

1 **Title:**

2 Phthalate exposure in pregnant women and their children in central Taiwan

3 **Authors names and affiliations:**

4 Susana Lin

5 E-mail: susanalin@nhri.org.tw

6 Affiliation: Division of Environmental Health and Occupational Medicine, National
7 Health Research Institutes, Taiwan

8 Hsiu-Ying Ku

9 E-mail: shiuo@nhri.org.tw

10 Affiliation: Graduate Institute of Life Science, National Defense Medical Center

11 Pen-Hua Su

12 E-mail: jen@csh.org.tw

13 Affiliation 1: Department of Pediatrics, Chung Shan Medical University Hospital,
14 Taichung, Taiwan.

15 Affiliation 2: School of Medicine, Chung Shan Medical University, Taichung, Taiwan.

16 Jein-Wen Chen

17 E-mail: jwchen@nhri.org.tw

18 Affiliation: Division of Environmental Health and Occupational Medicine, National
19 Health Research Institutes, Taiwan

20 Po-Ching Huang

21 E-mail: pchuang@nhri.org.tw

22 Affiliation: Division of Environmental Health and Occupational Medicine, National
23 Health Research Institutes, Taiwan

24 Jürgen Angerer

25 E-mail: juergen.angerer@ipasum.imed.uni-erlangen.de

26 Affiliation: Institute ad Outpatient Clinic of Occupational, Social and Environmental
27 Medicine, University of Erlangen-Nuremberg

28 Shu-Li Wang

29 E-mail: slwang@nhri.org.tw

30 Affiliation 1: Division of Environmental Health and Occupational Medicine, National
31 Health Research Institutes, Taiwan

32 Affiliation 2: School of Medicine, Chung Shan Medical University, Taichung, Taiwan

33 **Corresponding author:**

34 Shu-Li Wang

35 National Health Research Institutes

36 Division of Evironmental Health and Occupational Medicine

37 No35, Keyan Road, Zhunan Town, Miaoli County 35035, Taiwan.

38 E-mail: slwang@nhri.org.tw

39 Phone: +886-37-246166-36519

40 Fax: +886-37-587406

41

42 **Abstract**

43

44 Phthalate exposure has been found to be associated with endocrine disruption,
45 respiratory effects, and reproductive and developmental toxicity. The intensive use of plastics
46 may be increasing the exposure to phthalates in the Taiwanese population, particularly for
47 young children.

48 We studied phthalate metabolites in pregnant women and their newborns in a general
49 population in Central Taiwan. A total of 430 pregnant women agreed to participate and one
50 hundred maternal urine samples and thirty paired cord blood and milk samples were
51 randomly selected from those participants. Eleven phthalate metabolites (MEHP,
52 5OH-MEHP, 2cx-MEHP, 5cx-MEPP, 5oxo-MEHP, MiBP, MnBP, MBzP, OH-MiNP,
53 oxo-MiNP, and cx-MiNP) representing exposure to five commonly used phthalates (DEHP,
54 DiBP, DnBP, BBP, DiNP) were measured in the urine of pregnant women, cord serum and
55 breast milk after delivery, and in the urine of their children. Exposure was estimated based on
56 excretion factors and correlation among metabolites of the same parental compound. Thirty
57 and fifty-nine urinary samples from 2 and 5 years-old children, respectively, were randomly
58 selected from the 185 children who were followed successfully.

59 The total urinary phthalate metabolite concentration (geometric mean, $\mu\text{g/L}$) was
60 found to be higher in 2-year-olds (398.6) and 5-year-olds (333.7) than in pregnant women

61 (205.2). Metabolites in urine are mainly from DEHP. The proportion of DiNP metabolites
62 was higher in children's urine (4.39 and 8.31%, ages 2 and 5) than in that of adults (0.83%)
63 (p<0.01). When compared with urinary levels, phthalate metabolite levels were low in cord
64 blood (37.45) and milk (14.90). DEHP metabolite levels in women's urine and their
65 corresponding cord blood were significantly correlated. When compared to other populations
66 in the world, DEHP derived metabolites in maternal urine in Taiwan were higher, while
67 phthalate metabolite levels in milk and cord blood were similar. The levels of phthalate
68 metabolites in milk and cord blood were comparable to those found in other populations.
69 Further studies of the effects on health related to DEHP and DiNP exposure are necessary.

70 **Keywords:**

71 Phthalate; environmental exposure; Taiwan; cord blood

72

73

74 **Introduction**

75 Phthalates are chemicals widely used in commercial products such as plastic softeners
76 and solvents in personal care products, lubricants and insect repellents (Fay et al., 1999; Koo
77 and Lee, 2004; Lee et al., 2005). Potential sources of exposure for di(2-ethylhexyl)phthalate
78 (DEHP) include polyvinylchloride containing medical devices, food packaging, plastic toys,
79 furniture, and car upholstery. Di-n-butyl phthalate (DnBP) is present in medicines, cosmetics,

80 cellulose acetate plastics, latex adhesives, nail polish and other cosmetic products; butyl
81 benzyl phthalate (BBP) is found in vinyl flooring, adhesives, sealants, food packaging,
82 furniture upholstery, vinyl tile, carpet tiles, artificial leather, and di-isononyl phthalate (DiNP)
83 is widely used in children's toys (Sathyanarayana, 2008). Recent studies suggest that the
84 intensive use of plastic material in Taiwan may be increasing the exposure to DEHP in the
85 Taiwanese population (Chen et al., 2008).

86 According to some epidemiological studies, phthalate exposure is associated with
87 adverse health outcomes, such as shorter anogenital distances at birth (Swan, 2006),
88 respiratory effects (Hoppin et al., 2004; Jaakkola et al., 1999; Jaakkola et al., 2000), and
89 increased waist circumference and insulin resistance (Stahlhut et al., 2007). Exposure to
90 MnBP, mono-benzyl phthalate (MBzP), and Mono-2-ethylhexyl phthalate (MEHP) is
91 associated with an overall pattern of decline in sperm motility (Duty et al., 2004).

