

Oxidative DNA damage estimated by urinary 8-hydroxy-2'-deoxyguanosine and arsenic in glass production workers

Tser-Sheng Lin, Chin-Ching Wu, Jyun-De Wu and Chun-Han Wei
Toxicol Ind Health published online 27 October 2011
DOI: 10.1177/0748233711416945

The online version of this article can be found at:
<http://tih.sagepub.com/content/early/2011/10/18/0748233711416945>

Published by:



<http://www.sagepublications.com>

Additional services and information for *Toxicology and Industrial Health* can be found at:

Email Alerts: <http://tih.sagepub.com/cgi/alerts>

Subscriptions: <http://tih.sagepub.com/subscriptions>

Reprints: <http://www.sagepub.com/journalsReprints.nav>

Permissions: <http://www.sagepub.com/journalsPermissions.nav>

>> **Proof** - Oct 27, 2011

[What is This?](#)

Oxidative DNA damage estimated by urinary 8-hydroxy-2'-deoxyguanosine and arsenic in glass production workers

Tser-Sheng Lin¹, Chin-Ching Wu², Jyun-De Wu³, and Chun-Han Wei¹

Abstract

A total of 130 male glass workers, including 33 administrative workers, 18 batch house workers, 42 craftsmen, and 37 melting process workers, were recruited to investigate the potential DNA damage resulting from toxic element exposure. The occupational exposure to trace elements, including arsenic (As), cadmium (Cd), manganese (Mn), nickel (Ni), lead (Pb), and selenium (Se), was estimated by their urinary levels as internal doses. In addition, all participants filled a self-filled questionnaire indicating their individual information. The average levels of urinary As, Cd, Mn, Ni, Pb, Se, and 8-hydroxy-2'-deoxyguanosine (8-OHdG) were 282.3 ± 464.6 , 3.07 ± 5.39 , 3.81 ± 11.43 , 81.48 ± 138.9 , 18.23 ± 49.61 , 165.2 ± 224.9 , and 17.21 ± 26.34 $\mu\text{g/g}$ creatinine, respectively. The urinary levels of 8-OHdG and toxic elements were strongly associated with the work nature of the worker, with an exception of Mn and Pb. In contrast, the levels of toxic element were not influenced by age, smoking behavior, and alcohol consumption. The urinary 8-OHdG was found significantly higher in higher internal exposure groups of As, Cd, Ni, and Se. However, the stepwise multiple regression models showed that urinary 8-OHdG was only associated with urinary As and heat stress but inversely with age.

Keywords

8-OHdG, oxidative damage, toxic metals, heat stress, glass production

Introduction

Increased risks of stomach, colon, and lung cancers are reported in glassworkers (Cordioli et al., 1987; Dubrow and Wegman, 1983; Hall and Rosenman, 1991; Levin et al., 1988; Lyngne et al., 1986; Malker et al., 1990; Milne et al., 1983; Sankila et al., 1990; Wingren and Axelson, 1985, 1987; Wingren and Englander, 1990). These findings may be as a result of the fact that the glass production workers are potentially exposed to many hazardous chemicals including some carcinogens (Tagesson et al., 1996). A number of trace metals (i.e. arsenic [As], cadmium [Cd], manganese [Mn], nickel [Ni], lead [Pb], selenium [Se], etc.) will be released into the workplace during glass production (Apostoli et al., 1998; Wu et al., 2008).

Toxic metals, including As, Cd, Mn, Ni, and Pb, can induce oxidative damage possibly via a Fenton-like mechanism (Engstrom et al., 2010; Ercal et al.,

2001; Kim et al., 2004; Yamauchi et al., 2004). Urinary 8-hydroxy-2'-deoxyguanosine (8-OHdG) has been widely utilized to estimate the DNA damage caused by oxidative stress and thus used as a biomarker for cancers (Erhola et al., 1997; Honda et al., 2000; Kasai et al., 1984; Lagorio et al., 1994; Pilger

¹Department of Safety, Health, and Environmental Engineering, National United University, Miaoli, Taiwan

²Department of Public Health, China Medical University, Taichung, Taiwan

³Department of Occupational Safety and Health, Chang Jung Christian University, Tainan County, Taiwan

Corresponding author:

Tser-Sheng Lin, Department of Safety, Health, and Environmental Engineering, National United University, 2 Lien Da, Miaoli 360, Taiwan

Email: tslin@nuu.edu.tw

et al., 2000; Tagesson et al., 1996; Toraason et al., 2003).

It is noteworthy that many glass production workers were also exposed to heat stress and the reported Wet Bulb Globe Temperature (WBGT) was as high as 40°C (Srivastava et al., 2000). Animal experiments have well demonstrated that the heat exposure would induce oxidative stress (Bagnyukova et al., 2007; Lin et al., 2006; Lushchak and Bagnyukova, 2006a,b). To our best knowledge, the influence of heat stress on oxidative stress in humans has not been reported yet.

Earlier study cannot find the association between oxidative damage and toxic metal exposure in art glass workers (Tagesson et al., 1996). However, this study did not include the exposure to physical factors such as thermal stress and noise. Thus, this study aimed to investigate the potential oxidative DNA damage owing to the toxic element exposure and physical factors by measuring urinary 8-OHdG in glass production workers. Exposure to trace metals (As, Cd, Mn, Ni, Pb, and Se) was estimated by their urinary levels, while physical stress was defined by similar exposure groups.

