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Abstract	<p>Radiotherapy widely uses the polymer gel dosimeter. The advantage of polymer gel dosimetry is the mapped 3D absorbed dose distribution that other dosimeters cannot achieve. The Acrylamide (AAm) is a frequently used monomer; however, the extreme toxicity of Acrylamide (ORL-RAT LD50: 124 mg/kg) raises a concern. Therefore, this study developed a new type of Propylene acid based gel dosimeter, named DEMBIG gel. The following outlines the aim of this study: (1) using two-point formulation to find the optimal scan parameter of MRI according to the best sensitivity and linearity (correlation coefficient) of DEMBIG gel, (2) using the optimal scan parameter of MRI to observe the properties of DEMBIG gel, and (3) verifying the three-dimensional (3D) dose distributions of radiotherapy. This study obtained three major results: 1. The scan protocol of MRI was established. 2. The preliminary results of DEMBIG gel were: (1) The range of absorbed dose of DEMBIG gel: 0–20 Gy. (2) The sensitivity and correlation coefficient of DEMBIG gel at verification as slope: 0.181 sGy^{-1}, $R^2:0.997$. (3) There is no energy dependency of the DEMBIG gel. (4) The dose difference was 3% in the three-dimensional (3D) isocenter dose in clinical radiotherapy. These data show that DEMBIG gel is a potential candidate for the 3D dosimeter.</p>	
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Preliminary investigation of a new type of propylene based gel dosimeter (DEMBIG)

Bor-Tsung Hsieh · Chi-Tsung Chiang ·
Pi-Hui Hung · Chia-Hung Kao · Ji-An Liang

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Abstract Radiotherapy widely uses the polymer gel dosimeter. The advantage of polymer gel dosimetry is the mapped 3D absorbed dose distribution that other dosimeters cannot achieve. The Acrylamide (AAM) is a frequently used monomer; however, the extreme toxicity of Acrylamide (ORL-RAT LD50: 124 mg/kg) raises a concern. Therefore, this study developed a new type of Propylene acid based gel dosimeter, named DEMBIG gel. The following outlines the aim of this study: (1) using two-point formulation to find the optimal scan parameter of MRI according to the best sensitivity and linearity (correlation coefficient) of DEMBIG gel, (2) using the optimal scan parameter of MRI to observe the properties of DEMBIG gel, and (3) verifying the three-dimensional (3D) dose

distributions of radiotherapy. This study obtained three major results: 1. The scan protocol of MRI was established. 2. The preliminary results of DEMBIG gel were: (1) The range of absorbed dose of DEMBIG gel: 0–20 Gy. (2) The sensitivity and correlation coefficient of DEMBIG gel at verification as slope: 0.181 sGy^{-1} , $R^2:0.997$. (3) There is no energy dependency of the DEMBIG gel. (4) The dose difference was 3% in the three-dimensional (3D) isocenter dose in clinical radiotherapy. These data show that DEMBIG gel is a potential candidate for the 3D dosimeter.

Keywords Polymer gel dosimeter · 3D dosimeter

Introduction

Radiation therapy can induce many significant biological and chemical effects to tumor cells and surrounding normal tissue. Verifying radiation treatment planning is important to not only deliver an adequate prescribed dose to the target volume, but to also spare critical organs at risk, especially for application in radiosurgery.

Many systems of chemical dosimetry have been proposed with great achievement in the recent years, such as Fricke dosimeter [1], film, solid state methods, silicon diodes and other aqueous dosimeter [2, 3]. We may select the proper dosimeter based on their unique character.

Wagter [4] proposed the terms of an ideal dosimeter in 2004. Traditionally, the dosimeter used for verification in radiation oncology can only show a 2D dose map.

Polymer gel, a chemical dosimeter that reacts with the monomer and free radicals at the irradiated area, has the unique advantage to offer 3D dose distribution. The degree of polymerization is proportional to radiation dose. Alexander et al. [5] first proposed the polymer system to

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55 determine the effects of ionizing radiation on polymeth-
56 ylmethacrylate. Maryanski [6] first proposed the PAG gel
57 with acrylic monomers and *N,N'*-methylenebisacrylamide,
58 named BANANA in 1993.