92 Pregnant and lactating women represent a population of special concern because of
93 the potential impact of their exposure to phthalates on the fetus and nursing infant. Exposure
94 data for children under age 6 are scarce (Jahnke et al., 2005; McKee, 2004; NTP-CERHR,
95 2003a). Metabolites of DEHP, DBP and BBP have been monitored in children aged 2-6
96 (Koch et al., 2004; Koch et al., 2005). A decrease in the anogenital distance in male infants
97 has been found to be associated with phthalate exposure, as determined by urinary MEP,
98 MBP, MBzP and MiBP levels (Swan et al 2005). In another study, MBP in maternal urine

99 and amniotic fluid was found to be associated with a shorter anogenital distance only in
100 female infants (Huang et al., 2009). In pregnant women, urinary MBP is negatively correlated
101 with thyroxine, free thyroxine and FT4 levels (Huang et al., 2007).

102 Many experimental studies using different laboratory animals (primarily rats) have
103 examined the reproductive toxicity, developmental toxicity, endocrine disruption, and
104 genotoxicity that might be induced by phthalic acids. For example, anti-androgenic effects
105 including delayed puberty in F₀, decreased sperm production and fecundity in F₁,
106 malformations in F₁ reproductive organs, and decreased F₂ litter size, were reported for DBP
107 (NTP-CERHR, 2003b). DBP's metabolite, MBP, is responsible for the toxic effects
108 associated with DBP exposure. These include increased prenatal mortality, decreased fetal
109 weight, cleft palate, fused sternbrae, reduced anogenital distance in males, cryptorchidism,
110 hypospadias, and agenesis of the epididymides or seminal vesicles (NTP-CERHR, 2003b).
111 High doses of DiNP caused an increase in liver weight, peroxisomal proliferation, skeletal
112 variations and renal toxicity in a one-generation and a two-generation toxicity study
113 (Moorman et al., 2000; NTP-CERHR, 2003a). In rats, BBP exposure was associated with
114 decreased testicular weight, reduced ano-genital distance, increased incidence of nipple
115 retention and decreased birth weight in both sexes of the first filial generation (Gray et al.,
116 2000; Parks et al., 2000). Treatment with DEHP was also associated with altered ano-genital
117 distance and nipple retention (NTP-CERHR, 2005).

118 In vitro studies help in understanding the possible mechanisms of toxicity. Phthalates
119 and their metabolites can bind to several nuclear receptors and act as endocrine disruptors or
120 metabolic disruptors (Desvergne et al., 2009). In a series of reporter gene assays, DBP, MBP
121 and DEHP have been found to have both anti-androgenic and androgenic activities at
122 different concentrations. These compounds also showed thyroid receptor (TR) antagonistic
123 activity. Only DBP was reported to have estrogenic activity (Shen et al., 2009). BBP has an
124 affinity for binding to estrogen receptors (ER) (Blair et al., 2000; Hashimoto et al., 2000;
125 Matthews et al., 2000; Zacharewski et al., 1998), and activates ER-mediated transcription
126 (Coldham et al., 1997; Harris et al., 1997; Hashimoto et al., 2000; Nishihara et al., 2000;
127 Zacharewski et al., 1998). DEHP has a weak agonistic activity at aryl hydrocarbon receptors
128 (AhR) (Kruger et al., 2008), constitutive androstane receptors (CAR, Nr1i3) (Eveillard et al.,
129 2009), and Pregnane X nuclear receptors (PXR, Nr1i2) (Cooper et al., 2008; Hurst and
130 Waxman, 2004). Phthalates, especially MEHP, interfere with steroid production, particularly
131 estradiol production and aromatase expression. A possible mechanism for this interference is
132 through mediation at peroxisome proliferator-activated receptors (PPAR) (Lovekamp-Swan
133 et al., 2003; Lovekamp and Davis, 2001). MEHP is a true ligand for all three PPAR isotypes
134 and a selective modulator of PPAR gamma (Desvergne et al., 2009). BBP does not activate
135 progesterone receptor-mediated transcription (Tran et al., 1996) or AR-mediated transcription
136 (Sohoni and Sumpter, 1998). While BBP alone did not show a significant agonistic AhR

137 effect, it enhanced TCDD induced AhR activity in a dose-dependent manner (Kruger et al.,
138 2008). BBP exposure in female rats is also associated with a significant increase in liver
139 Ethoxyresorufin-O-deethylation (EROD) activity (Singletary et al., 1997). BBP also induces
140 human breast cancer cell proliferation (Harris et al., 1997; Korner et al., 1998; Soto et al.,
141 1997).

142 We monitored eleven phthalate metabolites (MEHP, 5OH-MEHP, 2cx-MEHP,
143 5cx-MEPP, 5oxo-MEHP, MiBP, MnBP, MBzP, OH-MiNP, oxo-MiNP, and cx-MiNP) in
144 pregnant women (urine, serum and milk), their newborns (cord blood) and prospectively in
145 their children at ages 2-3 and 5-6 (urine) from a general population in Central Taiwan. These
146 eleven metabolites were derived from exposure to five commonly used phthalates: DEHP
147 (MEHP, 5OH-MEHP, 2cx-MEHP, 5cx-MEPP, and 5oxo-MEHP), DiBP (MiBP), DnBP
148 (MnBP), BBP (MnBP and MBzP), and DiNP (OH-MiNP, oxo-MiNP, and cx-MiNP).

149 Exposure to phthalic acids was estimated based on the 95% confidence interval for the level
150 of each measured urinary metabolite, and excretion fraction published in the literature.

151 Correlations among metabolites of the same parental compounds and among different types
152 of samples from pregnant women and their corresponding children were also tested.