Materials and methods

Subject

A total of 130 male workers in 4 different similar exposure groups were randomly recruited in this study. Of these workers, 33 were administrative employees (Group A) without obvious occupational exposure, 18 working for batch house (Group B) were exposed to particulate matter, 42 craftsmen (Group C) were exposed to noise, and others mainly working on melting process (Group M) were exposed to heat stress.

According to Taiwan government regulation, this studied plant has to regularly report the workplace monitoring data measured by a certified laboratory following the approved methods. The last few year reports indicated that the WBGT Index (January) in the melting area ranged from 30.2 to 32.6°C, whereas the average ambient temperature was 15.3°C. The time weighing sound pressure level (SPL-TWA) ranged from 85.0 to 106.6 dB(A) in the craftsman working areas. The particulate matter $\leq 10\mu\text{m}$ (PM₁₀) in the batch house area was below 0.12 mg/m³.

Each participant received a written consent form and agreed to provide his urine sample for 8-OHdG, As, Cd, Ni, Pb, and Se measurements and

a self-filled questionnaire indicating his smoking behavior and alcohol consumption status. The study protocol was approved by our Institute's Human Ethics Committee.

The urine samples were collected in January 2008 and transported back to our laboratory within 2 hours and stored at -20°C until further analysis. In addition, they were required to stop seafood consumption 48 hours earlier than the urine sample collection. The first morning voids were decided to be used for 8-OHdG measurements for the following reasons: (1) it was successfully applied in the chronic environmental exposure to trace metals (Wong et al., 2005); (2) the half-life for the studied trace elements was long and they would accumulate in human body; and (3) a good relationship between urinary 8-OHdG levels in the pre-work and post-work shifts was observed (Wen et al., 2008).

The average age of Group A, Group B, Group C, and Group M workers were 41.3 ± 8.3 , 38.8 ± 10.0 , 38.3 ± 8.7 , and 37.9 ± 8.0 years, respectively. Smoking behavior and alcohol consumption in these four groups were not significantly different (smoker: 48.7–60.0%; alcohol: 20–53.3%).

Trace metal analysis and quality control

A Perkin-Elmer Elan DRCII inductively coupled plasma-mass spectrometer (ICP-MS) was utilized to quantify the concentrations of As, Cd, Ni, Pb, and Se in urine samples. The operation conditions were as follows: (1) carrier gas (argon, 99.999%): 0.8 L/min; (2) plasma gas (argon, 99.999%): 15 L/min; (3) auxiliary gas (argon, 99.999%): 0.9 L/min; (4) pump rate: 1.5 mL/min; and (5) radio frequency (RF) power: 1050 kW. The detection limits determined by three standard deviations of seven measurements of blanks for As, Cd, Mn, Ni, Pb, and Se were 0.22, 0.13, 0.13, 0.16, 0.18, and 0.25 $\mu\text{g/L}$, respectively.

Urine was diluted (1 + 9) with a solution containing 2% nitric acid (trace metal free grade, Sigma, St Louis, MO), 0.2% Triton X-100 (Aldrich, St Louis, MO) and an internal standard of platinum (Pt) for ICP-MS measurements. The reference materials including Seronorm Trace Element Quality (Urine LOT No. 2525) and BIO-RAD Lyphocheck Urine Metals (Urine LOT Level 2-69122) were analyzed to assure our trace metal quantification. The overall recovery was 103.7%, with a standard deviation of 0.03% and the results were detailed in Table 1. The labware clean procedure included soaking in acetone, cleaning with

Table 1. Comparison of measured and certified concentrations, intraday and interday variations

| | Concentration (ng/mL) | | Recovery (%) | Coefficient of Variation (C.V.) [%] | |
|---|-----------------------|-------------|--------------|-------------------------------------|----------|
| | Certified | Measured | | Intraday | Interday |
| Seronom Trace Element Quality Control | | | | | |
| Urine LOT NO2525 | | | | | |
| As | 184 ± 17 | 191 ± 15 | 103.8 | 3.3 | 6.9 |
| Cd | 5.06 ± 0.22 | 4.98 ± 0.13 | 98.4 | 2.8 | 6.7 |
| Mn | 11.1 ± 1.0 | 12.1 ± 1.2 | 109.0 | 4.3 | 6.8 |
| Ni | 41.5 ± 2.2 | 43.5 ± 2.6 | 104.8 | 4.7 | 8.9 |
| Pb | 91.1 ± 7.0 | 88.4 ± 5.2 | 97.0 | 3.1 | 5.2 |
| Se | 66.9 ± 7.1 | 71.2 ± 8.1 | 106.4 | 4.8 | 8.4 |
| BIO-RAD Lyphochek [®] Urine Metals Control | | | | | |
| Urine LOT Level 2-69122 | | | | | |
| As | 144 ± 60 | 147 ± 12 | 102.0 | 4.3 | 7.3 |
| Cd | 12.2 ± 4.5 | 12.8 ± 2.8 | 104.9 | 2.6 | 5.3 |
| Mn | 20.0 ± 4.0 | 22.1 ± 2.2 | 110.5 | 4.2 | 7.5 |
| Ni | 24.5 ± 4.9 | 25.4 ± 2.5 | 103.7 | 3.6 | 4.8 |
| Pb | 59.3 ± 20.2 | 61.2 ± 2.2 | 103.2 | 3.5 | 5.8 |

As: arsenic, Cd: cadmium, Mn: manganese, Ni: nickel, Pb: lead, Se: selenium.

soap, and four-step leaching with different acids (Lin and Nriagu, 1999) to minimize the possible contamination.

