

# Botanical Drugs and Stem Cells

Po-Cheng Lin,<sup>1,2\*</sup> Li-Fu Chang,<sup>1\*</sup> Po-Yen Liu,<sup>3\*</sup> Shinn-Zong Lin,<sup>2,4\*</sup> Wan-Chen Wu,<sup>5</sup>

Wuen-Shyong Chen<sup>6</sup>, Chang-Hai Tsai,<sup>7,8</sup> Tzyy-Wen Chiou,<sup>1\*#</sup> and Horng-Jyh Harn<sup>9\*#</sup>

<sup>1</sup>Department of Life Science and Graduate Institute of Biotechnology, National Dong Hwa University, Hualien, Taiwan, R.O.C.

<sup>2</sup>Center for Neuropsychiatry, China Medical University Hospital, Taichung, Taiwan, R.O.C.

<sup>3</sup>Graduate Institute of Chinese Medical Science, China Medical University, Taichung, Taiwan, R.O.C.

<sup>4</sup>China Medical University Beigang Hospital, Yun-Lin, Taiwan, R.O.C.

<sup>5</sup>Graduate Institute of Basic Medical Science, China Medical University, Taichung, Taiwan, R.O.C.

<sup>6</sup>Department of Stem Cell Applied Technology, Gwo Xi Stem Cell Applied Technology, Hsinchu, Taiwan, R.O.C.

<sup>7</sup>Department of Pediatrics, China Medical University Hospital, Taichung, Taiwan, R.O.C.

<sup>8</sup>Department of Healthcare Administration, Asia University, Taichung, Taiwan, R.O.C.

<sup>9</sup>Department of Pathology, China Medical University and Hospital, Taichung, Taiwan,  
R.O.C.

Running title: Botanical Drugs and Stem Cells

\*These authors contributed equally to this article

#To whom correspondence should be addressed

#Corresponding authors:

Dr. Tzyy-Wen Chiou, PhD

Institute of Biotechnology

National Dong-Hwa University

No. 1, Sec. 2, Da Hsueh Rd., Shou-Feng, Hualien, Taiwan, R.O.C.

Tel: 886-3-8633638

Fax: 886-3-8630262

E-mail address: [twchiou@mail.ndhu.edu.tw](mailto:twchiou@mail.ndhu.edu.tw)

Dr. Horng-Jyh Harn

Department of Pathology, China Medical University and Hospital, 2 Yuh-Der Road,

Taichung, Taiwan, 40447, R.O.C.

TEL: 886-4-22052121 ext.2661

FAX: 886-4-22052121 ext.2566

E-mail address: [d15977@mail.cmuh.org.tw](mailto:d15977@mail.cmuh.org.tw)

## **ABSTRACT**

The potential to generate virtually any differentiated cell type from stem cells offers the possibility of creating new sources of cells for regenerative medicine. To realize this potential, it will be essential to control stem cell differentiation. Chinese herbal medicine is a major aspect of traditional Chinese medicine and is a rich source of unique chemicals. As such, individual herbs or extracts may play a role in the proliferation and differentiation of stem cells. In this review, we discuss some of the Chinese herbal medicines that are used to treat human diseases such as neuronal degenerative diseases, cardiovascular diseases, and osteoporosis. We also describe the relationship between Chinese herbal medicines and stem cell regulation.

Key words: stem cell, traditional Chinese medicine (TCM), neuronal degenerative, cardiovascular, osteoporosis

## **INTRODUCTION**

Stem cells are capable of proliferation, self-renewal, and production of differentiated functional progeny that are characteristic of the organ from which they were derived (3,41). Stem cell biology is one of the most challenging areas of biomedicine. Stem cells are undifferentiated cells that have the ability to proliferate for

an extended period and to differentiate into specific cell types under appropriate conditions. Stem cells are typically classified as embryonic stem cells (ESCs) and adult stem cells. ESCs are derived from the inner cell mass of the blastocyst, and have an unlimited capacity for self-renewal and the potential to differentiate into cells of all three germ layers. In contrast, adult stem cells are found in differentiated tissues, have a limited self-renewal capacity, and only differentiate into cell types found in the tissues from which they are derived. These characteristics of stem cells suggest that they hold great potential for revolutionizing regenerative medicine and tissue engineering (86). In this discipline, one of the biggest challenges is finding convenient and efficient approaches to modulate the fate of stem cells. Although genetic methods have been widely used in this area, and significant progress has been made in recent years (107), we are still far from the goal of conveniently and easily controlling stem cell fate.

### **TRADITIONAL CHINESE MEDICINES (TCMs)**

Native cultures all over the world have traditionally used herbs to maintain health and treat illnesses. In China, more than 3,200 herbs and 300 mineral and animal extracts are used in more than 400 different formulas. Chinese herbal medicine is a major aspect of TCM, which focuses on restoring a balance of energy, body, and spirit

to maintain health rather than treating a particular disease or medical condition. Herbs are used with the goal of restoring balance by nourishing the body. Chinese herbal medicine is not based on conventional Western concepts of medical diagnosis and treatment, but instead treats patients' main complaints or the patterns of their symptoms rather than the underlying causes. Practitioners attempt to prevent and treat imbalances, such as those caused by cancer and other diseases, with complex combinations of herbs, minerals, and plant extracts. One aspect of Chinese herbal medicine aims to restore or strengthen immunity and resistance to disease. Treatments undertaken with this goal are called “Fu Zheng” and are given as complementary therapy intended to reduce the side effects of conventional Western treatments.

## **TCM AND NEURONAL DEGENERATIVE DISEASE**

Parkinson's disease, is a common and debilitating neuronal degenerative disease resulting from massive degenerative loss of dopamine neurons (13). In East Asia, TCM practitioners use certain TCMS to treat neuronal degenerative diseases. A classification of the herbs discussed in this review, as distinguished by function, is presented in Table 1. In addition, many Chinese herbs or herbal extracts such as green tea polyphenols or catechins (5,13,39,40,62,63,69,88), oil from ganoderma lucidum spores (153),

triptolide from tripterygium wilfordii hook (64,65), polysaccharides from nerium indicum flowers (13), panax ginseng and ginsenoside (15,94,95,104,128), ginkgo biloba and EGb 761 (8,96,100,115,117,141), and polygonum (66) are able to attenuate degeneration of dopamine neurons and the symptoms caused by the neurotoxins 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine and 6-hydroxydopamine in both in vitro and in vivo conditions. Neuroprotective strategies to rescue nigral dopamine neurons from progressive death are currently being explored, including the use of Chinese herbs and herbal extracts that have shown potential clinical benefits of attenuating the progression of Parkinson's disease (67,146,147). Generally speaking, Chinese herbs or herbal extracts may promote neuronal survival and neurite growth, and may facilitate functional recovery from brain injuries by invoking distinct mechanisms that are related to their neuroprotective roles as antioxidants (67,101), dopamine transporter inhibitors (149,150), monoamine oxidase inhibitors (115,152), free radical scavengers (39,40), modulators of cell survival genes and signaling (63,81), anti-apoptotic agents (53,143,152), and even agents that improve blood circulation in the brain (48).

## **STEM CELLS DIFFERENTIATE INTO NEURON-LIKE CELLS**

Neural precursor or stem cells derived from brain have the capacity to self-renew and to generate progeny capable of differentiating into multiple distinct cell lineages that produce neurons and glia (35,76,83,98). Transplantation of human neural stem cells obtained from embryonic tissue leads to neuronal and astrocytic differentiation (9,84,116) and remyelination in the newborn mouse brain (33). Neural precursor cells derived from adult human brain have been reported to remyelinate axons and provide recovery of impulse conduction following transplantation into demyelinated rodent spinal cords (2). These studies have engendered intense interest because they suggest the possibility of using such cells for repair strategies in neurological diseases (2,9,84,105,116,135). However, difficulties remain regarding the relative inaccessibility of these cells for autologous cell therapies and the need for cell expansion using trophic factors. The potential to generate virtually any differentiated cell type from ESCs offers the possibility of establishing new models of mammalian development and creation of new sources of cells for regenerative medicine (52,86). To realize this potential, it will be essential to control ESC differentiation and to direct specific gene expression in these cells leading to a specific phenotype (Fig. 1). The derivation of human ESCs has brought cell therapy-based regenerative medicine



significantly closer to clinical application. However, expansion of undifferentiated cells and their directed differentiation in vitro are difficult to control (123).