153

154 **Methods**

155

156 *Participants, specimen and data collection*

157 The subjects were pregnant women from Central Taiwan, aged between 25 and 35
158 and without clinical complications. We invited all pregnant women visiting the local medical
159 center between December 2000 and November 2001 to participate in this study. A total of
160 610 women were approached, and 430 (participation rate: 75%) agreed to be interviewed. To
161 the best of our knowledge, those who refused to participate did not differ in age or social
162 status from those who did participate. All of the participants completed a questionnaire
163 concerning maternal age, parity, baby's weight, educational level, disease history, dietary and
164 smoking habits, and breast-feeding history. Maternal urine was collected from subjects
165 during the third trimester of pregnancy (28-36 weeks), and umbilical-cord serum was
166 collected upon delivery. Women who agreed to collect samples of breast milk were trained in
167 collection procedures in order to minimize the risk of contamination. A total of 175
168 participants provided adequate breast-milk (>60 ml) samples. Newborns were followed again
169 when they were 2-3 years old (185 subjects, in 2003-2004) and 5-6 years old (185 subjects, in
170 2006-2007). For newborns and children aged 2-3, spot urine samples were collected in a
171 pediatric urine bag with the assistance of a parent at the hospital. For children aged 5-6, urine
172 samples were collected in a glass beaker. Immediately after collection, urine samples were
173 transferred into amber glass bottles and stored at -20°C for analyses of phthalate monoesters
174 and creatinine. One hundred maternal urine samples, fifty-nine urine samples from children

175 aged 5-6 , 30 urine samples from children aged 2-3 , and 30 paired cord blood samples and
176 milk samples were randomly retrieved and sent for analysis. The difference in the number of
177 children's samples was due to freshness and budget limitations, and the fact that one of the
178 samples had insufficient volume for creatinine analysis.

179

180 *Analysis of phthalate metabolites*

181 The concentrations of eleven phthalate metabolites (MEHP, 5OH-MEHP, 2cx-MEHP,
182 5cx-MEPP, 5oxo-MEHP, MiBP, MnBP, MBzP, OH-MiNP, oxo-MiNP, and cx-MiNP) in
183 urine, cord serum and breast milk were determined with LC-MS-MS methods as described in
184 previous publications (Koch et al., 2003; Preuss et al., 2005) by Dr Jürgen Angerer's lab at
185 the University of Erlangen, Germany. Metabolite concentrations are expressed as "µg/L" or
186 "µg/g creatinine". "Total metabolites" refers to the sum of metabolites calculated by adding
187 the concentrations of all metabolites.

188

189 *Determination of creatinine levels in urine*

190 Urinary creatinine levels were measured by Kaohsiung Medical University Chung-Ho
191 Memorial Hospital, using the spectrophotometric method, with picric acid as the reactive
192 agent, and read at 520nm.

193

194 ***Statistical methods***

195 We verified the distribution of data for phthalate metabolites for normality. As the
196 data were generally skewed slightly to the right, log transformations of phthalate metabolite
197 values and geometric means were applied in parametric statistical tests. Metabolite levels
198 under the limits of detection (<LOD) were recorded as half the LOD value. Samples from
199 male and female children were considered both separately and together to determine gender
200 differences. Pearson's correlation tests (r = Pearson correlation coefficient) were used to
201 check for correlations among values of metabolites from the same parental compound in
202 different matrices. The metabolite profile of the urine samples from each subject was
203 expressed as a percentage. The geometric means of the groups were used for parametric tests
204 and medians were used for non parametric tests. The Statistical Package for Social Science
205 (version 15.0; SPSS, Chicago, IL, USA) was used for statistical analysis.

206

207 ***Estimation of parental compound levels***

208 The amount of parental phthalate to which each subject might have been exposed was
209 estimated using excretion fractions reported in the literature (Wittassek et al., 2007). We
210 calculated the range of possible original parental compound levels, based on the 95%
211 confidence interval for the level of each urinary metabolite.

212

213 (1) Parental compound = metabolite concentration × excretion fraction

214

215 We also calculated the daily intake (Estimated Daily Intake, EDI) for these parental
216 compounds, taking into account an average body weight of 55 kg for Taiwanese women, 16.5
217 kg for children aged 2-3, and 20 kg for children aged 5-6 (DOH 2000). A daily urine
218 excretion of 0.8-2.2L was calculated for pregnant women, 0.6L for children aged 2-3 and 0.7
219 L for children aged 5-6 (Fleisher et al., 2002).

220

221 (2) Estimated daily intake = estimated parental compound concentration × daily urine

222 excretion × average body weight

223

224 As an example, to estimate the maximum exposure to BBP for children aged 5 and 6,
225 we used the MnBP concentration in their urine (GM max= 4.86), and an excretion rate of
226 73%, as reported by Wittassek et al 2007; therefore, the parental compounds from which this
227 metabolite originated should be $4.86 \times 100 \div 73 = 6.66 \mu\text{g/L}$. For the estimation of daily intake,
228 a daily urinary excretion of 0.7L urine/day for children this age was multiplied by the
229 estimated parental compound concentration and the result divided by the average body
230 weight of 20 kg. The estimated daily intake in this example would be $6.66 \times 0.7 = 0.23 \mu\text{g/kg}$
231 bw/day.

232

233 **Results**

234

235 *General characteristics of the population*

236 Table 1 shows the general characteristics of participating subjects, including maternal
237 age, maternal education level, and breast feeding patterns. The average age of the mothers
238 was 29, and the average pre-pregnant BMI was 21. Forty-one percent of the breast-feeding
239 mothers were taking supplements: vitamins, calcium, folic acid or Chinese herbs. Forty-six
240 percent of the infant subjects were male. Mean body weights at birth for the 2-3-year-old
241 cohort and the 5-6-year-old cohort were 3290±460g and 3240±450g, respectively.

242

243 *Urinary metabolite levels*

244 Urinary phthalate metabolite levels in children at ages 2 and 5 and pregnant women
245 were compared using values with and without creatinine adjustment (Table 2). Generally,
246 metabolite levels were higher in children than in pregnant women. This could be observed
247 whether creatinine adjustment was applied or not. Total phthalate metabolite concentrations
248 without creatinine adjustment were found to be higher in 2-year-old children (GM =
249 398.6µg/L, 282.6–562.3) and 5-year-old children (333.7µg/L, 251.8–442.2) than in pregnant
250 women (205.2µg/L, 172.7–243.8). When creatinine adjustment was applied, children
251 appeared to have even higher phthalate metabolite concentrations; however, we should take

252 into account the fact that the level of creatinine in pregnant women's urine was higher than
253 that in children (PW: 76.60µg/L; 5Y: 59.53µg/L; and 2Y: 62.28µg/L).