8-OHdG and creatinine analysis

Urine samples were centrifuged at 2000g for 10 minutes and then 50 µL of the supernatants was taken for the determination of 8-OHdG levels with a competitive enzyme-linked immunosorbent assay (ELISA) kit (Japan Institute for the Control of Aging, Japan). The determination range was reported to be 0.5–200 ng/mL by the manufacturer. The analysis procedure was briefly described as follows: the 50 µL of 8-OHdG monoclonal antibody and the urine samples were added into the wells of ELISA kit and incubated at 37°C for 1 hour. After washing three times with phosphate-buffered saline (PBS), the antibodies that remained bound to the 8-OHdG in the sample were further bound with the horseradish peroxidase-conjugated secondary antibody for another 1 hour at 37°C. Again after washing for three times with PBS and subsequent addition of 3,3',5,5'-tetramethylbenzidine would allow the development of color intensity proportional to the amount of bound antibodies. The color reaction was then terminated in 15 minutes by stop solution (1 M phosphoric acid) and the absorbance was measured using a spectrophotometric plate reader (VersaMax[™], Kelowna International Scientific Inc., Taipei, Hitachi, Tokyo) at 450 nm wavelength. Urinary creatinine was determined with a Hitachi

7170 autoanalyser using the Jaffé reaction. Finally, urinary 8-OHdG levels were adjusted by urinary creatinine levels.

Statistical analysis

This study used SPSS 12.0 to conduct statistical analysis. The Kolmogorow-Smirnov's test was carried out to examine whether the variables had a normal distribution ($p < 0.05$). Each individual measurement of urinary As, Cd, Mn, Ni, Pb, Se, and 8-OHdG was adjusted with urinary creatinine level and log transformed to stabilize the variance and to approach the normal distribution. We used the analysis of variance (ANOVA) F test to examine the statistical significance of differences in the urinary As, Cd, Mn, Ni, Pb, Se, and 8-OHdG between four groups. The generalized linear model (GLM) was used to examine the association among variables. The stepwise multiple regression models were conducted to test the correlation between variables. The level of statistical significance was $p < 0.05$.

Results

After adjusting with creatinine, the concentrations of urinary As, Cd, Mn, Ni, Pb, Se, and 8-OHdG were 282.6 ± 464.6 , 3.07 ± 5.39 , 3.81 ± 11.43 , 81.48 ± 138.9 , 18.23 ± 49.61 , 165.2 ± 224.9 , and 17.21 ± 26.34 µg/g creatinine, respectively. The results are detailed in Table 2. In addition, they were all logarithmic normal distribution. Thus, further statistical

Table 2. Urinary As, Cd, Mn, Ni, Pb, Se, and 8-OHdG levels ($\mu\text{g/g}$ creatinine; geometric mean) in male glass production workers^a

| | Administrative workers (n = 33) | Batch house workers (n = 18) | Craftsmen (n = 42) | Melting workers (n = 37) | Total workers (n = 130) |
|--------|---------------------------------|------------------------------|---------------------------|---------------------------|---------------------------|
| As | 259.2 \pm 206.3 (174.7) | 106.6 \pm 80.0 (84.7) | 407.9 \pm 694.3 (191.7) | 246.0 \pm 384.6 (136.7) | 282.3 \pm 464.6 (151.9) |
| Cd | 4.43 \pm 8.72 (1.82) | 1.23 \pm 1.16 (0.86) | 3.33 \pm 4.35 (1.91) | 2.45 \pm 3.21 (1.25) | 3.07 \pm 5.39 (1.50) |
| Mn | 3.42 \pm 6.31 (0.94) | 2.87 \pm 2.22 (1.93) | 4.42 \pm 12.49 (0.99) | 3.91 \pm 15.87 (0.57) | 3.81 \pm 11.43 (0.92) |
| Ni | 96.18 \pm 175.9 (40.7) | 47.48 \pm 42.59 (32.5) | 109.2 \pm 176.0 (47.5) | 53.5 \pm 54.49 (29.6) | 81.5 \pm 138.9 (37.9) |
| Pb | 40.53 \pm 86.14 (7.45) | 10.10 \pm 13.99 (4.11) | 15.27 \pm 34.73 (3.65) | 5.67 \pm 9.80 (2.22) | 18.23 \pm 49.61 (3.86) |
| Se | 140.6 \pm 212.9 (48.5) | 36.99 \pm 44.40 (15.58) | 225.5 \pm 251.5 (141.5) | 180.9 \pm 234.0 (78.9) | 165.2 \pm 224.9 (67.2) |
| 8-OHdG | 8.14 \pm 10.71 (3.32) | 11.06 \pm 9.19 (9.19) | 17.28 \pm 18.82 (9.72) | 28.23 \pm 41.48 (12.71) | 17.21 \pm 26.34 (7.80) |

As: arsenic, Cd: cadmium, Mn: manganese, Ni: nickel, Pb: lead, Se: selenium, 8-OHdG: 8-hydroxy-2'-deoxyguanosine.