Recent work has demonstrated that mesenchymal stem cells (MSCs) have the potential to differentiate into neurons both in vivo and in vitro (28,112-114). Under appropriate experimental conditions, MSCs can differentiate into several types of mesenchymal cells, including osteocytes, chondrocytes, and adipocytes, and they can also differentiate into non-mesenchymal cells, such as neural cells. Many protocols for inducing neuro-differentiation of MSCs in vitro have been reported (6,38,47,61,87,91,103,119,131), which may offer new methods of cell treatment for several types of neural diseases.

### **TCMs INDUCE NEURO-DIFFERENTIATION**

TCMs may enhance mentality, strengthen brain function, promote blood circulation, remove stasis, awaken patients, delay senescence, and modulate the immune system. Recent reports have shown differentiation of human MSCs into neuron-like cells with danshen injection (125,138,139). Increasing numbers of studies have indicated that many Chinese herbs or herbal extracts such as danshen (75,77,78), tetramethylpyrazine (79), panax notoginseng saponins (74,109), *Astragalus*

*mongholicus* (29,118), rhizoma gastrodiae (31), and panax ginseng (81,82,127) are able to direct the differentiation of human stem cells into neuron-like cells.

In one of our studies, the natural compound n-butylidenephthalide, which is isolated from a chloroform extract of *Angelica sinensis*, was investigated for its antitumor effects on glioblastoma multiform brain tumors both in vitro and in vivo (121). In a subsequent study, we observed that *Nurr1*, which plays an important role in stem cell differentiation into dopamine neurons (Fig. 1), was found to be upregulated immediately after n-butylidenephthalide treatment (16,71). Thus, many TCM compounds may have important uses in stem cell biology.

A Chinese herbal medicine study reported that ESC differentiation can be directed toward neurons and suggested that an effective inducible pharmaceutical dosage to produce neurons for clinical transplant may be possible. This study also suggested a new approach for developing herbal applications for the differentiation of MSCs and ESCs. New pharmaceutical strategies for treating Parkinson's disease will hopefully be developed by understanding the various active molecules and valuable combinations that contribute to the biological effects of Chinese herbs and herbal extracts.

## **EFFECTS OF STEM CELL THERAPY COMBINED WITH CHINESE HERBS**

## **ON MYOCARDIAL INFARCTION**

Cardiovascular diseases are the most serious groups of diseases in the world, and the prevalence and lethality rate are increasing every year. In America, more than 71 million people suffer from cardiovascular diseases, which are also the leading cause of death in many other countries, including most European and developed countries. Thus, a combination of stem cell therapy and Chinese herbs potentially could be used to treat cardiovascular diseases such as myocardial infarction and stenosis.

Adult cardiomyocytes do not regenerate, and therefore cannot be used to replace damaged cells following myocardial infarction (60,89,142). Recently, some progenitor or stem cells have been shown to differentiate into cardiomyocytes. Because of this property, many groups have attempted to use these progenitor or stem cells to replace damaged cardiomyocytes following myocardial infarction. However, only a portion of bone marrow–derived stem cells (MSCs) differentiate into cardiomyocytes, which distribute in the less-ischemic border zone. Thus, transplanted MSCs cannot be used to form functional cardiac tissue, making transplantation of MSCs for myocardial infarction therapy less than ideal. Consequently, considerable efforts have focused on the search for a new approach to enhance cardiogenic differentiation efficiency. Therefore, determining what kind of stem cell to use (i.e., ESCs, hematopoietic stem

cells, or MSCs) and the important molecular steps required to increase their differentiation is important. Surprisingly, many herbal medicines can increase the cardiogenic differentiation ability of MSCs. For example, in a search for molecules that promote cardiogenic differentiation, Cheng et al. found that the traditional herb “*Geum japonicum*,” which is frequently used by the Miao ethnic group in China’s Guizhou province, can enhance the cardiogenic differentiation ability of MSCs and significantly repair infarcted hearts (18,68). Further, this group identified a compound from *Geum japonicum* called cardiogenin, and showed that it is a major compound involved in cardiogenic differentiation (18).

In addition to increasing the differentiation efficiency, promoting stem cells to migrate to the infarcted heart from other tissues such as bone marrow is another strategy under investigation. Yang et al. found that the TCM “Wenyang Huoxue Recipe,” which consists of *Panax ginseng*, *Radix Aconiti lateralis preparata*, *Radix Ilex pubesceus*, and *Herbal Leonuri*, promotes stem cell mobilization. In this study, acute myocardial infarction patients were treated with Wenyang Huoxue Recipe for 10 successive days, which resulted in significantly increased numbers of peripheral CD34<sup>+</sup> cells. Through increasing the stem cell mobilization, more stem cells migrate to the infarcted heart area and may differentiate to the cardiomyocyte for cardiogenic repair.

Thus, Wenyang Huoxue Recipe can reduce the infarcted heart area and improved heart function (140).

A third important strategy for myocardial infarction therapy is to increase angiogenesis and endothelial cells in the infarcted heart. In TCM, ginseng has been shown to induce ESCs to differentiate into cardiomyocytes in vitro (106). Furthermore, saponin, a ginseng component, can stimulate myocardial tissue to secrete granulocyte colony-stimulating factor, which induces migration of MSCs to the myocardium. These MSCs can then differentiate into vascular endothelial cells, which can promote capillary regeneration in infarcted myocardial tissue and maintain the blood supply for protection of damaged cardiomyocytes (124,126).

#### STEM CELL THERAPY COMBINED WITH CHINESE HERBS FOR STENOSIS

Stenosis is one of the most common cardiovascular diseases in the world and is most commonly caused by atherosclerosis, a chronic inflammatory response in the walls of arteries. The process of atherosclerosis is divided into three stages. In the first stage, endothelial cells undergo inflammatory activation, which can be caused by smoking, hypertension, diabetes, autoimmune diseases, infections, or high cholesterol. Inflamed endothelial cells cannot secrete normal amounts of nitric oxide or

antithrombotic and vasodilating cytokines. Further, oxidative low-density lipoprotein (ox-LDL) accumulates in the arterial walls, and macrophages are attracted to engulf the ox-LDL. Large numbers of macrophages with ox-LDL, which are called foam cells, form the fatty streaks that are seen on the artery wall. In the second stage, accumulated platelets and foam cells secrete cytokines and growth factors, such as platelet-derived growth factor, epidermal growth factor, and fibroblast growth factor, which induce smooth muscle cells to migrate from the media and proliferate in the intima. The proliferating smooth muscle cells also secrete extracellular matrix, and the fatty streaks become fibrous plaques. In the third stage, fibrous plaques rupture, and platelets aggregate and form thrombi. This phenomenon reduces the lumen area and decreases the blood supply. Thus, atherosclerosis leads to stenosis and can lead to myocardial infarction in the heart during its most serious stages.

Currently, there are two strategies for combining stem cell therapy with TCM for treatment of stenosis. The first is to reduce the inflammatory effect, and the second is to regenerate functional endothelial cells. In 2008, Forte et al. used mesenchymal stem cells to treat balloon-injured rat arteries, an animal model of stenosis. They reported that mesenchymal stem cells can reduce the expression of proinflammatory genes, such as interleukin-1 and chemoattractant protein-1, thus limiting stenosis (34). Many TCMs

have been shown to promote stem cell migration and increase the number of stem cells in the circulatory system. For example, the Wenyang Huoxue Recipe can promote stem cell migration to the stenosis area (140), possibly reducing inflammation and the extent of stenosis.