254 **Analysis of the proportion of metabolites in urine helps identify the parental**
255 **compound and source of exposure.** Phthalate metabolites in urine were mainly those from
256 DEHP, followed by metabolites from DnBP or BBP and those from DiNP. The sums of
257 urinary DEHP metabolites, with a GM of 102.2 µg/L for pregnant women, 152.3µg/L for
258 5-year old children and 200.3µg/L for 2-year old children, were proportionally higher. The
259 second most abundant metabolite was MnBP (Pw: 72.29µg/L; 5y: 75.09µg/L; 2y:
260 100.44µg/L). It was followed by MiBP (Pw: 12.49µg/L; 5y: 25.24µg/L; 2y: 17.21µg/L), and
261 MBzP (Pw: 0.96µg/L; 5y: 3.61µg/L; 2y: 3.40µg/L). The ratio of DiNP metabolites/Total
262 phthalate metabolites observed in children's urine samples was higher than that in adult
263 samples. The total DiNP metabolite concentration was 1.71µg/L for pregnant women,
264 27.73µg/L for 5-year old children and 17.46µg/L for 2-year old children. The proportion of
265 each metabolite over total phthalate metabolite is shown in Table 3. **No gender difference**
266 **was found at any age, either for the total concentration or for any particular metabolite.**

267

268 *Phthalate metabolites in cord sera and milk samples*

269 Only MEHP, MiNP and MnBP were detected in some of the breast milk samples
270 (Table 4). MEHP was detected in 73% of the samples. When compared to urine, the

271 concentrations of phthalate metabolites were much lower in milk and cord blood samples.
272 The compositions of the phthalate metabolites found in these matrices were also very
273 different. The most abundant metabolite in cord blood serum samples was MEHP, rather than
274 oxidized metabolites of DEHP (Table 4).

275

276 *Correlational studies*

277 We found that DEHP metabolite concentrations in urine samples of pregnant women
278 (without creatinine adjustment) and their corresponding cord blood samples were
279 significantly correlated for two of the metabolites, 5cxMEPP ($r=0.53$, $p<0.01$) and
280 2cxMMHP ($r=0.44$, $p<0.01$) (Table 5). This correlation was still observed when the
281 creatinine adjustment was applied to the urine samples (data not shown). Oxo-MiNP
282 concentrations in urinary samples of children at both ages were well correlated ($r =0.41$,
283 $p<0.05$).

284 Concentrations of different metabolites derived from DEHP and DiNP within the
285 same sample were well correlated as were the sum of metabolites from the same parental
286 compound (Pearson's correlation between 0.7 and 0.988). DBzP-derived metabolites are
287 MBzP and MnBP. We observed no correlation between the concentrations of these two
288 metabolites (data not shown). DnBP is another source for MnBP. This lack of DBzP
289 derivation suggests that DnBP is the main contributing source for the formation of MnBP.

290

291 *Estimation of parental compound exposure*

292 The estimated parental contribution to actual phthalate metabolite levels in urinary
293 samples and the estimated daily intake of these compounds suggested that children were
294 more exposed to phthalate than were pregnant women. Children aged 2-3 seem to have been
295 exposed to more total phthalate, particularly to DEHP and DnBP at that younger age than
296 when they were 5-6 years old. An estimated daily intake of each parental compound and a
297 tolerable daily intake (TDI) reference are shown in Table 6. The exposure of the participants
298 in this project did not exceed tolerable daily intake as determined by the European Food
299 Safety Authority (2005).

300

301 **Discussion**

302 Compared to a CDC study of a population aged 6 and older in the USA (EPA, 2005),
303 the phthalate metabolite levels in the current study (as a geometric mean) were higher for
304 MEHP, 5OH MEHP, 5oxoMEHP, 5cxMEPP and MnBP, and lower for MBzP. When
305 compared to a German study of nursery school children aged 2-6 (Koch et al., 2004), the
306 level of MEHP in our study (as a median) was higher, while the levels of 5OH MEHP, 5oxo
307 MEHP, MnBP and MBzP were lower. When compared to levels in children aged 3-5 in the
308 GerES IV study (Becker et al 2009), only the MEHP level in our study was higher. The levels

309 of 5OH MEHP, 5oxo MEHP and MnBP in our study were lower than those in a Japanese
310 study performed exclusively on pregnant women (Suzuki et al., 2009). As stated in previous
311 reports about human DEHP metabolism, the urinary DEHP metabolites are oxidized products,
312 rather than monoesters, and the simple monoester MEHP was the dominant metabolite in
313 blood serum (Wittassek et al., 2007). The MEHP concentration was also found to be higher
314 than that of other oxidized metabolites in our cord serum samples (Table 4). Although
315 phthalate metabolite levels in milk and cord sera were low, cord blood metabolite levels were
316 well correlated with maternal urinary metabolite levels (Table 5); therefore, maternal urinary
317 metabolite levels may be useful in prenatal exposure studies.

318 The phthalate metabolites detected in breast milk in this study were mainly MEHP
319 and MnBP (Table 4). The levels of MEHP in milk were lower than those reported from the
320 USA (Calafat et al 2004), Denmark (Mortensen et al 2005, Main et al 2006), Finland (Main
321 et al 2006) and Italy (Latini et al 2009). Levels of MnBP were second highest in the studies in
322 Finland and Italy. Concentrations of phthalate metabolites in milk, blood and serum were
323 close to the limit of detection; therefore, phthalate metabolites in urine maybe more
324 informative than those in milk or serum (Hogberg et al., 2008).