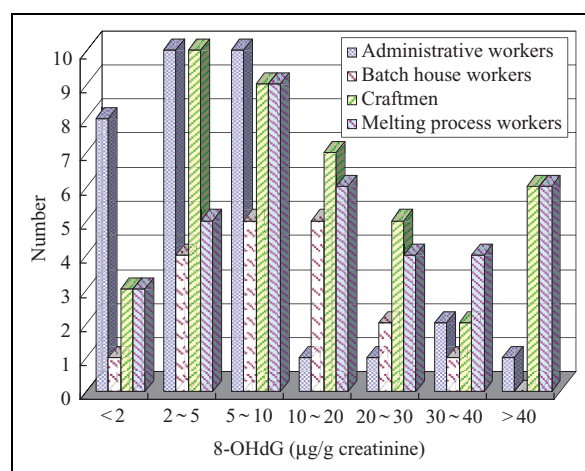
^aAll differences were significant after log transformation among various groups, except for Ni.

analysis was conducted with log-transformed values. The ANOVA indicated that all urinary trace elements were significantly different among these four groups of workers, with an exception of Ni. Most of the lowest urinary trace element values were observed in batch house workers, with an exception of Pb. By contrast, the highest urinary trace element levels were found in the craftsmen, except for Pb. It is noteworthy that the highest urinary Pb level was found in the administrative workers. Moreover, the urinary trace elements were not associated with age, smoking behavior, and alcohol consumption.

The urinary 8-OHdG levels in glass workers of Groups A, B, C, and M were 8.14 ± 10.71 , 11.06 ± 9.19 , 17.28 ± 18.82 , and $28.22 \pm 41.48 \mu\text{g/g}$ creatinine, respectively; and the distribution is shown in Figure 1. It is obvious that the urinary 8-OHdG in Group M workers was much higher than other groups (log transformed, $p < 0.001$).

The influence of As, Cd, Mn, Ni, Pb, and Se exposure on the levels of urinary 8-OHdG was analyzed using urinary trace element levels as internal exposure doses. Thus, the participants were equally divided into three groups, according to their urinary trace element levels. The results are presented in Table 3. As shown, the urinary 8-OHdG was strongly associated with the levels of urinary As, Cd, Ni, and Se. The GLM was further conducted to examine the correlations by controlling age, smoking behavior, and alcohol consumption. All these toxic elements were significantly associated with the urinary 8-OHdG levels, except for Mn and Pb (Table 3). In addition, the urinary 8-OHdG concentration was not affected by smoking and alcohol consumption but inversely associated with age.

The stepwise multiple regression models were utilized to estimate the net effect of a single variable adjusted for other factors. First, the effects of urinary As, Cd, Mn, Ni, Pb, and Se levels, and dust, heat, and

**Figure 1.** The distribution of urinary 8-hydroxy-2'-deoxyguanosine (8-OHdG) in different work groups.

noise exposure on urinary 8-OHdG were therefore analyzed while adjusting the potential confounders including age, smoking behavior, and alcohol consumption. Because we did not measure the personal exposure to dust, noise, and heat stress, these exposures were defined by similar working groups. Exposure to dust, heat, and noise as well as smoking behavior and alcohol consumption were thus considered dummy variables. For heat stress, only group M was set as 1 and others were set as 0. For noise exposure, only group C was set as 1 and others were set as 0. For dust exposure, only group B was set as 1 and others were set as 0. The results (Table 4) clearly indicate that the urinary 8-OHdG level was significantly influenced by heat stress ($p = 0.004$), age ($p = 0.018$), and urinary As ($p = 0.041$).

Discussion

Urinary As, Cd, Mn, Ni, and Pb levels in glass workers were also reported to be higher than those in

Table 3. Urinary 8-OHdG concentrations in different urinary trace element level groups

| Trace element | Low level | Medium level | High level |
|-----------------|---------------|---------------|---------------|
| As ^a | 8.43 ± 8.55 | 16.50 ± 17.62 | 26.73 ± 39.59 |
| Cd ^a | 8.52 ± 8.27 | 18.06 ± 28.48 | 25.04 ± 33.03 |
| Mn | 11.25 ± 10.25 | 22.36 ± 32.40 | 17.91 ± 29.76 |
| Ni ^b | 8.98 ± 7.63 | 15.56 ± 19.06 | 27.14 ± 39.07 |
| Pb | 13.14 ± 18.55 | 17.14 ± 28.61 | 21.35 ± 30.22 |
| Se ^b | 9.85 ± 11.37 | 13.35 ± 16.21 | 28.53 ± 39.12 |

As: arsenic, Cd: cadmium, Mn: manganese, Ni: nickel, Pb: lead, Se: selenium, 8-OHdG: 8-hydroxy-2'-deoxyguanosine.

^a $p < 0.05$.

^b $p < 0.01$.