In 2005, Yang et al. reported that the traditional Chinese herbs *Astragalus membranaceus* and *Panax notoginseng* can increase the proliferation of endothelial progenitor cells (136). In 2007, Ji et al. found that *Danshen* can significantly enhance the functional activity of endothelial progenitor cells in patients with hypercholesterolemia (46). Furthermore, in 2009, Shi et al. showed that the ginsenoside Rg1 can promote migration, adhesion, and proliferation of endothelial progenitor cells by increasing the number of endothelial progenitor cells in S phase and by decreasing the number of cells in the G0/G1 phase (108). Therefore, using *Astragalus membranaceus*, *Panax notoginseng*, and the ginsenoside Rg1 in stenosis therapy to regenerate functional endothelial cells may be beneficial. However, these studies have only been done in vitro, and thus future in vivo studies are required. The traditional Chinese herbs that contribute to stem cell therapy for myocardial infarction and stenosis are shown in Table 2.

## FAVORABLE EFFECTS OF TCM ON BONE HEALTH

Accumulating evidence suggests that natural products, such as herbal extracts and dietary supplements, have favorable effects on bone health (93,132). For example, turmeric root (*Curcuma longa*), which has long been used to treat inflammatory diseases, and its active constituents, curcuminoids, are known to have favorable effects on bone metabolism. *Acanthopanax senticosus* extract affects biochemical markers of bone turnover and increases in bone density in postmenopausal women (44). *Fructus Ligustri Lucidi*, a kidney-tonifying Chinese herb, can increase osteoblastic differentiation in rat osteoblast-like UMR-106 cells. *Fructus Ligustri Lucidi* extract improves bone properties in aged rats possibly via its direct action on osteoblastic cells to enhance the mineralization process (148). *To-Sa-Za* (TSZ-AE), the seed of *Cuscuta chinensis*, induces osteogenic activity in human osteoblast-like MG-63 cells (137). *Cistanche salsa*, a Chinese herb was found to contain an anti-osteoporotic compound, (2E,6R)-8-Hydroxy-2,6-dimethyl-2-octenoic Acid, an extract of *Cistanche salsa* significantly suppresses the bone weight loss that occurs in ovariectomized mice (134). In the search for new naturally occurring anti-osteoporosis agents in plants, Deyama et al. found that *Eucommia ulmoides* Oliv., also called Du-Zhong or Tu-Chung, is one of the earliest and most important edible crude herbs used for various medicinal purposes



in China, Japan, and Korea. The leaf and bark of this plant are rich in polyphenolic compounds such as lignans, phenolic acid, and flavonoids (27).

## **EFFECT OF STEM CELL THERAPY COMBINED WITH CHINESE HERBS ON OSTEOPOROSIS**

The two major cell types responsible for bone remodeling are osteoclasts, which resorb bone, and osteoblasts, which form new bone (37). Osteoporosis results from an imbalance in bone resorption and bone formation with a net bone loss that may be induced by several conditions, such as hormonal imbalance, disease, or medications (e.g., corticosteroids or anti-epileptic agents) (37). In TCM, herbs that strengthen bones can be used to treat bone diseases with the same symptoms as osteoporosis (Table 3). The currently available treatment, estrogen replacement therapy, is based on inhibition of bone resorption to prevent further bone loss. Many osteoporotic patients, however, have already lost a substantial amount of bone, and thus a method to increase bone mass by stimulating new bone formation is needed. During bone formation, osteoblasts are the key cell type for bone matrix formation and calcification. Because osteoclasts are responsible for bone resorption, they are one of the main targets for treatment of osteoporosis. Icariin is a prenylated flavonol glycoside contained in the herb

Epimedium, which has long been used to improve bone fracture healing or prevent osteoporosis because of the belief that the herb has bone-strengthening action. Icarin strengthens bones, and one of the mechanisms of this activity is the stimulation of proliferation and enhancement of the osteogenic differentiation of MSCs (12). Icarin can also inhibit the formation of osteoclasts as well as their bone resorption activity, which suggests that this compound may be the component responsible for the bone-strengthening action of Epimedium (11). In another study, osteoclasts were isolated and cultured for in vitro study of the effects of the Chinese herb Guizhou epimedium (Epimedium Leptorrhizum Stearn). Epimedium inhibited osteoclast-mediated bone resorption (144). Furthermore, curcumin (a mixture of curcuminoids) can prevent osteoclastogenesis, and it is also has anti-inflammatory and antioxidant effects (93). Studies of the effect of paeonol on RANKL-induced NF- $\kappa$ B activation and on osteoclastogenesis in osteoclast precursor cells have shown that paeonol inhibits RANKL-induced ERK, p38, and NF- $\kappa$ B activation in macrophages (7).

## **EFFECT OF STEM CELL THERAPY COMBINED WITH CHINESE HERBS ON OSTEOARTHRITIS**

Osteoarthritis is one of the most common chronic age-related disorders, which affects at least one-third of adults over years of age (36). Osteoarthritis is a progressive and multifactorial disease characterized by cartilage degeneration, osteophytes, and synovial fibrosis (49). Current treatments for osteoarthritis are unsatisfactory for restoring full function or returning tissue to its native normal state (122). Until recently, pharmacological management of most patients with osteoarthritis has targeted symptoms of the disease rather than the underlying cause and pathophysiology (19). Analgesics and non-steroidal anti-inflammatory drugs represent the mainstay of treatment. These drugs mainly alleviate symptoms but have many side effects (80). Chinese herbal remedies are the most prevalent and effective treatments for the management of chronic illnesses in many Asian countries. Because indigenous herbs have anti-inflammatory, analgesic, anti-arthritic, and blood microcirculation-enhancing functions and the ability to inhibit enzymatic cartilage degeneration, they have been widely used to treat various inflammatory diseases (59,99). A recent report showed that extracts from several TCM formulae alleviate osteoarthritis symptoms and are beneficial to the pathophysiology of the disease. For example, Diacerein, which is isolated from *Rheum* plants, is as effective as non-steroidal anti-inflammatory drugs for knee and hip osteoarthritis (43). *Clematis* is commonly known for its anti-spasmodic,

anti-inflammatory, and analgesic effects (59). SKI 306X, a processed extract from a formula of three TCM herbs (*Clematis manshurica*, *Trichosanthes kirilowii*, and *Prunella vulgaris*), has been used as a anti-arthritic agent (20). Yanghe decoction protects articular cartilage, an effect that is achieved by regulating the expression of HIF-1alpha mRNA (10).

mesenchymal stem cells (MSCs) are an attractive source for tissue engineering and regeneration owing to their ability to differentiate into multiple mesenchymal lineages, including bone, cartilage, muscle, ligament, tendon, adipose, and marrow stroma (92). MSCs have been suggested to play an important role in osteoarthritis treatment because this cell population is resistant to the degenerative changes in bone and cartilage in osteoarthritis (25). Several studies have demonstrated that MSCs can form cartilage *in vivo* and *in vitro* (92). The proliferative capacity of MSCs from patients with osteoarthritis is substantially reduced, but the osteogenic potential of MSCs is equivalent to or greater than that of control subjects (85). Herbal medicine may affect MSCs by promoting osteoblastic differentiation (50,70). TCMs contain agents that can contribute to the proliferation and differentiation of MSCs. Recently, great effort has been devoted to examining the effects of TCMs on cell protection for osteoarthritis treatment using cell culture systems and animal models. For example,

*Clematis* was shown to be protective in osteoarthritis by inhibiting apoptosis in chondrocytes. *Rheum* (Diacerein) and *Clematis* can improve the cell proliferative capacity (20,59,99) of MSCs. These results suggest that TCM may play an important therapeutic role in patients with osteoarthritis by improving the proliferative capacity of MSCs and inhibiting mineralization by MSCs (14).

## **CONCLUSION**

In conclusion, growing evidence indicates that Chinese herbs or herbal extracts can greatly impact stem cell biology. For example, 20070721-GX, a health food, contains a mixture of Chinese herb extract and is a potent stem cell growth factor. In the unpublished study, the CD34<sup>+</sup> cell numbers of eleven volunteers, after administer 20070721-GX for at least two months, are increased to 4.9 times ( $1.47 \pm 0.11\%$ ) than placebo-controlled ( $0.3 \pm 0.1\%$ ).