59 Recently, we used the less toxic material (DEMA,
60 2-(Dimethylamino) ethyl acrylate, ORL-RAT LD50:1751
61 mg/kg [7]) as a monomer in composition with the new
62 polymer gel named DEMBIG (2-(Dimethylamino) ethyl
63 acrylate, *N,N'*-methylene-bisacrylamide, Gelatin).

64 This research used MRI as a measurement method of
65 DEMBIG gel to obtain an optimal scan parameter to verify
66 the correlation coefficient of dose response, temporal sta-
67 bility of dose response, and energy dependence for DEM-
68 BIG gel by optimal MRI parameters. This study also used
69 DEMBIG gel and optimal MRI parameters to simulate the
70 radiotherapy process and to verify the isocenter dose.

71 Materials and methods

72 Polymer gel manufacture

73 The composition of DEMA gel were Gelatin (Sigma-
74 Aldrich 7%), 2-(Dimethylamino) ethyl methacrylate (DEMA,
75 Sigma-Aldrich 5%), *N,N'*-methylene-bisacrylamide (BIS,
76 Merck Chemical Company 4%), and deionized water
77 prepared under a controlled argon atmosphere inside a
78 glove box. To begin gel manufacture, the water was filled
79 with argon (20 psi/min) for 20 min, followed by adding
80 gelatin to the water and magnetically stirring for 10 min at
81 room temperature. The solution was further heated and
82 stirred to 45 °C to dissolve the gelatin. After 15 min,
83 DEMA and BIS were added to the solution and kept
84 magnetically stirred for 30 min until complete dissolution.
85 Finally, the gel was filled with argon for 30 min and then
86 poured into Pyrex screw test tubes (16 mm OD, 100 mm
87 length, No. 9826,) and wrapped in aluminum foil to pre-
88 vent photo-polymerization, and placed in a refrigerator
89 (4 ± 1 °C) for 48 h to irradiate.

90 Irradiation of the gel

91 Irradiation was performed by a linear accelerator (Clinac
92 21 EX, Varian Medical Systems, Palo Alto, CA, USA).
93 The center of the short side of a customized 30 cm ×
94 30 cm × 4 cm acrylic phantom was punctured with a
95 16 mm diameter hole, to accommodate the Pyrex test tube.
96 To discover the precise location, 3.5 cm and 16.5 cm
97 acrylic sticks were placed in the upper and lower sides of a
98 test tube. Acrylic phantom was placed in the middle
99 between two pieces of 3 cm solid water phantoms. The
100 setup criteria of the linear accelerator were: gantry: 0°, field
101 size: 10 × 10 cm², depth: 5 cm, photon energy: 6MV.

Magnetic resonance imaging

103 The T2 relaxation times of gel samples were determined
104 24 h after irradiation, using a head coil in the MRI facility
105 (Signa 0.5 T, GE Medical System). The vials were imaged
106 upright in a single slice in the axial plane at 22 °C using a
107 2-echo spin-echo sequence. Dose–response curves were
108 calculated by taking the mean (and standard deviation) of a
109 region of interest within each vial. All calculations were
110 performed using Image J (free software). This study used
111 fast spin-echo sequences to acquire optimal echo time, the
112 formula of *R2* calculation as formula (1)

$$R2 = \frac{1}{T2} = \frac{1}{TB2 - TB2} \times \frac{S(TB2)}{S(TB2)} \quad (1)$$

114 The two acquired images matched the long echo time and
115 the short echo time. The DEMBIG gel was imaged by the
116 spine echo sequence and the fast spine echo sequence in 14
117 protocols. The spine echo sequence set of TE matches
118 using TE1 are as follows: 30, 40 ms and TE2: 100, 120,
119 130, 140, 150, 160, 170, 180 ms with TR of 3 s, slice
120 thickness of 5 mm, FOV of 256 mm. The fast spine echo
121 sequence set of TE matches were TE1:31.5 ms and
122 TE2:158 ms, the other parameter was the same as the
123 spine echo sequence. Table 1 lists the parameter setting of
124 MRI.