325 We found no correlation among levels of metabolites in different matrices from the
326 same person, i.e., urine sample, milk sample, and cord blood. This observation is in
327 agreement with previous studies, where authors concluded that phthalate concentrations in

328 urine were higher than those in serum, milk or saliva, and did not reflect the concentrations of
329 oxidative metabolites in other body fluids, especially milk (Hines et al., 2009, Hogberg et al.,
330 2008). The composition of phthalate metabolites was very different in urine and milk samples.
331 Although it is not clear how phthalate metabolites are secreted into different body fluids or
332 travel across the placenta, they probably have different rates of secretion. An in vitro
333 experiment where the placenta was perfused with different phthalate monoesters showed
334 differential diffusion rates across the placenta for different phthalate metabolites (Mose et al
335 2007). This suggested that physical-chemical properties of the compounds may influence
336 the tissue distribution of the metabolites. This may also explain the lack of correlation among
337 different matrices in the same subject. In general, DEHP and DiNP derived metabolites better
338 represented the overall phthalate exposure as they were the most important contributors in
339 pregnant women and children, respectively.

340 The phthalate exposure profile suggests that the major health risks that might be
341 associated with this population could be anti-androgenic activity and tyrosine receptor
342 antagonistic activity related to MBP, DEHP, and DBP exposure. Mixtures of phthalate esters
343 exhibit cumulative, largely dose-additive effects on male rat reproductive tract development
344 when administered during sexual differentiation in utero (Howdeshell, Furr et al., 2008).
345 DBP+DEHP increased the incidence of many reproductive malformations including
346 epididymal agenesis and reduced androgen-dependent organ weights (Howdeshell et al 2007).

347 Mixtures including BBP, DBP, DEHP, DiBP and DPP reduced testosterone production in a
348 dose-additive manner (Howdeshell, Wilson et al 2008).

349 DiNP is widely used in toys and the proportions of its metabolites differed among
350 urine samples from 2-3 year-olds, 5-6 year-olds, and mothers. Our levels tended to be lower
351 than those in Japanese (Suzuki et al., 2009), US (CDC, 2005; Swan et al., 2005), and German
352 populations (Becker et al., 2009). Higher exposure to DiNP in children could be associated
353 with renal toxicity or problems in skeletal development.

354 MnBP and MBzP are breakdown products of BBP. In an experiment where stable
355 isotope-labelled BBP was administered to eight volunteers in a single dose, 73% was
356 excreted as MBzP and 6% as MnBP on a molar basis (Anderson et al., 2001). We found that
357 the concentration of MnBP was much larger than that of MBzP in our urine samples. Because
358 MnBP can also be derived from DnBP, our results suggest that DnBP is probably the
359 principal contributor of MnBP in this population.

360 Although estimated daily intake values were below the tolerable daily intake
361 according to CSTE 1998 (Koch et al, 2003) and EFSA (2005), MiBP and MnBP levels in
362 maternal urine were higher than those reported for mothers who gave birth to newborns with
363 shorter ano-genital distances (Swan, 2006; Swan et al., 2005), indicating a potential risk for
364 anti-androgenic effects in this population.

365 Higher exposures to DEHP in general and to DiNP in children seem to characterize
366 the phthalate exposure in this population. Although the daily intake values estimated from
367 urinary metabolites were within the tolerable daily intake levels as published by the European
368 Food Safety Authority (EFSA, 2005), it is worth noting that phthalate acids are currently
369 banned for use in toys and school supplies and, therefore, TDI is no longer applicable.
370 Further studies will look deeper into the health effects reported as a result of exposure to
371 DEHP and DiNP, e.g., gender-related behaviour, obesity, liver function, bone density, and
372 allergy, in the population we have been following for several years.

373

374 **Conclusions**

375 Eleven phthalate metabolites derived from exposure to five commonly used phthalates
376 were monitored. Higher exposures to DEHP in general, to DiNP for children, and lower
377 exposure to BBP are the characteristics of phthalate exposure for pregnant women and their
378 children in Central Taiwan.

379 We found no association between metabolite levels in the mothers and in their
380 children at ages 2 or 5. This may be due to differences in types of ingested food and habits.
381 Metabolite levels in different matrices from the same person, i.e., urine sample, milk sample,
382 and cord blood were not associated. Metabolites derived from the same parental compound,
383 as in the cases of DEHP and DiNP, were well correlated. Although the metabolites derived

384 from a particular phthalic acid are inter-correlated, the additive effects and variability of the
385 secretion fraction suggest that the measurement of individual metabolites is necessary in
386 order to estimate the total exposure.

387 Compared to other populations in the world, the phthalate metabolite levels found in
388 this study were higher for DEHP derived metabolites in urine, whether or not the data was
389 adjusted by creatinine. The levels of phthalate metabolites in milk and cord blood were
390 comparable to those found in other populations. Further follow up studies on health effects
391 related to DEHP and DiNP exposure are necessary.

392 **Acknowledgements**

393 We greatly appreciate the excellent assistance of Ms. Hsiao-Yen Chen for the various
394 specimen collections.

395

396 **Grant**

397 This work was supported by National Health Research Institutes, Taiwan (EO-97-PP-05 and
398 EO-98-PP-03)

399

400

401 **References**

402

403 Anderson, W.A., Castle, L., Scotter, M.J., Massey, R.C., Springall, C., 2001. A biomarker
404 approach to measuring human dietary exposure to certain phthalate diesters. Food
405 Addit. Contam. 18(12):1068-74.

406 Becker, K. Göen, T., Seiwert M., Conrad, A., Pick-Fuss, H., Müller, J., Wittassek, M.,
407 Schultz, C., Kolossa-Gehring, M., 2009. GerES IV: Phthalate metabolites and
408 bisphenol A in urine of German children. Int. J. Hyg. Environ. Health
409 212(6):685-692.

410 Blair, R.M., Fang, H., Branham, W.S., Hass, B.S., Dial, S.L., Moland, C.L., Tong, W., Shi,
411 L., Perkins, R., Sheehan, D.M., 2000. The estrogen receptor relative binding affinities
412 of 188 natural and xenochemicals: structural diversity of ligands. Toxicol. Sci.
413 54(1):138-53.