Chinese herbs may be able to efficiently regulate stem cell fate, proliferation, and differentiation. Many kinds of Chinese herbs have been shown to regulate stem cell activity. Most recent reports about the effects of Chinese herbs on stem cell regulation have focused on their effective treatment of various conditions. Few reports have examined the effects of Chinese herbs on stem cell regulation, however. Many scientists do not yet accept the stem cell regulation effects of Chinese herbs because of

the lack of molecular biology evidence. Thus, examination of the molecular mechanisms of Chinese herbs on stem cell regulation is the next challenge.

#### Acknowledgements

The work was supported by grants (No. 097426K9) from the Gwo Xi Stem Cell Applied Technology, Hsinchu, Taiwan, R.O.C. This study is supported in part by Taiwan Department of Health Clinical Trial and Research Center of Excellence (DOH99-TD-B-111-004).

Table 1. Herbs in traditional Chinese medicines are used by traditional Chinese medicine practitioners to treat neuronal diseases.

Category of use	Latin name (English equivalent)	Chinese name (pinyin)	Referenc(s)
Blood Circulation-Promoting and Stasis-Removing Drugs	<i>Radix Paeoniae Rubra</i>	Chi Shao	(30,129,130)
	<i>Radix Achyranthis Bidentatae</i>	Niu Xi	(130)
	<i>Radix Salviae Miltiorrhizae</i>	Dan Shen	(73,133)
	<i>Rhizoma Corydalis</i>	Yan Hu Suo	
	<i>Radix Notoginseng</i>	San Qi	(21,22)
	<i>Rhizoma Chuanxiong</i>	Chuan Xiong	(51)
	<i>Stigma Croci</i>	Xi Hong Hua	
Qi-Regulating Drugs	<i>Fructus Toosendan</i>	Chuan Lian Zi	(109,110)
	<i>Fructus Aurantii Immaturus</i>	Zhi Shi	
	<i>Rhizoma Cyperi</i>	Xiang Fu	(57)
Expectorants, Antitussives, and Anti-asthmatics	<i>Unprocessed Rhizoma Pinelliae</i>	Sheng Ban Xia	(24,55)
	<i>Ginkgo Leaf</i>	Yin Xing Ye Pian	(1,32,97)
	<i>Unprocessed Rhizome Arisaematis</i>	Sheng Tian Nan Xing	(54)
	<i>Fructus Perillae</i>	Zi Su Zi	(151)
Heat-Clearing Drugs	<i>Cortex Moutan</i>	Mu Dan Pi	
	<i>Radix Sophorae Flavescentis</i>	Ku Shen	
	<i>Herba Patriniae</i>	Bai Jiang Cao	
	<i>Herba Houttuyniae</i>	Yu Xing Cao	
	<i>Fructus Gardeniae</i>	Zhi Zi	(42,124)
	<i>Spica Prunellae</i>	Xia Ku Cao	
	<i>Rhizoma Coptidis</i>	Huang Lian	(26,45)
	<i>Radix Scutellariae</i>	Huang Qin	(45,47)
Awakening Drugs	<i>Borneolum Syntheticum</i>	Bing Pian	
	<i>Rhizoma Acori Tatarinowii</i>	Shi Chang Pu	
Tranquilizers	<i>Radix Polygalae</i>	Yuan Zhi	(23,58,145)
	<i>Semen Ziziphi Spinosa</i>	Suan Zao Ren	(102)
	<i>Cortex Albiziae</i>	He Huan Pi	
	<i>Succinum</i>	Hu Po	
Tonics	<i>Radix Ginseng</i>	Ren Shen	(21,22,111)
	<i>Radix Paeniae Alba</i>	Bai Shao	(7,72)
	<i>Radix Codonopsis</i>	Dang Shen	
	<i>Radix Glycyrrhizae</i>	Gan Cao	(17,145)

\* Please refer to lkum Web site (<http://www.lkum.com/pages/QJKojpmi.html>).

Table 2. Chinese herbs used in stem cell therapy for cardiovascular disease.

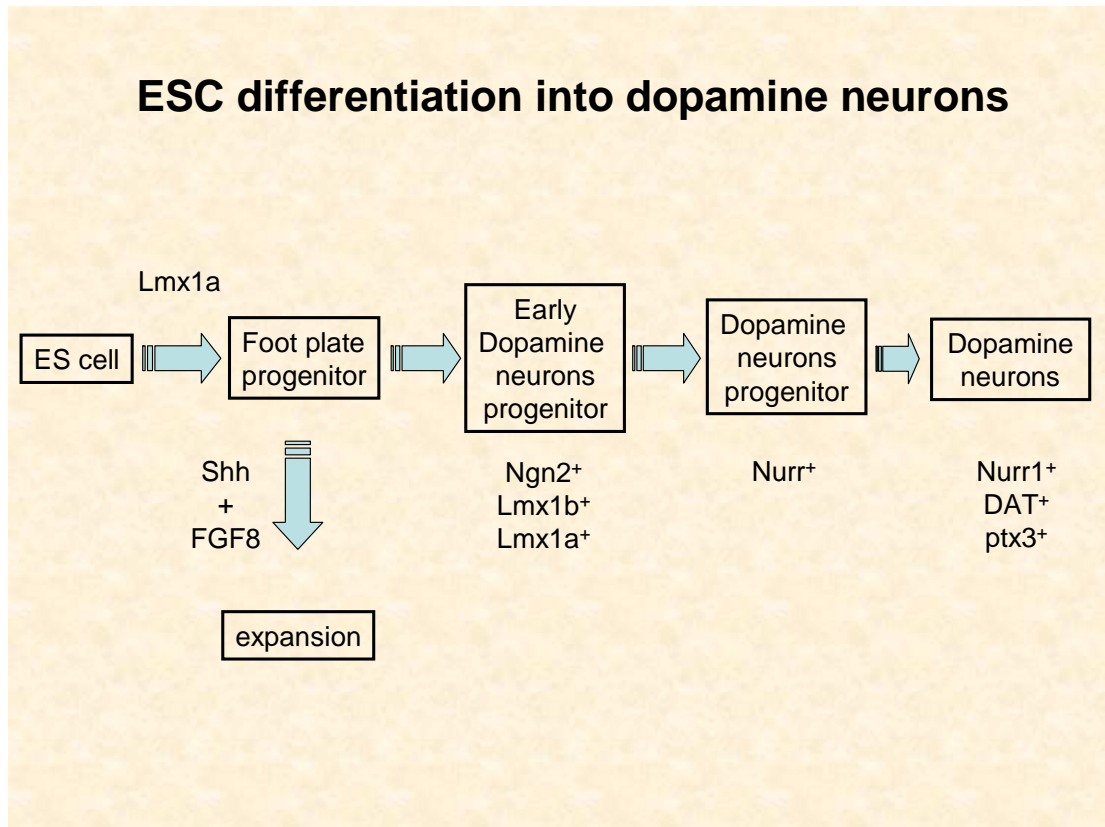
Disease	Therapeutic strategy	Chinese herb or compound extracted from Chinese herb	Reference
Myocardial infarction	Increase the cardiogenic differentiation ability of stem cells	<i>Geum Japonicum</i>	(18)
		Cardiogenin from <i>Geum Japonicum</i> <i>Panax ginseng</i>	(18) (140)
	Increase the migration ability of stem cells	<i>Panax ginseng</i>	(140)
		<i>Aconitum carmichaeli</i>	(140)
		<i>Ilex pubescens</i>	(140)
		<i>Leonuri herba</i>	(140)
	Increase angiogenesis	Saponin from <i>Panax quinquefolius</i>	(124,126)
Stenosis	Increase the migration ability of stem cells and reduce inflammation	<i>Panax ginseng</i>	(140)
		<i>Aconitum carmichaeli</i>	(140)
		<i>Ilex pubescens</i>	(140)
		<i>Leonuri herba</i>	(140)
	Regenerate functional endothelial cells	Ginsenoside Rg1 from <i>Panax ginseng</i>	(108)
		<i>Astragalus membranaceus</i>	(136)
		<i>Panax notoginseng</i>	(136)
		<i>Salvia miltiorrhiza</i> (Danshen)	(46)



Table 3. Traditional Chinese medicine applications in osteoporosis

<b>Function</b>	<b>Herb name</b>	<b>Effective composition</b>	<b>Reference(s)</b>
Osteoblastic differentiation	<i>Wedelia calendulacea</i>	Wedelolactone	(4)
	<i>Fructus Ligustri Lucidi</i>	Ethanol extract	(148)
	<i>Cistanche salsa</i>	(R)-HDOA	(134)
	<i>Eucommia ulmoides</i>	Cortex extract	(27)
	<i>Acanthopanax senticosus</i>	Water extract	(44)
	<i>Cuscuta chinensis</i>	Flavonoids	(137)
	<i>Glycine max</i>	Menaquinone-7	(42)
	<i>Green tea</i>	Epigallocatechin-3-gallate	(56)
Inhibits bone resorption activity	<i>Paeonia lactiflora Pallas</i>	Paeonol	(120)
	<i>Curcuma longa Linn</i>	curcumin	(90)
	<i>Guizhou epimedium</i>	Epimedium extract	(144)
	<i>Epimedium</i>	Icariin	(12)

Figure 1. Differentiation of embryonic stem cells (ESCs) into dopamine neurons and the genes involved.