Table 4. Results of stepwise multiple regression of urinary As, Cd, Mn, Ni, Pb, Se levels, and heat exposure and confounder with respect to urinary 8-OHdG levels^a

| Model | Unstandardized coefficient (B) | Standardized coefficient | p Value |
|---------------|--------------------------------|--------------------------|---------|
| 1 Constant | 0.741 | | 0.000 |
| Heat exposure | 0.400 | 0.286 | 0.004 |
| 2 Constant | 1.499 | | 0.000 |
| Heat exposure | 0.391 | 0.279 | 0.004 |
| Age | -0.019 | -0.237 | 0.014 |
| 3 Constant | 0.801 | | 0.082 |
| Heat exposure | 0.409 | 0.293 | 0.002 |
| Age | -0.018 | -0.224 | 0.018 |
| As | 0.296 | 0.194 | 0.041 |

^alog ($\times \mu\text{g/g creatinine}$).

control subjects (Apostoli et al., 1998). Compared with their values, our values for As and Ni were much higher although Cd, Mn, and Pb were lower. The disparity may be due to the different composite of the raw materials used. Compared with occupationally nonexposed population (Batariova et al., 2006; Heitland and Koster, 2006; Paschal et al., 1998), our values were certainly higher. For example, the urinary As, Cd, Mn, Ni, Pb, and Se for occupationally nonexposed German (Heitland and Koster, 2006) were 34, 0.18, 0.087, 0.756, 0.8, and 14 $\mu\text{g/L}$, respectively. The urinary As in healthy Japanese was reported to be $149 \pm 125 \mu\text{g/g creatinine}$ (Yamauchi et al., 2004). Obviously, the urinary As level in the glass workers was much higher and posed a health threat. The lowest values observed in the batch house workers might be because their workplace was separate and far from the production buildings, while the administrative offices were adjacent to the production areas and receiving traffic-related emissions. In addition, the batch house workers were legislatively required to wear suitable personal protection equipment. Thus, the suggested source of As, Mn, Ni, and Se was the fugitive emission from melting processes. The craftsmen also worked closer to the fugitive

sources than the workers involved in the melting process.

The urinary 8-OHdG levels reported for occupationally nonexposed population ranged from 1.84 to 15.7 $\mu\text{g/g creatinine}$ (Engstrom et al., 2010; Lee et al., 2010; Liu et al., 2009; Lu et al., 2007; Rossner et al., 2008; Tagesson et al., 1996; Yamauchi et al., 2004), whereas those with occupational exposure ranged from 5.0 to 61.4 $\mu\text{g/g creatinine}$ (Kim et al., 2004; Lee et al., 2010; Liu et al., 2009; Lu et al., 2007; Rossner et al., 2008; Tagesson et al., 1996; Wen et al., 2008). Our measurements for administrative workers were in the range of $7.9 \pm 10.5 \mu\text{g/g creatinine}$. The urinary 8-OHdG levels in exposed workers (12.44 for craftsmen and 28.22 for melting process workers) are obviously higher. However, the urinary 8-OHdG measurements for the same population at different periods have shown a considerable variation (Rossner et al., 2008); thus the comparison of urinary 8-OHdG reported in different studies is difficult.

Oxidative damage owing to reactive oxygen species (ROS) and radical-related damage to DNA have been suggested to play an important role in cancers,

arteriosclerosis, arthritis, and so on (Valko et al., 2006). Urinary 8-OHdG has been widely used as a biomarker of oxidative damage after it was identified in human urine (Valko et al., 2006). Our multiple regression models indicated the urinary 8-OHdG level was not associated with tobacco and alcohol consumptions which agree with the observations by Nia et al. (2001), Lodovici et al. (2000), Liu et al. (2009), and Van Zeeland et al. (1999). In addition, an earlier study on art glass workers (Tagesson et al., 1996) also demonstrated that the urinary 8-OHdG level was not statistically associated with smoking behavior in male workers. In contrast, the urinary 8-OHdG was significantly elevated in female smokers. In addition, the elevation of urinary 8-OHdG level owing to smoking behavior was also reported by Kasai et al. (2001). It seems the urinary 8-OHdG would be elevated by tobacco consumption, but occupational exposure will be more significant in inducing oxidative damage. Very few studies reported the relationship between urinary 8-OHdG and alcohol consumption. Liu et al. (2009) reported that the urinary 8-OHdG was not significantly affected by alcohol consumption. The urinary 8-OHdG level was observed to decrease due to aging both in male rats and in humans (Fraga et al., 1990; Kasai et al., 2001). Our regression models also demonstrate this tendency.

Many studies have found a considerable increase in urinary 8-oxo-7, 8-dihydro-2'-deoxyguanine (8-OHdG) owing to occupational exposure to chemicals such as Mn, Ni, and Pb (Kim et al., 2004). In addition, the urinary 8-OxodG level was strongly associated with urinary As and Cd but not urinary Pb in early pregnancy women (Engstrom et al., 2010). This study also showed a higher 8-OHdG level in workers with higher urinary As, Cd, Ni, and Se levels (smoking behavior, alcohol consumption, and age). An increase in 8-OHdG excretion was also demonstrated both in individuals with acute and chronic As exposure (Yamauchi et al., 2004). Our stepwise regression models also showed the association between urinary 8-OHdG and As levels.

A Fenton-type reduction has been suggested as an important mechanism for the oxidative damage produced by metals (Moriwaki et al., 2008; Soares et al., 2008). Hence, the chemical species of a metal may significantly influence its oxidative stress. For example, metavanadate would result in more severe oxidative damage than decavanadate (Soares et al., 2008). In addition, the accumulation of metals may also maintain its oxidative stress on biota (Soares

et al., 2008). However, the interaction between metal ions included suppressive, additive, and synergistic effects on DNA oxidative stress (Moriwaki et al., 2008).