414 Calafat, A.M., Slakman, A.R., Silva, M.J., Herbert, A.R., Needham, L.L., 2004. Automated
415 solid phase extraction and quantitative analysis of human milk for 13 phthalate
416 metabolites. J of Chromatogr B Analyt Technol Biomed Life Sci, 805:49-56.

417 Chen, M.L., Chen, J.S., Tang, C.L., Mao, I.F., 2008. The internal exposure of Taiwanese to
418 phthalate - an evidence of intensive use of plastic materials. Environ. Int. 34(1):79-85.

419 Coldham, N.G., Dave, M., Sivapathasundaram, S., McDonnell, D.P., Connor, C., , Sauer,
420 M.J., 1997. Evaluation of a recombinant yeast cell estrogen screening assay. Environ.
421 Health Perspect. 105(7):734-42.

422 Cooper, B.W., Cho, T.M., Thompson, P.M., Wallace, A.D., 2008. Phthalate induction of
423 CYP3A4 is dependent on glucocorticoid regulation of PXR expression. Toxicol. Sci.
424 103(2):268-77.

425 Desvergne, B., Feige, J.N., Casals-Casas, C., 2009. PPAR-mediated activity of phthalates: A
426 link to the obesity epidemic? Mol. Cell. Endocrinol. 304(1-2):43-8.

427 DOH (Department of Health, Taiwan), 2000. Nutrition and Health Survey in Taiwan
428 (NAHSIT) 1993-1996.

429 Duty, S.M., Calafat, A.M., Silva, M.J., Brock, J.W., Ryan, L., Chen, Z., Overstreet, J.,
430 Hauser, R., 2004. The relationship between environmental exposure to phthalates and
431 computer-aided sperm analysis motion parameters. J. Androl. 25(2):293-302.

432 European Food Safety Authority. 2005. [BBP] The EFSA Journal. 241: 1-14

433 European Food Safety Authority. 2005. [DBP] The EFSA Journal. 242:1-17

434 European Food Safety Authority. 2005. [DEHP] The EFSA Journal. 243: -20

435 European Food Safety Authority. 2005. [DINP] The EFSA Journal. 244:1-18

436 European Food Safety Authority. 2005. [DIDP] The EFSA Journal. 245:1-14

437 Eveillard, A., Mselli-Lakhal, L., Mogha, A., Lasserre, F., Polizzi, A., Pascussi, J.M., Guillou,
438 H., Martin, P.G., Pineau, T., 2009. Di-(2-ethylhexyl)-phthalate (DEHP) activates the
439 constitutive androstane receptor (CAR): a novel signalling pathway sensitive to
440 phthalates. *Biochem. Pharmacol.* 77(11):1735-46.

441 Fay, M., Donohue, J.M., De Rosa, C., 1999. ATSDR evaluation of health effects of
442 chemicals. VI. Di(2-ethylhexyl)phthalate. Agency for Toxic Substances and Disease
443 Registry. *Toxicol. Ind. Health* 15(8):651-746.

444 Fleisher, G., Ludwig, S. 2002. *Synopsis of Pediatric Emergency Medicine Philadelphia, PA.*
445 Lippincott Williams & Wilkins.

446 Gray, L.E.Jr., Ostby, J., Furr, J., Price, M., 2000. Veeramachaneni DN, Parks L. Perinatal
447 exposure to the phthalates DEHP, BBP, and DINP, but not DEP, DMP, or DOTP,
448 alters sexual differentiation of the male rat. *Toxicol. Sci.* 58(2):350-65.

449 Harris, C.A., Henttu, P., Parker, M.G., Sumpter, J.P., 1997. The estrogenic activity of
450 phthalate esters in vitro. *Environ. Health Perspect.* 105(8):802-11.

451 Hashimoto, Y., Moriguchi, Y., Oshima, H., Nishikawa, J., Nishihara, T., Nakamura, M., 2000.
452 Estrogenic activity of chemicals for dental and similar use in vitro. *J. Mater. Sci.*
453 *Mater. Med.* 11(8):465-8.

454 Hines, E.P., Calafat, A.M., Silva, M.J., Mendola, P., Fenton, S.E., 2009. Concentrations of
455 phthalate metabolites in milk, urine, saliva, and serum of lactating North Carolina
456 women. *Environ. Health Perspect.* 117(1):86-92.

457 Högberg, J., Hanberg, A., Berglund, M., Skerfving, S., Remberger, M., Calafat, A.M.,
458 Filipsson, A.F., Jansson, B., Johansson, N., Appelgren, M., Håkansson, H., 2008.
459 Phthalate diesters and their metabolites in human breast milk, blood or serum, and
460 urine as biomarkers of exposure in vulnerable populations. *Environ. Health Perspect.*
461 116(3):334-9.

462 Hoppin, J.A., Ulmer, R., London, S.J., 2004. Phthalate exposure and pulmonary function.
463 *Environ. Health Perspect.* 112(5):571-4.

464 Howdeshell, K.L., Rider, C.V., Wilson, V.S., Gray, L.E. Jr., 2008. Mechanisms of actions of
465 phthalate esters, individually and in combination, to induce abnormal reproductive
466 development in male laboratory rats. *Environ Res* 108:168-176.

467 Howdeshell, K.L., Wilson, V.S., Furr, J., Lambright, C.R., Rider, C.V., Blystone, C.R.,
468 Hotchkiss, A.K., Gray, L.E. Jr., 2008. A Mixture of five phthalate esters inhibits fetal
469 testicular testosterone production in the Sprague-Dawley rat in a cumulative,
470 dose-additive manner. *Toxicolog Sci* 105(1), 153-165.

471 Howdeshell, K.L., Furr, J., Lambright, C.R., Rider, C.V., Wilson, V.S., Gray, L.E. Jr. 2007.
472 Cumulative Effects of Dibutyl Phthalate and Diethylhexyl Phthalate on Male Rat

473 Reproductive Tract Development: Altered Fetal Steroid Hormones and Genes.
474 Toxicolog Sci 99(1):190-202.

475 Huang, P.C., Kuo, P.L., Chou, Y.Y., Lin, S.J., Lee, C.C., 2009. Association between prenatal
476 exposure to phthalates and the health of newborns. Environ. Int. 35(1):14-20.