## References

1. Ahlemeyer, B.; Kriegelstein, J. Neuroprotective effects of Ginkgo biloba extract. *Cell Mol Life Sci* 60(9):1779-1792; 2003.
2. Akiyama, Y.; Honmou, O.; Kato, T.; Uede, T.; Hashi, K.; Kocsis, J. D. Transplantation of clonal neural precursor cells derived from adult human brain establishes functional peripheral myelin in the rat spinal cord. *Exp Neurol* 167(1):27-39; 2001.
3. Alsberg, E.; von Recum, H. A.; Mahoney, M. J. Environmental cues to guide stem cell fate decision for tissue engineering applications. *Expert Opin Biol Ther* 6(9):847-866; 2006.
4. Annie, S.; Prabhu, R. G.; Malini, S. Activity of *Wedelia calendulacea* Less. in post-menopausal osteoporosis. *Phytomedicine* 13(1-2):43-48; 2006.
5. Bournival, J.; Quessy, P.; Martinoli, M. G. Protective Effects of Resveratrol and Quercetin Against MPP(+)-Induced Oxidative Stress Act by Modulating Markers of Apoptotic Death in Dopaminergic Neurons. *Cell Mol Neurobiol*; 2009.
6. Brazelton, T. R.; Rossi, F. M.; Keshet, G. I.; Blau, H. M. From marrow to brain: expression of neuronal phenotypes in adult mice. *Science* 290(5497):1775-1779; 2000.
7. Cao, B. Y.; Yang, Y. P.; Luo, W. F.; Mao, C. J.; Han, R.; Sun, X.; Cheng, J.; Liu, C. F. Paeoniflorin, a potent natural compound, protects PC12 cells from MPP(+) and acidic damage via autophagic pathway. *J Ethnopharmacol*.
8. Cao, F.; Sun, S.; Tong, E. T. Experimental study on inhibition of neuronal toxic effect of levodopa by ginkgo biloba extract on Parkinson disease in rats. *J Huazhong Univ Sci Technolog Med Sci* 23(2):151-153; 2003.
9. Chalmers-Redman, R. M.; Priestley, T.; Kemp, J. A.; Fine, A. In vitro propagation and inducible differentiation of multipotential progenitor cells from human fetal brain. *Neuroscience* 76(4):1121-1128; 1997.
10. Chen, C. W.; Chen, Y. Q. [Experiment research on the influence of Yanghe decoction on the expression of HIF-1 $\alpha$  mRNA in osteoarthritis]. *Zhongguo Gu Shang* 21(6):432-434; 2008.
11. Chen, K. M.; Ge, B. F.; Liu, X. Y.; Ma, P. H.; Lu, M. B.; Bai, M. H.; Wang, Y. Icariin inhibits the osteoclast formation induced by RANKL and macrophage-colony stimulating factor in mouse bone marrow culture. *Pharmazie* 62(5):388-391; 2007.
12. Chen, K. M.; Ge, B. F.; Ma, H. P.; Liu, X. Y.; Bai, M. H.; Wang, Y. Icariin, a flavonoid from the herb *Epimedium* enhances the osteogenic differentiation of rat primary bone marrow stromal cells. *Pharmazie* 60(12):939-942; 2005.

13. Chen, L. W.; Wang, Y. Q.; Wei, L. C.; Shi, M.; Chan, Y. S. Chinese herbs and herbal extracts for neuroprotection of dopaminergic neurons and potential therapeutic treatment of Parkinson's disease. *CNS Neurol Disord Drug Targets* 6(4):273-281; 2007.
14. Chen, M.; Feng, W.; Cao, H.; Zou, L.; Chen, C.; Baatrup, A.; Nielsen, A. B.; Li, H.; Kassem, M.; Zou, X. and others. A traditional Chinese medicine formula extracts stimulate proliferation and inhibit mineralization of human mesenchymal stem cells in vitro. *J Ethnopharmacol*; 2009.
15. Chen, X. C.; Zhu, Y. G.; Zhu, L. A.; Huang, C.; Chen, Y.; Chen, L. M.; Fang, F.; Zhou, Y. C.; Zhao, C. H. Ginsenoside Rg1 attenuates dopamine-induced apoptosis in PC12 cells by suppressing oxidative stress. *Eur J Pharmacol* 473(1):1-7; 2003.
16. Chen, Y. L.; Jian, M. H.; Lin, C. C.; Kang, J. C.; Chen, S. P.; Lin, P. C.; Hung, P. J.; Chen, J. R.; Chang, W. L.; Lin, S. Z. and others. The induction of orphan nuclear receptor Nur77 expression by n-butylphenylthalide as pharmaceuticals on hepatocellular carcinoma cell therapy. *Mol Pharmacol* 74(4):1046-1058; 2008.
17. Chen, Z. A.; Wang, J. L.; Liu, R. T.; Ren, J. P.; Wen, L. Q.; Chen, X. J.; Bian, G. X. Liquiritin potentiate neurite outgrowth induced by nerve growth factor in PC12 cells. *Cytotechnology* 60(1-3):125-132; 2009.
18. Cheng, L.; Chen, H.; Yao, X.; Qi, G.; Liu, H.; Lee, K.; Lee, K.; Zhang, J.; Chen, S.; Lin, X. and others. A plant-derived remedy for repair of infarcted heart. *PloS one* 4(2):e4461; 2009.
19. Cho, C. H.; Nuttall, M. E. Emerging techniques for the discovery and validation of therapeutic targets for skeletal diseases. *Expert Opin Ther Targets* 6(6):679-689; 2002.
20. Choi, J. H.; Kim, D. Y.; Yoon, J. H.; Youn, H. Y.; Yi, J. B.; Rhee, H. I.; Ryu, K. H.; Jung, K.; Han, C. K.; Kwak, W. J. and others. Effects of SKI 306X, a new herbal agent, on proteoglycan degradation in cartilage explant culture and collagenase-induced rabbit osteoarthritis model. *Osteoarthritis Cartilage* 10(6):471-478; 2002.
21. Choi, R. C.; Zhu, J. T.; Leung, K. W.; Chu, G. K.; Xie, H. Q.; Chen, V. P.; Zheng, K. Y.; Lau, D. T.; Dong, T. T.; Chow, P. C. and others. A flavonol glycoside, isolated from roots of *Panax notoginseng*, reduces amyloid-beta-induced neurotoxicity in cultured neurons: signaling transduction and drug development for Alzheimer's disease. *J Alzheimers Dis* 19(3):795-811.
22. Choi, R. C.; Zhu, J. T.; Leung, K. W.; Chu, G. K.; Xie, H. Q.; Chen, V. P.;