Animal experiments have demonstrated that the heat exposure will result in oxidative damage (Bagnyukova et al., 2007; Lin et al., 2006; Lushchak and Bagnyukova, 2006a,b). For example, thiobarbituric acid reacting substances (TBARS) would be significantly increased in 5-week broil chickens after a 6-hour exposure to 32°C (Lin et al., 2006) which indicated that the oxidative stress was induced. A similar phenomenon was also observed in the rotan (Bagnyukova et al., 2007). For example, the TBARS would be elevated by 2–3 folds after the exposure to 32°C for 12 hours; moreover, the activity of superoxide dismutase (SOD) would increase 3 folds. The experiments on goldfish found that the SOD activities would increase with heat exposure (35°C) but was reversible with a lower temperature (19°C) recovery (Lushchak and Bagnyukova, 2006b). To our best knowledge, no association between heat stress and oxidative stress in human has been documented. Our results indicated that the exposure to heat may pose strong oxidative stress resulting in severe DNA damage. Animal studies also indicated that the heat stress might also interfere with the metabolic processes; however, we did not measure blood As, Cd, Mn, Ni, Pb, and Se levels as well as their ambient levels. Thus, the interference of heat stress on workers cannot be drawn and need further studies.

The oxidative damage of cochlear hair cells due to noise was reported and can be prevented by antioxidants (Rabinowitz et al., 2002). Our study did not find the association between urinary 8-OHdG level and noise hearing. The possible explanations are the following. First, all craftsmen were required to wear protective equipment against noise. Second, we did not measure personal exposure, thus the analysis with dummy variable cannot reflect the true relationship.

In addition, we did not collect the urine samples of both pre-work and post-work shifts, for analysis; so it may be of a concern that the influence of heat exposure on oxidative stress in human cannot be directly examined since the urinary 8-OHdG may be reversed after work. The studies of heat stress exposure in goldfish showed that the oxidative damage markers had been increasing for 24 hours after heat exposure (Lushchak and Bagnyukova, 2006a). The results of single injection of Cd to marine teleost showed

that the oxidative stress markers would maintain at a significantly higher levels at least for 7 days (Soares et al., 2008). In addition, the oxidative damage has been increasing at least for 72 hours after intense noise exposure in rats (Van Campen et al., 2002). For human, the urinary 8-OHdG may need 180 days to be recovered after a single strong exposure (Yamauchi et al., 2004). In spite of the rapidly decreasing urinary As level after acute As poisoning, the urinary 8-OHdG level peaked at 30 days after that acute exposure (Yamauchi et al., 2004). For the oxidative damage regarding chronic exposure, the urinary 8-OHdG in first morning voids was not statistically different from the average value in the 24-hour average urinary levels (Thompson et al., 1999; Wong et al., 2005). The urinary 8-OHdG levels in the before and end work samples were significantly correlated (Kim et al., 2004; Wen et al., 2008). Although the urinary 8-OHdG level after work may be higher than that before work, Kim's study (2004) also showed that the first day urinary 8-OHdG level was higher than those after work on days 2, 4, and 5. These phenomena reveal that the intensity of hazard exposure might significantly influence the recovery from oxidative damage. Assuming that a worse recovery from oxidative damage due to chronic exposure may pose a higher health risk, the urinary 8-OHdG level in the first morning voids may represent oxidative damage due to chronic exposure hazards.

In conclusion, this investigation points out that higher internal dose of As, Cd, Ni, and Se may result in significant oxidative stress; however, heat stress may play an important role on oxidative damage. Thus warrants further studies to detail the interference of heat stress on oxidative damage.

Acknowledgements

The authors greatly appreciated all the participants for their help during the urine sample collection and questionnaire filling.

Funding

The project was partly supported by the Taiwan National Science Council (NSC97-2221-E-239-017).

References

- Apostoli P, Giusti S, Bartoli D, Perico A, Bavazzano P, and Alessio L (1998) Multiple exposure to arsenic, antimony, and other element in art glass manufacturing. *American Journal of Industrial Medicine* 34(1): 65–72.
- Bagnyukova TV, Danyliv SI, Zinko OS, and Lushchak VI (2007) Heat shock induces oxidative stress in rotan *Perccottus glenii* tissues. *Journal of Thermal Biology* 32: 255–260.
- Batariova A, Spevackova V, Benes B, Cejchanova M, Smid J, and Cerna M (2006) Blood and urine levels of Pb, Cd and Hg in the general population of the Czech Republic and proposed reference values. *International Journal of Hygiene and Environmental Health* 209(4): 359–366.
- Cordioli G, Cuoghi L, Solari L, Berrino F, Crosignani P, and Riboli E (1987) Mortalita per tumore in una coorte di lavoratori della industria de vetro. *Epidemiologia e Prevenzione* 9(30): 16–18.
- Dubrow R, Wegman DH (1983) *Occupational Characteristics of Cancer Victims in Massachusetts: 1971-1973*. DHHS (NIOSH) Publ. no. 84 109. Cincinnati, OH: National Institute for Occupational, Safety and Health.
- Engstrom KS, Vahter M, Johansson G, Lindh CH, Teichert F, Singh R, et al. (2010) Chronic exposure to cadmium and arsenic strongly influences concentrations of 8-oxo-7,8-dihydro-2'-deoxyguanosine in urine. *Free Radical Biology and Medicine* 48(9): 1211–1217.
- Ercal N, Gurer-Orhan H, and Aykin-Burns N (2001) Toxic metals and oxidative stress part I: mechanisms involved in metal induced oxidative damage. *Current Topics in Medicinal Chemistry* 1(6): 529–539.
- Erhola M, Toyokuni S, Okada K, Tanaka T, Hiai H, Ochi H, et al. (1997) Biomarker evidence of DNA oxidation in lung cancer patients: association of urinary 8-hydroxy-2'-deoxyguanosine excretion with radiotherapy, chemotherapy, and response to treatment. *FEBS Letters* 409(2): 287–291.
- Fraga CG, Shigenaga MK, Park JW, Degan P, and Ames BN (1990) Oxidative damage to DNA during aging: 8-hydroxy-2'-deoxyguanosine in rat organ DNA and Urine. *Proceedings of the National Academy of Sciences of the United States of America* 87(12): 4533–4537.
- Hall NEL, Rosenman KD (1991) Cancer by industry: analysis of a population-based cancer registry with an emphasis on blue collar workers. *American Journal of Industrial Medicine* 19(2): 145–159.
- Heitland P, Koster HD (2006) Biomonitoring of 30 trace elements in urine of children and adults by ICP-MS. *Clinica Chimica Acta* 365(1–2): 310–318.
- Honda M, Yamada Y, Tomonaga M, Ichinose H, and Kamihira S (2000) Correlation of urinary 8-hydroxy-2'-deoxyguanosine (8-OHdG), a biomarker of oxidative DNA damage, and clinical features of hematological disorders: a pilot study. *Leukemia Research* 24(6): 461–468.
- Kasai H, Hayami H, Yamaizumi Z, Saito H, and Nishimura S (1984) Detection and identification of mutagens and carcinogens as their adducts with guanosine derivatives. *Nucleic Acids Research* 12(4): 2127–2136.