477 Huang, P.C., Kuo, P.L., Guo, Y.L., Liao, P.C., Lee, C.C., 2007. Associations between urinary
478 phthalate monoesters and thyroid hormones in pregnant women. Hum. Reprod.
479 22(10):2715-22.

480 Hurst, C.H., Waxman, D.J., 2004. Environmental phthalate monoesters activate pregnane X
481 receptor-mediated transcription. Toxicol Appl Pharmacol 199(3):266-74.

482 Jaakkola, J.J., Oie, L., Nafstad, P., Botten, G., Samuelsen, S.O., Magnus, P., 1999. Interior
483 surface materials in the home and the development of bronchial obstruction in young
484 children in Oslo, Norway. Am. J. Public Health 89(2):188-92.

485 Jaakkola, J.J., Verkasalo, P.K., Jaakkola, N., 2000. Plastic wall materials in the home and
486 respiratory health in young children. Am. J. Public Health 90(5):797-9.

487 Jahnke, G.D., Iannucci, A.R., Scialli, A.R., Shelby, M.D., 2005. Center for the evaluation of
488 risks to human reproduction--the first five years. Birth Defects. Res. B. Dev. Reprod.
489 Toxicol. 74(1):1-8.

490 Koch, H.M., Drexler, H., Angerer, J., 2004. Internal exposure of nursery-school children and
491 their parents and teachers to di(2-ethylhexyl)phthalate (DEHP). *Int J Hyg Environ*
492 *Health*; 207(1):15-22.

493 Koch, H.M., Gonzalez-Reche, L.M., Angerer, J., 2003. On-line clean-up by multidimensional
494 liquid chromatography-electrospray ionization tandem mass spectrometry for high
495 throughput quantification of primary and secondary phthalate metabolites in human
496 urine. *J. Chromatogr. B. Analyt. Technol. Biomed. Life Sci.* 784(1):169-82.

497 Koch, H.M., Preuss, R., Drexler, H., Angerer, J., 2005. Exposure of nursery school children
498 and their parents and teachers to di-n-butylphthalate and butylbenzylphthalate. *Int*
499 *Arch Occup Environ Health* 78(3):223-9.

500 Koo, H.J., Lee, B.M., 2004. Estimated exposure to phthalates in cosmetics and risk
501 assessment. *J. Toxicol. Environ. Health A* 67(23-24):1901-14.

502 Korner, W., Hanf, V., Schuller, W., Bartsch, H., Zwirner, M., Hagenmaier, H., 1998.
503 Validation and application of a rapid in vitro assay for assessing the estrogenic
504 potency of halogenated phenolic chemicals. *Chemosphere* 37(9-12):2395-407.

505 Kruger, T., Long, M., Bonefeld-Jorgensen, E.C., 2008. Plastic components affect the
506 activation of the aryl hydrocarbon and the androgen receptor. *Toxicology*
507 246(2-3):112-23.

508 Latini, G., Wittassek, M., Del Vecchio, A., Presta, G., De Felice, C., Angerer, J., 2009.
509 Lactational exposure to phthalates in Southern Italy. *Environ. Int.* 35(2):236-9.

510 Lee, S.K., Owens, G.A., Veeramachaneni, D.N., 2005. Exposure to low concentrations of
511 di-n-butyl phthalate during embryogenesis reduces survivability and impairs
512 development of *Xenopus laevis* frogs. *J. Toxicol. Environ. Health A* 68(10):763-72.

513 Lovekamp-Swan T, Jetten AM, Davis BJ. 2003. Dual activation of PPARalpha and
514 PPARgamma by mono-(2-ethylhexyl) phthalate in rat ovarian granulosa cells. *Mol*
515 *Cell Endocrinol* 201(1-2):133-41.

516 Lovekamp, T.N., Davis, B.J., 2001. Mono-(2-ethylhexyl) phthalate suppresses aromatase
517 transcript levels and estradiol production in cultured rat granulosa cells. *Toxicol. Appl.*
518 *Pharmacol.* 172(3):217-24.

519 Main, K.M., Mortensen, G.K., Kaleva, M.M., Boisen, K.A., Damgaard, I.N., Chellakooty, M.,
520 Schmidt, I.M., Suomi, A.M., Virtanen, H.E., Petersen, D.V., Andersson, A.M.,
521 Toppari, J., Skakkebaek, N.E., 2006. Human Breast Milk Contamination with
522 Phthalates and Alterations of Endogenous Reproductive Hormones in Infants Three
523 Months of Age. *Environ Health Perspect* 114(2):270-276.

524 Matthews, J., Celius, T., Halgren, R., Zacharewski, T., 2000. Differential estrogen receptor
525 binding of estrogenic substances: a species comparison. *J. Steroid Biochem. Mol. Biol.*
526 74(4):223-34.

527 McKee, R.H., 2004. Phthalate exposure and early thelarche. *Environ. Health Perspect.*
528 112(10):A541-3.

529 Moorman, W.J., Ahlers, H.W., Chapin, R.E., Daston, G.P., Foster, P.M., Kavlock, R.J.,
530 Morawetz, J.S., Schnorr, T.M., Schrader, S.M., 2000. Prioritization of NTP
531 reproductive toxicants for field studies. *Reprod. Toxicol.* 14(4):293-301.

532 Mortensen, G.A., Main, K.M., Andersson, A.M., Leffers, H., Skakkebaek, N.E., 2005.
533 Determination of phthalate monoesters in human milk, consumer milk, and infant
534 formula by tandem mass spectrometry (LC-MS-MS). *Anal Bioanal Chem*
535 382:1084-1092.