- Zheng, K. Y.; Lau, D. T.; Dong, T. T.; Chow, P. C. and others. A Flavonol Glycoside, Isolated From Roots of *Panax notoginseng*, Reduces Amyloid-beta-Induced Neurotoxicity in Cultured Neurons: Signaling Transduction and Drug Development for Alzheimer's Disease. *J Alzheimers Dis*; 2009.
23. Chung, I. W.; Moore, N. A.; Oh, W. K.; O'Neill, M. F.; Ahn, J. S.; Park, J. B.; Kang, U. G.; Kim, Y. S. Behavioural pharmacology of polygalasaponins indicates potential antipsychotic efficacy. *Pharmacol Biochem Behav* 71(1-2):191-195; 2002.
  24. Chung, T. W.; Koo, B. S.; Choi, E. G.; Kim, M. G.; Lee, I. S.; Kim, C. H. Neuroprotective effect of a chuk-me-sun-dan on neurons from ischemic damage and neuronal cell toxicity. *Neurochem Res* 31(1):1-9; 2006.
  25. Creamer, P.; Hochberg, M. C. Osteoarthritis. *Lancet* 350(9076):503-508; 1997.
  26. Cui, H. S.; Matsumoto, K.; Murakami, Y.; Hori, H.; Zhao, Q.; Obi, R. Berberine exerts neuroprotective actions against in vitro ischemia-induced neuronal cell damage in organotypic hippocampal slice cultures: involvement of B-cell lymphoma 2 phosphorylation suppression. *Biol Pharm Bull* 32(1):79-85; 2009.
  27. Deyama, T.; Nishibe, S.; Nakazawa, Y. Constituents and pharmacological effects of *Eucommia* and Siberian ginseng. *Acta Pharmacol Sin* 22(12):1057-1070; 2001.
  28. Ding, D. C.; Shyu, W. C.; Chiang, M. F.; Lin, S. Z.; Chang, Y. C.; Wang, H. J.; Su, C. Y.; Li, H. Enhancement of neuroplasticity through upregulation of beta1-integrin in human umbilical cord-derived stromal cell implanted stroke model. *Neurobiology of disease* 27(3):339-353; 2007.
  29. Dong, L. H.; Wang, Y.; Lu, C. Q.; Wang, F. [Effect of *Astragalus mongholicus* on inducing differentiations of rat bone marrow-derived mesenchymal stem cells into neurocyte-like cells]. *Sichuan Da Xue Xue Bao Yi Xue Ban* 38(3):417-420; 2007.
  30. Dong, X. P.; Xu, T. L. *Radix paeoniae rubra* suppression of sodium current in acutely dissociated rat hippocampal CA1 neurons. *Brain Res* 940(1-2):1-9; 2002.
  31. Dong, X. X.; Liu, J. B.; Dong, Y. X. [Experimental study on effect of gastrodia in inducing the differentiation of mesenchymal stem cells into neuron-like cells]. *Zhongguo Zhong Xi Yi Jie He Za Zhi* 24(1):51-54; 2004.
  32. Fan, L. H.; Wang, K. Z.; Cheng, B. [Protective effects of ginkgo leaf extracts on neurons in spinal cord after ischemia-reperfusion injury in rabbits]. *Zhong*

- Xi Yi Jie He Xue Bao 4(2):181-184; 2006.
33. Flax, J. D.; Aurora, S.; Yang, C.; Simonin, C.; Wills, A. M.; Billingham, L. L.; Jendoubi, M.; Sidman, R. L.; Wolfe, J. H.; Kim, S. U. and others. Engraftable human neural stem cells respond to developmental cues, replace neurons, and express foreign genes. *Nat Biotechnol* 16(11):1033-1039; 1998.
  34. Forte, A.; Finicelli, M.; Mattia, M.; Berrino, L.; Rossi, F.; De Feo, M.; Cotrufo, M.; Cipollaro, M.; Cascino, A.; Galderisi, U. Mesenchymal stem cells effectively reduce surgically induced stenosis in rat carotids. *Journal of cellular physiology* 217(3):789-799; 2008.
  35. Gage, F. H.; Coates, P. W.; Palmer, T. D.; Kuhn, H. G.; Fisher, L. J.; Suhonen, J. O.; Peterson, D. A.; Suhr, S. T.; Ray, J. Survival and differentiation of adult neuronal progenitor cells transplanted to the adult brain. *Proc Natl Acad Sci U S A* 92(25):11879-11883; 1995.
  36. Garstang, S. V.; Stitik, T. P. Osteoarthritis: epidemiology, risk factors, and pathophysiology. *Am J Phys Med Rehabil* 85(11 Suppl):S2-11; quiz S12-14; 2006.
  37. Goltzman, D. Discoveries, drugs and skeletal disorders. *Nat Rev Drug Discov* 1(10):784-796; 2002.
  38. Goolsby, J.; Marty, M. C.; Heletz, D.; Chiappelli, J.; Tashko, G.; Yarnell, D.; Fishman, P. S.; Dhib-Jalbut, S.; Bever, C. T., Jr.; Pessac, B. and others. Hematopoietic progenitors express neural genes. *Proc Natl Acad Sci U S A* 100(25):14926-14931; 2003.
  39. Guo, S.; Bezar, E.; Zhao, B. Protective effect of green tea polyphenols on the SH-SY5Y cells against 6-OHDA induced apoptosis through ROS-NO pathway. *Free Radic Biol Med* 39(5):682-695; 2005.
  40. Guo, S.; Yan, J.; Yang, T.; Yang, X.; Bezar, E.; Zhao, B. Protective effects of green tea polyphenols in the 6-OHDA rat model of Parkinson's disease through inhibition of ROS-NO pathway. *Biol Psychiatry* 62(12):1353-1362; 2007.
  41. Hall, P. A.; Watt, F. M. Stem cells: the generation and maintenance of cellular diversity. *Development* 106(4):619-633; 1989.
  42. Han, S. H.; Chung, M. J.; Lee, S. J.; Rhee, C. Digestion-resistant fraction from soybean [*Glycine max* (L.) Merrill] induces hepatic LDL receptor and CYP7A1 expression in apolipoprotein E-deficient mice. *J Nutr Biochem* 17(10):682-688; 2006.
  43. Hunter, D. J.; Wise, B. Review: diacerein is more effective than placebo and is as effective as NSAIDs for knee and hip osteoarthritis. *Evid Based Med* 12(3):74; 2007.

44. Hwang, Y. C.; Jeong, I. K.; Ahn, K. J.; Chung, H. Y. The effects of *Acanthopanax senticosus* extract on bone turnover and bone mineral density in Korean postmenopausal women. *J Bone Miner Metab*; 2009.
45. Hwang, Y. S.; Shin, C. Y.; Huh, Y.; Ryu, J. H. Hwangryun-Hae-Dok-tang (Huanglian-Jie-Du-Tang) extract and its constituents reduce ischemia-reperfusion brain injury and neutrophil infiltration in rats. *Life Sci* 71(18):2105-2117; 2002.
46. Ji, K. T.; Zhang, H. Q.; Tang, J. F.; Li, H. Y. [Effects of Danshen on number and activity of endothelial progenitor cells of patients with hypercholesterolemia]. *Zhongguo Zhong yao za zhi = Zhongguo zhongyao zazhi = China journal of Chinese materia medica* 32(12):1214-1217; 2007.
47. Jia, Y.; Yang, Y.; Zhou, Y.; Song, Y.; Liu, L.; Song, J.; Wang, X.; Zhong, L.; Yu, X. [Differentiation of rat bone marrow stromal cells into neuron induced by baicalin]. *Zhonghua Yi Xue Za Zhi* 82(19):1337-1341; 2002.
48. Jingyi, W.; Yasuhiro, M.; Naoya, H.; Seok, R. C.; Yoshiharu, Y.; Nagara, T.; Fumiko, T.; Shigeru, M.; Junji, K. Observation on the effects of Chinese medicine zhenxuanyin for improving cerebral blood flow in rats with cerebral ischemia. *J Tradit Chin Med* 17(4):299-303; 1997.
49. Johnston, S. A. Osteoarthritis. Joint anatomy, physiology, and pathobiology. *Vet Clin North Am Small Anim Pract* 27(4):699-723; 1997.
50. Kang, S. K.; Kim, K. S.; Byun, Y. S.; Suh, S. J.; Jim, U. H.; Kim, K. H.; Lee, I. S.; Kim, C. H. Effects of *Ulmus davidiana* planch on mineralization, bone morphogenetic protein-2, alkaline phosphatase, type I collagen, and collagenase-1 in bone cells. *In Vitro Cell Dev Biol Anim* 42(7):225-229; 2006.
51. Kao, T. K.; Ou, Y. C.; Kuo, J. S.; Chen, W. Y.; Liao, S. L.; Wu, C. W.; Chen, C. J.; Ling, N. N.; Zhang, Y. H.; Peng, W. H. Neuroprotection by tetramethylpyrazine against ischemic brain injury in rats. *Neurochem Int* 48(3):166-176; 2006.
52. Kim, D. S.; Kim, J. Y.; Kang, M.; Cho, M. S.; Kim, D. W. Derivation of functional dopamine neurons from embryonic stem cells. *Cell transplantation* 16(2):117-123; 2007.
53. Kim, E. H.; Jang, M. H.; Shin, M. C.; Shin, M. S.; Kim, C. J. Protective effect of aqueous extract of *Ginseng radix* against 1-methyl-4-phenylpyridinium-induced apoptosis in PC12 cells. *Biol Pharm Bull* 26(12):1668-1673; 2003.
54. Kim, H. Neuroprotective herbs for stroke therapy in traditional eastern medicine. *Neurol Res* 27(3):287-301; 2005.