- Kasai H, Iwamoto-Tanaka N, Miyamoto T, Kawanami K, Kawanami S, Kido R, et al. (2001) Life style and urinary 8-hydroxydeoxyguanosine, a marker of oxidative DNA damage: effects of exercise, working conditions, meat intake, body mass index, and smoking. *Japanese Journal of Cancer Research* 92(1): 9–15.
- Kim JY, Mukherjee S, Ngo L, and Christiani DC (2004) Urinary 8-hydroxy-2'-deoxyguanosine as a biomarker of oxidative DNA damage in workers exposed to fine particulates. *Environmental Health Perspectives* 112(6): 666–671.
- Lagorio S, Tagesson C, Forastiere F, Iavarone I, Axelson O, and Carere A (1994) Exposure to benzene and urinary concentrations of 8-hydroxydeoxyguanosine, a biological marker of oxidative damage to DNA. *Occupational and Environmental Medicine* 51(11): 739–743.
- Lee M, Chen M, Lung S, Tsai C, Yin X, and Mao I (2010) Exposure assessment of PM_{2.5} and urinary 8-OHdG for diesel exhaust emission inspector. *Science of the Total Environment* 408(3): 505–510.
- Levin LI, Zheng W, Blot WJ, Gao Y-T, and Fraumeni JF Jr (1988) Occupation and lung cancer in Shanghai: a case-control study. *British Journal of Industrial Medicine* 45(7): 450–458.
- Lin H, Decuyper E, and Buyse J (2006) Acute heat stress induces oxidative stress in broiler chickens. *Comparative Biochemistry and Physiology - Part A: Molecular and Integrative Physiology* 144(1): 11–17.
- Lin T, Nriagu J (1999) Thallium speciation in river waters with Chelex-100 resin. *Analytica Chimica Acta* 395: 301–307.
- Liu H, Lin M, Liu P, Chan C, and Chen H (2009) Health risk assessment by measuring plasma malondialdehyde (MDA), urinary 8-hydroxydeoxyguanosine (8-OH-dG) and DNA strand breakage following metal exposure in foundry workers. *Journal of Hazardous Materials* 170(2–3): 699–704.
- Lodovici M, Casalini C, Cariaggi R, Michelucci L, and Dolara P (2000) Levels of 8-hydroxydeoxyguanosine as a marker of DNA damage in human leukocytes. *Free Radical Biology and Medicine* 28(1): 13–17.
- Lu C, Ma Y, Lin J, Chuang C, and Sung F (2007) Oxidative DNA damage estimated by urinary 8-hydroxydeoxyguanosine and indoor air pollution among non-smoking office employees. *Environmental Research* 103(3): 331–337.
- Lushchak VI, Bagnyukova TV (2006a) Temperature increase results in oxidative stress in goldfish tissues. 1. Indices of oxidative stress. *Comparative Biochemistry and Physiology - Part C: Toxicology and Pharmacology* 143(1): 30–35.
- Lushchak VI, Bagnyukova TV (2006b) Temperature increase results in oxidative stress in goldfish tissues. 2. Antioxidant and associated enzymes. *Comparative Biochemistry and Physiology - Part C: Toxicology and Pharmacology* 143(1): 36–41.
- Lynge L, Kurppa K, Kristofersen L, Malker H, and Sauli H (1986) Silica dust and lung cancer: results from the Nordic occupational mortality and cancer incidence registers. *Journal of the National Cancer Institute* 77(4): 883–889.
- Malker HSR, McLaughlin JK, Weiner JA, Silverman DT, Blot WJ, Ericsson JLE, et al. (1990) Occupational risk factors for nasopharyngeal cancer in Sweden. *British Journal of Industrial Medicine* 47(3): 213–214.
- Milne R, Sandler DP, Everson RB, and Brown SM (1983) Lung cancer and occupation in Alameda county: a death certificate case control study. *American Journal of Industrial Medicine* 4(4): 565–575.
- Moriwaki H, Osborne MR, and Phillips DH (2008) Effects of mixing metal ions on oxidative DNA damage mediated by a Fenton-type reduction. *Toxicology in Vitro* 22(1): 36–44.
- Nia AB, Van Schooten FJ, Schilderman PAEL, De Kok TCM, Haenen GR, Van Herwijnen MHM, et al. (2001) A multi-biomarker approach to study the effects of smoking on oxidative DNA damage and repair and antioxidative defense mechanisms. *Carcinogenesis* 22(3): 395–401.
- Paschal DC, Ting BG, Morrow JC, Pirkle JL, Jackson RJ, Sampson EJ, et al. (1998) Trace metals in urine of United States residents: reference range concentration. *Environmental Research* 76(1): 53–59.
- Pilger A, Germadnik D, Schaffer A, Theiler A, Pils P, Sluka F, et al. (2000) 8-Hydroxydeoxyguanosine in leukocyte DNA and urine of quartz-exposed workers and patients with silicosis. *International Archives of Occupational and Environmental Health* 73(5): 305–310.
- Rabinowitz PM, Wise JP Sr, Moberg BH, Antonucci PG, Powell C, and Slade M (2002) Antioxidant status and hearing function in noise-exposed workers. *Hearing Research* 173(1–2): 164–171.
- Rossner P Jr, Svecova V, Milcova A, Lnenickova Z, Solansky I, and Sram RJ (2008) Seasonal variability of oxidative stress markers in city bus drivers. Part I. Oxidative damage to DNA. *Mutation Research* 642(1–2): 14–20.
- Sankila R, Karjalainen S, Pukkala E, Oksanen H, Hakulinen T, Teppo L, et al. (1990) Cancer risk among glass factory workers: an excess of lung cancer? *British Journal of Industrial Medicine* 47(12): 815–818.
- Soares SS, Martins H, Gutierrez-Merino C, and Aureliano M (2008) Vanadium and cadmium *in vivo* effects in