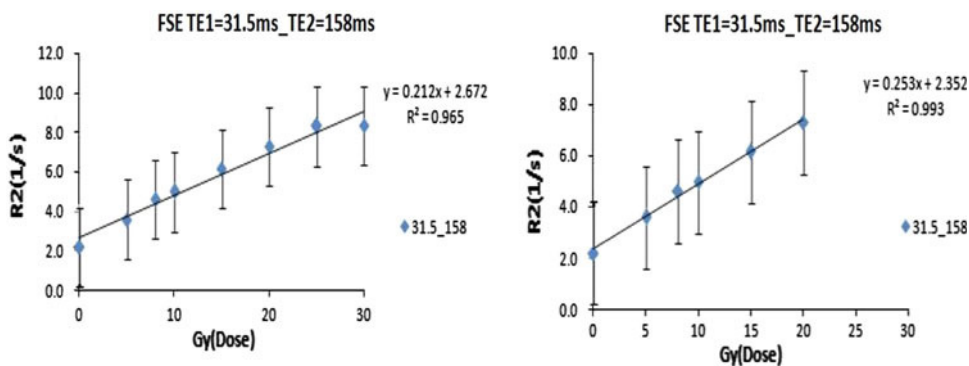
Verification of dose distribution in radiotherapy

125 As a base for treatment planning, CT-images of the gel
126 phantom were acquired using a spiral CT scanner (Hispeed
127 NX/I, GE Medical System). The slice thickness was 2 mm.
128 We used the treatment planning system (Eclipse, Varian
129 Medical Systems, Palo Alto, CA, USA) to generate the 6
130 MV radiosurgery plan, with gantry angle rotation 0, 72,
131 144, 216, 288 degree and the prescribed target dose of
132 20 Gy. The output was 400 MU/min during radiation.
133 A 270 mL gel phantom was irradiated by a linear accel-
134 erator. There were six vials for the calibration curve. 135

Table 1 Scan parameter setting of MRI

Parameter	
Matrix size (MS)	256 × 256
Slice Thickness (mm)	5
Repetition Time (ms)	3000
Echo Time 1_short TE1 (ms)	31.5, 30, 40
Echo Time 2_long TE2 (ms)	100, 120, 130, 140, 150, 158, 160, 170, 180
Number of slices	9
Number of echo	2
Bandwidth (Hz/pixel)	10.42

Fig. 1 Dose response curve of DEMBIG gel at optimal TE match protocol



136 **Results**

137 Optimal scan parameter-Echo Time, TE

138 DEMBIG acquired the image of optimal echo times by a
 139 fast spin-echo sequence, analyzing the relationship
 140 between R_2 and the absorbed dose by a two-point method
 141 at different ranges of dose response. The R_2 -dose response
 142 of the DEMBIG polymer gel dosimeter was linear between
 143 0 and 30 Gy doses. Figure 1 shows the dose response curve
 144 of DEMBIG gel at optimal TE match protocol. The fast
 145 spine-echo sequence set of the TE1:31.5 ms and TE2:158
 146 acquired superior R_2 -dose sensitivities and correlation
 147 coefficient. We found an improved correlation coefficient
 148 at 0–20 Gy compared to 0–30 Gy, when the absorbed dose
 149 over 25 Gy the DEMBIG was saturated.

150 Verification optimal protocol of MRI

151 *Correlation coefficient of dose response*

152 The DEMBIG polymer gel formulation by % mass con-
 153 sisted of 4% N,N' -methylene-bis-acrylamide (bis), 5%
 154 DEMA, and 7% gelatin irradiated up to 20 Gy and imaged
 155 by MRI optimal protocol. Table 2 lists the optimal setting.
 156 Figure 2 shows the DEMBIG gel for three batches using
 157 the optimal protocol.

158 *Temporal stability of DEMBIG gel*

159 The DEMBIG gel was irradiated up to 30 Gy and imaged
 160 at 2, 4, 6, 12, 18, 24, 48, and 72 h respectively. Figure 3
 161 shows the correlation coefficient of 0–30, 0–25, and
 162 0–20 Gy, including the temporal stability of DEMBIG gel
 163 at 24 h in 0–20 Gy after post irradiation.