55. Koo, B. S.; Choi, E. G.; Park, J. B.; Cho, C. H.; Chung, K. H.; Kim, C. H. Neuroprotective effect of Chuk-Me-Sun-Dan on NMDA- and AMPA-evoked nitric oxide synthase activity in mouse brain. *Immunopharmacol Immunotoxicol* 27(3):499-514; 2005.
56. Lee, A. H.; Fraser, M. L.; Binns, C. W. Possible role for green tea in ovarian cancer prevention. *Future Oncol* 1(6):771-777; 2005.
57. Lee, C. H.; Hwang, D. S.; Kim, H. G.; Oh, H.; Park, H.; Cho, J. H.; Lee, J. M.; Jang, J. B.; Lee, K. S.; Oh, M. S. Protective effect of Cyperi rhizoma against 6-hydroxydopamine-induced neuronal damage. *J Med Food* 13(3):564-571.
58. Lee, H. J.; Ban, J. Y.; Koh, S. B.; Seong, N. S.; Song, K. S.; Bae, K. W.; Seong, Y. H. Polygalae radix extract protects cultured rat granule cells against damage induced by NMDA. *Am J Chin Med* 32(4):599-610; 2004.
59. Lee, S. W.; Chung, W. T.; Choi, S. M.; Kim, K. T.; Yoo, K. S.; Yoo, Y. H. Clematis mandshurica protected to apoptosis of rat chondrocytes. *J Ethnopharmacol* 101(1-3):294-298; 2005.
60. Leferovich, J. M.; Bedelbaeva, K.; Samulewicz, S.; Zhang, X. M.; Zwas, D.; Lankford, E. B.; Heber-Katz, E. Heart regeneration in adult MRL mice. *Proceedings of the National Academy of Sciences of the United States of America* 98(17):9830-9835; 2001.
61. Lei, Z.; Yongda, L.; Jun, M.; Yingyu, S.; Shaoju, Z.; Xinwen, Z.; Mingxue, Z. Culture and neural differentiation of rat bone marrow mesenchymal stem cells in vitro. *Cell Biol Int* 31(9):916-923; 2007.
62. Levites, Y.; Weinreb, O.; Maor, G.; Youdim, M. B.; Mandel, S. Green tea polyphenol (-)-epigallocatechin-3-gallate prevents N-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced dopaminergic neurodegeneration. *J Neurochem* 78(5):1073-1082; 2001.
63. Levites, Y.; Youdim, M. B.; Maor, G.; Mandel, S. Attenuation of 6-hydroxydopamine (6-OHDA)-induced nuclear factor-kappaB (NF-kappaB) activation and cell death by tea extracts in neuronal cultures. *Biochem Pharmacol* 63(1):21-29; 2002.
64. Li, F. Q.; Cheng, X. X.; Liang, X. B.; Wang, X. H.; Xue, B.; He, Q. H.; Wang, X. M.; Han, J. S. Neurotrophic and neuroprotective effects of triphlorolide, an extract of Chinese herb *Tripterygium wilfordii* Hook F, on dopaminergic neurons. *Exp Neurol* 179(1):28-37; 2003.
65. Li, F. Q.; Lu, X. Z.; Liang, X. B.; Zhou, H. F.; Xue, B.; Liu, X. Y.; Niu, D. B.; Han, J. S.; Wang, X. M. Triptolide, a Chinese herbal extract, protects dopaminergic neurons from inflammation-mediated damage through inhibition of microglial activation. *J Neuroimmunol* 148(1-2):24-31; 2004.



66. Li, J. X.; Wang, Z. B.; Zhu, L. Q.; Zhu, F.; Cui, W. [Cell proliferation inhibitive and apoptosis promoting effects of sanchi extract on GES-1 cell after being transformed by MNNG]. *Zhongguo Zhong Xi Yi Jie He Za Zhi* 25(8):719-722; 2005.
67. Li, L. [Protective effects of schisanhenol, salvianolic acid A and SY-L on oxidative stress induced injuries of cerebral cells and their mechanisms]. *Sheng Li Ke Xue Jin Zhan* 29(1):35-38; 1998.
68. Li, M.; Yu, C. M.; Cheng, L.; Wang, M.; Gu, X.; Lee, K. H.; Wang, T.; Sung, Y. T.; Sanderson, J. E. Repair of infarcted myocardium by an extract of *Geum japonicum* with dual effects on angiogenesis and myogenesis. *Clinical chemistry* 52(8):1460-1468; 2006.
69. Li, R.; Huang, Y. G.; Fang, D.; Le, W. D. (-)-Epigallocatechin gallate inhibits lipopolysaccharide-induced microglial activation and protects against inflammation-mediated dopaminergic neuronal injury. *J Neurosci Res* 78(5):723-731; 2004.
70. Liang, B.; Song, X.; Liu, G.; Li, R.; Xie, J.; Xiao, L.; Du, M.; Zhang, Q.; Xu, X.; Gan, X. and others. Involvement of TR3/Nur77 translocation to the endoplasmic reticulum in ER stress-induced apoptosis. *Exp Cell Res* 313(13):2833-2844; 2007.
71. Lin, P. C.; Chen, Y. L.; Chiu, S. C.; Yu, Y. L.; Chen, S. P.; Chien, M. H.; Chen, K. Y.; Chang, W. L.; Lin, S. Z.; Chiou, T. W. and others. Orphan nuclear receptor, Nurr-77 was a possible target gene of butyridenepthalide chemotherapy on glioblastoma multiform brain tumor. *J Neurochem* 106(3):1017-1026; 2008.
72. Liu, H. Q.; Zhang, W. Y.; Luo, X. T.; Ye, Y.; Zhu, X. Z. Paeoniflorin attenuates neuroinflammation and dopaminergic neurodegeneration in the MPTP model of Parkinson's disease by activation of adenosine A1 receptor. *Br J Pharmacol* 148(3):314-325; 2006.
73. Liu, J.; Kuang, P.; Wu, W.; Zhang, F.; Wan, F.; Huang, Y.; Ding, A. Radix *Salviae miltiorrhizae* protects rat hippocampal neuron in culture from anoxic damage. *J Tradit Chin Med* 18(1):49-54; 1998.
74. Liu, J. W.; Tian, S. J.; de Barry, J.; Luu, B. Panaxadiol glycosides that induce neuronal differentiation in neurosphere stem cells. *J Nat Prod* 70(8):1329-1334; 2007.
75. Liu, Y. R.; Qu, S. X.; Maitz, M. F.; Tan, R.; Weng, J. The effect of the major components of *Salvia Miltiorrhiza* Bunge on bone marrow cells. *J Ethnopharmacol* 111(3):573-583; 2007.
76. Lois, C.; Alvarez-Buylla, A. Proliferating subventricular zone cells in the adult