- teleost cardiac muscle: metal accumulation and oxidative stress markers. *Comparative Biochemistry and Physiology - Part C: Toxicology and Pharmacology* 147(2): 168–178.
- Srivastava A, Kumar R, Joseph E, and Kumar A (2000) Heat exposure study in the workplace in a glass manufacturing unit in India. *The Annals of Occupational Hygiene* 44(6): 449–453.
- Tagesson C, Magnus K, and Wingren G (1996) Urinary malondialdehyde and 8-hydroxydeoxyguanosine as potential markers of oxidative stress in industrial art glass workers. *International Archives of Occupational and Environmental Health* 69(1): 5–13.
- Thompson HJ, Heimendinger J, Haegle A, Sedlacek SM, Gillette C, O'Neill C, et al. (1999) Effect of increased vegetable and fruit consumption on markers of oxidative cellular damage. *Carcinogenesis* 20(12): 2261–2266.
- Toraason M, Butler MA, Ruder A, Forrester C, Taylor L, Ashley DL, et al. (2003) Effect of perchloroethylene, smoking, and race on oxidative DNA damage in female dry cleaners. *Mutation Research* 539(1–2): 9–18.
- Van Campen LE, Murphy WJ, Franjs JR, Mathias PI, and Toraason MA (2002) Oxidative DNA damage is associated with intense noise exposure on the rat. *Hearing Research* 164(1–2): 29–38.
- Valko M, Thodes CJ, Moncol J, Izakovic M, and Mazur M (2006) Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chemico-Biological Interactions* 160(1): 1–40.
- Van Zeeland AA, de Groot AJL, Hall J, and Donato F (1999) 8-Hydroxydeoxyguanosine in DNA from leukocytes of healthy adults: relationship with cigarette smoking, environmental tobacco smoke, alcohol and coffee consumption. *Mutation Research* 439(2): 249–257.
- Wen S, Yang F, Gong Y, Zhang X, Hui Y, Li J, et al. (2008) Elevated levels of urinary 8-hydroxy-2'-deoxyguanosine in male electrical and electronic equipment dismantling workers exposed to high concentrations of polychlorinated dibenzo-p-dioxins and dibenzofurans, polybrominated diphenyl ethers, and polychlorinated biphenyls. *Environmental Science and Technology* 42(11): 4202–4207.
- Wingren G, Axelson O (1985) Mortality pattern in a glass producing area in SE Sweden. *British Journal of Industrial Medicine* 42(6): 411–414.
- Wingren G, Axelson O (1987) Mortality in the Swedish glassworks industry. *Scandinavian Journal of Work Environment and Health* 13(5): 412–416.
- Wingren G, Englander V (1990) Mortality and cancer morbidity in a cohort of Swedish glassworkers. *International Archives of Occupational and Environmental Health* 62(3): 253–257.
- Wong R, Kuo C, Hsu M, Wang T, Chang P, Wu T, et al. (2005) Increased levels of 8-hydroxy-2'-deoxyguanosine attributable to carcinogenic metal exposure among schoolchildren. *Environmental Health Perspectives* 113(10): 1386–1390.
- Yamauchi H, Aminaka Y, Yoshida K, Sun G, Pi J, and Waalkes MP (2004) Evaluation of DNA damage in patients with arsenic poisoning: urinary 8-hydroxydeoxyguanine. *Toxicology and Applied Pharmacology* 198(3): 291–296.