Table 2 Optimal setting of MRI

TE1 (ms)	TE2 (ms)	TR (ms)	Thickness (mm)	Scan sequence
31.5	158	3000	5	Fast spin echo

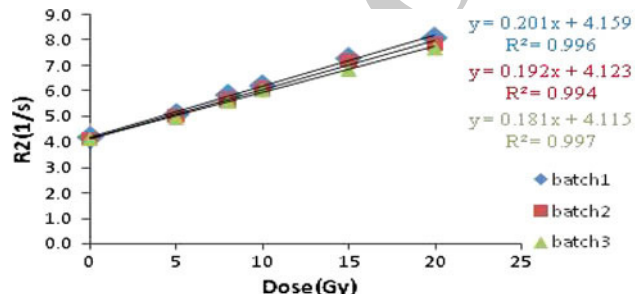


Fig. 2 Reproduction of DEMBIG gel for three batches using optimal protocol

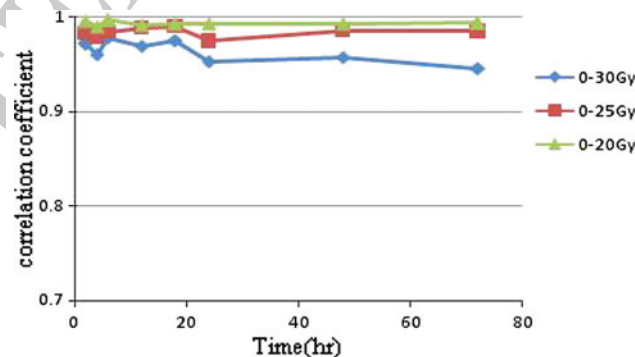


Fig. 3 Temporal stability of DEMBIG gel

Verification energy dependence of DEMBIG gel dosimeter 164

165 Figure 4 shows the DEMBIG gel dosimeter R_2 -dose
 166 response with different photon energies; therefore no sig-
 167 nificant energy effects in DEMBIG gel have been observed
 168 using the optimal protocol of MRI evaluation when photon
 169 energy used 6 and 10 MV.

Verification of isocenter dose in clinical practice 170

171 Figure 5 shows calibration R_2 dependence on the absorbed
 172 dose for DEMBIG gel at optimal MR setup. The data
 173 revealed no significant difference in dose sensitivity and R_2
 174 (0) parameters, which were important for further

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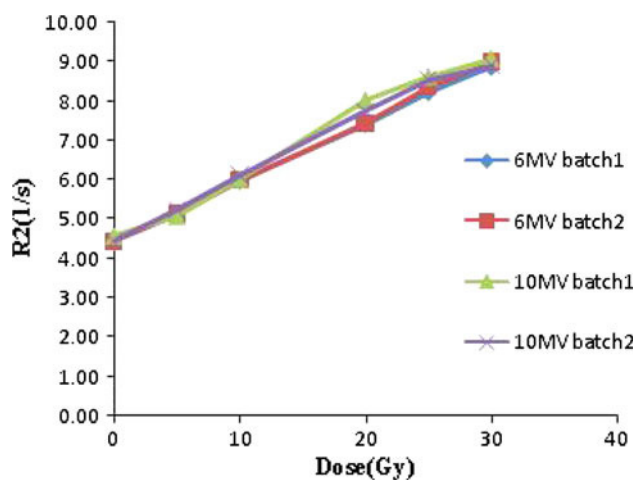


Fig. 4 Energy dependence of DEMBIG gel

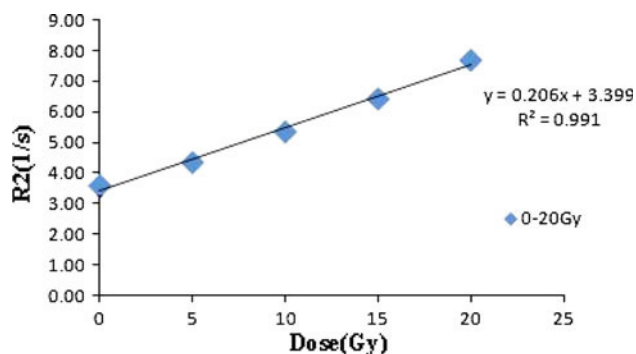


Fig. 5 Calibration R_2 dependence for DEMBIG gel

Table 3 Comparison of isocenter dose with treatment plan and DEMBIG gel

Item	Dose of isocenter (Gy)	Dose percent of isocenter (%)	Difference (%)
Treatment plan	20	100.1	0
DEMBIG gel	20.41	102.0	2

175 calculations of dose distribution. The fit curve function is
 176 $R_2 = 3.399 + 0.206D$ with a dose range of 0–20 Gy. The
 177 isocenter dose of 270 ml gel phantom is 20.41 Gy using
 178 the fit curve function. Table 3 compares the original
 179 treatment plan with the gel phantom. The difference was
 180 2% compared with the original treatment plan.