536 Mose, T., Mortensen, G.K., Hedegaard, M., Knudsen, L.E., 2007. Phthalate monoesters in
537 perfusate from a dual placenta perfusion system, the placenta tissue and umbilical
538 cord blood. *Reprod Toxicology* 23:83-91.

539 Nishihara, E., Nagayama, Y., Inoue, S., Hiroi, H., Muramatsu, M., Yamashita, S., Koji, T.,
540 2000. Ontogenetic changes in the expression of estrogen receptor alpha and beta in rat
541 pituitary gland detected by immunohistochemistry. *Endocrinology* 141(2):615-20.

542 National Toxicology Program – Center for the Evaluation of Risks to Human Reproduction.
543 2000. NTP-CERHR Monograph on the Potential Human Reproductive and
544 Developmental Effects of Di-isononyl Phthalate (DINP). Ntp Cerhr DINP 00.

545 National Toxicology Program – Center for the Evaluation of Risks to Human Reproduction.
546 2000. NTP-CERHR Monograph on the Potential Human Reproductive and
547 Developmental Effects of Di-n-Butyl Phthalate (DBP). Ntp Cerhr DBP 00.

548 Parks, L.G., Ostby, J.S., Lambright, C.R., Abbott, B.D., Klinefelter, G.R., Barlow, N.J., Gray,
549 L.E.Jr., 2000. The plasticizer diethylhexyl phthalate induces malformations by
550 decreasing fetal testosterone synthesis during sexual differentiation in the male rat.
551 *Toxicol Sci* 58(2):339-49.

552 Preuss, R., Koch, H.M., Angerer, J. 2005. Biological monitoring of the five major
553 metabolites of di-(2-ethylhexyl)phthalate (DEHP) in human urine using
554 column-switching liquid chromatography-tandem mass spectrometry. *J Chromatogr B*
555 *Analyt Technol Biomed Life Sci* 816(1-2):269-80.

556 Rastogi, S.K., Kesavachadran, C., Mahdi, F., Pandey, A., 2006. Phthalate exposure and
557 health outcomes. *Indian J Occup Environ Med* 10(3):111-115.

558 Sathyanarayana, S., 2008. Phthalates and Children's Health. *Curr Probl Pediatr Adolesc*
559 *Health Care* 38:34-49.

560 Shen, O., Du, G., Sun, H., Wu, W., Jiang, Y., Song, L., Wang, X., 2009. Comparison of in
561 vitro hormone activities of selected phthalates using reporter gene assays. *Toxicol*
562 *Lett.* 191:9-14.

563 Singletary, K., MacDonald, C., Wallig, M., 1997. The plasticizer benzyl butyl phthalate (BBP)
564 inhibits 7,12-dimethylbenz(a)anthracene (DMBA)-induced rat mammary DNA adduct
565 formation and tumorigenesis. *Carcinogenesis* 18(8):1669-73.

566 Sohoni, P., Sumpter, J.P., 1998. Several environmental oestrogens are also anti-androgens. *J.*
567 *Endocrinol.* 1998; 158(3):327-39.

568 Soto, A.M., Fernandez, M.F., Luizzi, M.F., Oles Karasko, A.S., Sonnenschein, C., 1997.
569 Developing a marker of exposure to xenoestrogen mixtures in human serum. *Environ.*
570 *Health Perspect.* 105 Suppl 3:647-54.

571 Stahlhut, R.W., van Wijngaarden, E., Dye, T.D., Cook, S., Swan, S.H., 2007. Concentrations
572 of urinary phthalate metabolites are associated with increased waist circumference
573 and insulin resistance in adult U.S. males. *Environ. Health Perspect.* 115(6):876-82.

574 Suzuki, Y., Niwa, M., Yoshinaga, J., Watanabe, C., Mizumoto, Y., Serizawa, S., Shiraishi, H.
575 2009. Exposure assessment of phthalate esters in Japanese pregnant women by using
576 urinary metabolite analysis. *Environ Health Prev Med* 14:180-187.

577 Swan, S.H., 2006. Prenatal phthalate exposure and anogenital distance in male infants.
578 *Environ. Health Perspect.* 114(2):A 88-9.

579 Swan, S.H., Main, K.M., Liu, F., Stewart, S.L., Kruse, R.L., Calafat, A.M., Mao, C.S.,
580 Redmon, J.B., Ternand, C.L., Sullivan S, Teague, JL. 2005.. Decrease in anogenital

581 distance among male infants with prenatal phthalate exposure. Environ Health
582 Perspect 113(8):1056-61.

583 Tran, D.Q., Klotz, D.M., Ladlie, B.L., Ide, C.F., McLachlan, J.A., Arnold, S.F., 1996.
584 Inhibition of progesterone receptor activity in yeast by synthetic chemicals. Biochem.
585 Biophys. Res. Commun. 229(2):518-23.

586 USEPA, 2010. Urinary Phthalate Level.
587 <[http://cfpub.epa.gov/eroe/index.cfm?fuseaction=detail.viewMidImg&lShowInd=0&](http://cfpub.epa.gov/eroe/index.cfm?fuseaction=detail.viewMidImg&lShowInd=0&subtop=208&lv=list.listByAlpha&r=188260#9581)
588 [subtop=208&lv=list.listByAlpha&r=188260#9581](http://cfpub.epa.gov/eroe/index.cfm?fuseaction=detail.viewMidImg&lShowInd=0&subtop=208&lv=list.listByAlpha&r=188260#9581)> Last updated on Wednesday,
589 January 20th, 2010.

590 Wittassek, M., Wiesmüller, G.A., Koch, H.M., Eckard, R., Dobler, L., Muller, J., Angerer, J.,
591 Schlüter, C., 2007. Internal phthalate exposure over the last two decades--a
592 retrospective human biomonitoring study. Int. J. Hyg. Environ. Health
593 210(3-4):319-33.

594 Zacharewski, T.R., Meek, M.D., Clemons, J.H., Wu, Z.F., Fielden, M.R., Matthews, J.B.,
595 1998. Examination of the in vitro and in vivo estrogenic activities of eight commercial
596 phthalate esters. Toxicol Sci 46(2):282-93.