, Chopra I. Genetic basis of resistance to fusidic acid in staphylococci. *Antimicrob Agents Chemother* 2007;51:1737–40.
- [43] O'Neill AJ, Larsen AR, Skov R, Henriksen AS, Chopra I. Characterization of the epidemic European fusidic acid-resistant impetigo clone of *Staphylococcus aureus*. *J Clin Microbiol* 2007;45:1505–10.
- [44] O'Neill AJ, Larsen AR, Henriksen AS, Chopra I. A fusidic acid-resistant epidemic strain of *Staphylococcus aureus* carries the *fusB* determinant, whereas *fusA* mutations are prevalent in other resistant isolates. *Antimicrob Agents Chemother* 2004;48:3594–7.
- [45] Laurent F, Tristan A, Croze M, Bes M, Meugnier H, Lina G, et al. Presence of the epidemic European fusidic acid-resistant impetigo clone (EEFIC) of *Staphylococcus aureus* in France. *J Antimicrob Chemother* 2009;63:420–1.
- [46] Katopodis GD, Grivea IN, Tsantsaridou AJ, Pournaras S, Petinaki E, Syrogiannopoulos GA. Fusidic acid and clindamycin resistance in community-associated, methicillin-resistant *Staphylococcus aureus* infections in children of central Greece. *BMC Infect Dis* 2010;10:351.
- [47] Udo EE, Sarkhoo E. Genetic analysis of high-level mupirocin resistance in the ST80 clone of community-associated methicillin-resistant *Staphylococcus aureus*. *J Med Microbiol* 2010;59:193–9.
- [48] Song JH, Hsueh PR, Chung DR, Ko KS, Kang CI, Peck KR, et al. Spread of methicillin-resistant *Staphylococcus aureus* between the community and the hospitals in Asian countries: an ANSORP study. *J Antimicrob Chemother* 2011;66:1061–9.
- [49] Lee K, Lee HS, Jang SJ, Park AJ, Lee MH, Song WK, et al. Antimicrobial resistance surveillance of bacteria in 1999 in Korea with a special reference to resistance of enterococci to vancomycin and Gram-negative bacilli to third generation cephalosporin, imipenem, and fluoroquinolone. *J Korean Med Sci* 2001;16:262–70.
- [50] Kim HB, Jang HC, Nam HJ, Lee YS, Kim BS, Park WB, et al. In vitro activities of 28 antimicrobial agents against *Staphylococcus aureus* isolates from tertiary-care hospitals in Korea: a nationwide survey. *Antimicrob Agents Chemother* 2004;48:1124–7.
- [51] Udo EE, Al-Sweih N, Mokaddas E, Johnny M, Dhar R, Gomaa HH, et al. Antibacterial resistance and their genetic location in MRSA isolated in Kuwait hospitals, 1994–2004. *BMC Infect Dis* 2006;6:168.
- [52] Alreshidi MA, Mariana NS. Increasing rate of detection of fusidic acid resistance in methicillin-resistant *Staphylococcus aureus* isolated from clinical samples in Malaysia. *Med J Malaysia* 2011;66:276.
- [53] Hsu LY, Koh TH, Singh K, Kang ML, Kurup A, Tan BH. Dissemination of multisusceptible methicillin-resistant *Staphylococcus aureus* in Singapore. *J Clin Microbiol* 2005;43:2923–5.
- [54] Lai CC, Wang CY, Chu CC, Tan CK, Lu CL, Lee YL, et al. Correlation between antimicrobial consumption and resistance among *Staphylococcus aureus* and enterococci causing healthcare-associated infections at a university hospital in Taiwan from 2000 to 2009. *Eur J Clin Microbiol Infect Dis* 2011;30:265–71.
- [55] Chen CM, Huang M, Chen HF, Ke SC, Li CR, Wang JH, et al. Fusidic acid resistance among clinical isolates of methicillin-resistant *Staphylococcus aureus* in a Taiwanese hospital. *BMC Microbiol* 2011;11:98.
- [56] Chen HJ, Hung WC, Tseng SP, Tsai JC, Hsueh PR, Teng LJ. Fusidic acid resistance determinants in *Staphylococcus aureus* clinical isolates. *Antimicrob Agents Chemother* 2010;54:4985–91.
- [57] Liu Y, Geng W, Yang Y, Wang C, Zheng Y, Shang Y, et al. Susceptibility and resistance determinants to fusidic acid in *Staphylococcus aureus* isolated from Chinese children with skin and soft tissue infections. *FEMS Immunol Med Microbiol* 2011. <http://dx.doi.org/10.1111/j.1574-695X.2011.00887.x> [Epub ahead of print].
- [58] Ng J, Gosbell IB. Successful oral pristinamycin therapy for osteoarticular infections due to methicillin-resistant *Staphylococcus aureus* (MRSA) and other *Staphylococcus* spp. *J Antimicrob Chemother* 2005;55:1008–12.
- [59] Nather A, David V, Hee HT, Thambiah J. Pyogenic vertebral osteomyelitis: a review of 14 cases. *J Orthop Surg (Hong Kong)* 2005;13:240–4.
- [60] Chiang HL, Chia YY, Chen YS, Hung CC, Liu K, Lo Y. Epidural abscess in an obstetric patient with patient-controlled epidural analgesia—a case report. *Int J Obstet Anesth* 2005;14:242–5.
- [61] Donaldson AD, Chan RC, Gosbell IB. Community-acquired methicillin-resistant *Staphylococcus aureus* in bone and joint infections: development of rifampicin resistance. *Med J Aust* 2006;184:196.
- [62] Inverarity D, Coia J, O'Neill G, MacTier R. MRSA vertebral discitis managed successfully using linezolid as a component of an oral antibiotic regimen. *J R Coll Physicians Edinb* 2006;36:112–14.
- [63] Aboltins CA, Page MA, Buising KL, Jenney AW, Daffy JR, Choong PF, et al. Treatment of staphylococcal prosthetic joint infections with debridement, prosthesis retention and oral rifampicin and fusidic acid. *Clin Microbiol Infect* 2007;13:586–91.
- [64] Apisarnthanarak A, Mundy LM. Successful treatment of disseminated methicillin-resistant *Staphylococcus aureus* with fosfomicin, cefoperazone/sulbactam and rifampin followed by fusidic acid and rifampin. *Int J Infect Dis* 2007;11:283–4.
- [65] Murray RJ, Pearson JC, Coombs GW, Flexman JP, Golledge CL, Speers DJ, et al. Outbreak of invasive methicillin-resistant *Staphylococcus aureus* infection associated with acupuncture and joint injection. *Infect Control Hosp Epidemiol* 2008;29:859–65.
- [66] Ahamed Puthiyaveetil S. Osteomyelitis—a case report. *Aust Fam Physician* 2009;38:521–3.
- [67] Ferry T, Uçkay I, Vaudaux P, François P, Schrenzel J, Harbarth S, et al. Risk factors for treatment failure in orthopedic device-related methicillin-resistant *Staphylococcus aureus* infection. *Eur J Clin Microbiol Infect Dis* 2010;29:171–80.
- [68] O'Neill BJ, Hirpara KM, Kaar TK. Successful treatment of chronic osteomyelitis of the radius. *ISRN Pediatrics* 2011:1–2.
- [69] Chiang ER, Su YP, Chen TH, Chiu FY, Chen WM. Comparison of articulating and static spacers regarding infection with resistant organisms in total knee arthroplasty. *Acta Orthop* 2011;82:460–4.
- [70] Wolfe CR. Case report: treatment of chronic osteomyelitis. *Clin Infect Dis* 2011;52(Suppl. 7):S538–41.