181 Discussion

182 Accurate measurement of absorbed dose from ionizing
 183 radiation is important. The DEMBIG gel is one hydrogel of

Table 4 Sensitivity of different polymer gel formulations [5]

No.	Type	Sensitivity (1/s)
1	PAG	0.33
2	PAGAS	0.008
3	nMAG	2.1
4	HEMA	0.046
5	DEMBIG	0.181

184 chemical dosimeter with feature of water or tissue equiv-
 185 alent and operation dose range was from 0 to 20 Gy read
 186 out by MRI to fit in clinical application.

187 Free radical reactions initiate the gel polymerization
 188 process. Nevertheless, molecular oxygen is an efficient
 189 “scavenger” of free radicals and inhibits polymerization of
 190 the gel dosimeter. The polymer gel dosimeter includes
 191 hypoxic and anoxic gel dosimeters.

192 Tetrakis (hydroxymethyl) phosphonium chloride (THPC),
 193 a new antioxidant, can solve the oxygen problem. Formu-
 194 lation of DEMBIG gel did not put in the THPC as an
 195 antioxidant, because the pH value of DEMBIG is 14 and
 196 THPC is 3, solidifying the solution from neutralization.
 197 Another method to remove oxygen uses nitrogen or argon
 198 as an antioxidant. Argon (20 psi/min) replaces oxygen with
 199 a concentration below 1% in the fabrication process to
 200 prevent the oxygen from dissolving into the solution.

201 De Deene [8] mentioned MRI as a non-destructive mea-
 202 surement method of the gel dosimeter in his review article.
 203 The MRI takes the R_1 mapping sequence; R_2 mapping
 204 sequence, and magnetization transfer (MT) to obtain an
 205 image for mapping dose distribution. Polymer gel dosime-
 206 ters are based on the conversion of comonomers to polymer
 207 aggregates upon irradiation. This reaction alters the mobility
 208 of surrounding water molecules, resulting in a change in R_1
 209 and R_2 . The dose–response of R_2 in gelatin based polymer
 210 gel dosimeters, however, is more pronounced than that of
 211 R_1 . The R_2 mapping sequence uses single- spin echo
 212 sequence, fast single-spin echo sequence, and multi-spin
 213 echo sequence for measuring the gel dosimeter.

214 The most important consideration of the polymer gel
 215 dosimeter is the correlation coefficient and dose response
 216 sensitivity. The range of dose response and sensitivity of
 217 DEMBIG gel were 0–20 Gy and 0.181(1/s), respectively.
 218 Jirasek listed the sensitivity to radiation of different poly-
 219 mer gel formulations [9], shown in Table 4. He investi-
 220 gated polymer gel solutions with various gelling agents,
 221 such as gelatin and agarose. The gel dosimeter used gelatin
 222 as gelling agents to improve sensitivity on MRI due to a
 223 low background. Overall sensitivity was between 0.008 and
 224 2.1(1/s). The sensitivity of the DEMBIG dosimeter was
 225 0.181(1/s) in gelatin with development potential. Future

226 work needs to prove the other physical properties of clinical
227 perspectives with DEMBIG.

228 In conclusion, the optimal scan parameter of MRI
229 as a measurement method of DEMBIG gel were short
230 TE:31.5 ms, long TE:158 ms, TR = 3 s, slice thickness =
231 5 mm, FOV 256*256 using the fast spin echo. The range of
232 dose response was 0–20 Gy, correlation coefficient difference
233 was 0.997, sensitivity was 0.181(1/s), and temporal
234 stability at 24 h for DEMBIG gel was based on the MRI
235 optimal scan parameter. DEMBIG gel does not possess
236 energy dependence. The difference of DEMBIG gel was less
237 than 3% at 0–15 Gy in verifying isocenter dose in the clinical
238 radiosurgery process. These data show that DEMBIG gel is a
239 potential candidate for the 3D dosimeter.

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