- mammalian forebrain can differentiate into neurons and glia. *Proc Natl Acad Sci U S A* 90(5):2074-2077; 1993.
77. Ma, L.; Cui, B. L.; Feng, X. Y.; Law, F. D.; Jiang, X. W.; Yang, L. Y.; Xie, Q. D.; Huang, T. H. [Biological characteristics of human umbilical cord-derived mesenchymal stem cells and their differentiation into neurocyte-like cells]. *Zhonghua Er Ke Za Zhi* 44(7):513-517; 2006.
  78. Ma, L.; Feng, X. Y.; Cui, B. L.; Law, F.; Jiang, X. W.; Yang, L. Y.; Xie, Q. D.; Huang, T. H. Human umbilical cord Wharton's Jelly-derived mesenchymal stem cells differentiation into nerve-like cells. *Chin Med J (Engl)* 118(23):1987-1993; 2005.
  79. Ma, L.; Liu, W. K.; Zhang, Y. K.; Ju, Y. [Impacts of tetramethylpyrazine on BDNF, bFGF expression and neuron-protection in severe brain injury tissue of rat]. *Sichuan Da Xue Xue Bao Yi Xue Ban* 39(2):207-210; 2008.
  80. McKellar, G.; Madhok, R.; Singh, G. The problem with NSAIDs: what data to believe? *Curr Pain Headache Rep* 11(6):423-427; 2007.
  81. Mizumaki, Y.; Kurimoto, M.; Hirashima, Y.; Nishijima, M.; Kamiyama, H.; Nagai, S.; Takaku, A.; Sugihara, K.; Shimizu, M.; Endo, S. Lipophilic fraction of *Panax ginseng* induces neuronal differentiation of PC12 cells and promotes neuronal survival of rat cortical neurons by protein kinase C dependent manner. *Brain Res* 950(1-2):254-260; 2002.
  82. Mohri, T.; Chiba, K.; Yamazaki, M.; Shimizu, M.; Morita, N. Activation of PC12 cells by lipophilic components of *Panax ginseng*. *Planta Med* 58(4):321-323; 1992.
  83. Morshead, C. M.; Reynolds, B. A.; Craig, C. G.; McBurney, M. W.; Staines, W. A.; Morassutti, D.; Weiss, S.; van der Kooy, D. Neural stem cells in the adult mammalian forebrain: a relatively quiescent subpopulation of subependymal cells. *Neuron* 13(5):1071-1082; 1994.
  84. Moyer, M. P.; Johnson, R. A.; Zompa, E. A.; Cain, L.; Morshed, T.; Hulsebosch, C. E. Culture, expansion, and transplantation of human fetal neural progenitor cells. *Transplant Proc* 29(4):2040-2041; 1997.
  85. Murphy, J. M.; Dixon, K.; Beck, S.; Fabian, D.; Feldman, A.; Barry, F. Reduced chondrogenic and adipogenic activity of mesenchymal stem cells from patients with advanced osteoarthritis. *Arthritis Rheum* 46(3):704-713; 2002.
  86. Murry, C. E.; Keller, G. Differentiation of embryonic stem cells to clinically relevant populations: lessons from embryonic development. *Cell* 132(4):661-680; 2008.
  87. Nandoe Tewarie, R. D.; Hurtado, A.; Levi, A. D.; Grotenhuis, J. A.; Oudega,

- M. Bone marrow stromal cells for repair of the spinal cord: towards clinical application. *Cell transplantation* 15(7):563-577; 2006.
88. Pan, T.; Fei, J.; Zhou, X.; Jankovic, J.; Le, W. Effects of green tea polyphenols on dopamine uptake and on MPP<sup>+</sup>-induced dopamine neuron injury. *Life Sci* 72(9):1073-1083; 2003.
  89. Poss, K. D.; Wilson, L. G.; Keating, M. T. Heart regeneration in zebrafish. *Science (New York, N.Y)* 298(5601):2188-2190; 2002.
  90. Prasad, C. P.; Rath, G.; Mathur, S.; Bhatnagar, D.; Ralhan, R. Potent growth suppressive activity of curcumin in human breast cancer cells: modulation of Wnt/beta-catenin signaling. *Chem Biol Interact*; 2009.
  91. Priller, J.; Persons, D. A.; Klett, F. F.; Kempermann, G.; Kreutzberg, G. W.; Dirnagl, U. Neogenesis of cerebellar Purkinje neurons from gene-marked bone marrow cells in vivo. *J Cell Biol* 155(5):733-738; 2001.
  92. Prockop, D. J. Marrow stromal cells as stem cells for nonhematopoietic tissues. *Science* 276(5309):71-74; 1997.
  93. Putnam, S. E.; Scutt, A. M.; Bicknell, K.; Priestley, C. M.; Williamson, E. M. Natural products as alternative treatments for metabolic bone disorders and for maintenance of bone health. *Phytother Res* 21(2):99-112; 2007.
  94. Radad, K.; Gille, G.; Moldzio, R.; Saito, H.; Ishige, K.; Rausch, W. D. Ginsenosides Rb1 and Rg1 effects on survival and neurite growth of MPP<sup>+</sup>-affected mesencephalic dopaminergic cells. *J Neural Transm* 111(1):37-45; 2004.
  95. Radad, K.; Gille, G.; Moldzio, R.; Saito, H.; Rausch, W. D. Ginsenosides Rb1 and Rg1 effects on mesencephalic dopaminergic cells stressed with glutamate. *Brain Res* 1021(1):41-53; 2004.
  96. Ramassamy, C.; Clostre, F.; Christen, Y.; Costentin, J. Prevention by a Ginkgo biloba extract (GBE 761) of the dopaminergic neurotoxicity of MPTP. *J Pharm Pharmacol* 42(11):785-789; 1990.
  97. Ramassamy, C.; Longpre, F.; Christen, Y. Ginkgo biloba extract (EGb 761) in Alzheimer's disease: is there any evidence? *Curr Alzheimer Res* 4(3):253-262; 2007.
  98. Reynolds, B. A.; Weiss, S. Generation of neurons and astrocytes from isolated cells of the adult mammalian central nervous system. *Science* 255(5052):1707-1710; 1992.
  99. Rezende, M. U.; Gurgel, H. M.; Vilaca Junior, P. R.; Kuroba, R. K.; Lopes, A. S.; Phillipi, R. Z.; Hernandez, A. J. Diacerhein versus glucosamine in a rat model of osteoarthritis. *Clinics (Sao Paulo)* 61(5):461-466; 2006.
  100. Rojas, P.; Garduno, B.; Rojas, C.; Viguera, R. M.; Rojas-Castaneda, J.; Rios,

- C.; Serrano-Garcia, N. EGb761 blocks MPP<sup>+</sup>-induced lipid peroxidation in mouse corpus striatum. *Neurochem Res* 26(11):1245-1251; 2001.
101. Rojas, P.; Serrano-Garcia, N.; Mares-Samano, J. J.; Medina-Campos, O. N.; Pedraza-Chaverri, J.; Ogren, S. O. EGb761 protects against nigrostriatal dopaminergic neurotoxicity in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine-induced Parkinsonism in mice: role of oxidative stress. *Eur J Neurosci* 28(1):41-50; 2008.
  102. Rong, C. L.; Dai, Y. X.; Cui, Y. [Effects of Semen Ziziphi Spinosae on the anxiety behavior of the yin deficiency mice]. *Zhong Yao Cai* 31(11):1703-1705; 2008.
  103. Roybon, L.; Ma, Z.; Asztely, F.; Fosum, A.; Jacobsen, S. E.; Brundin, P.; Li, J. Y. Failure of transdifferentiation of adult hematopoietic stem cells into neurons. *Stem Cells* 24(6):1594-1604; 2006.
  104. Rudakewich, M.; Ba, F.; Benishin, C. G. Neurotrophic and neuroprotective actions of ginsenosides Rb(1) and Rg(1). *Planta Med* 67(6):533-537; 2001.
  105. Ryu, M. Y.; Lee, M. A.; Ahn, Y. H.; Kim, K. S.; Yoon, S. H.; Snyder, E. Y.; Cho, K. G.; Kim, S. U. Brain transplantation of neural stem cells cotransduced with tyrosine hydroxylase and GTP cyclohydrolase 1 in Parkinsonian rats. *Cell transplantation* 14(4):193-202; 2005.
  106. Sasaki, T.; Oh, K. B.; Matsuoka, H.; Saito, M. [Effect of Panax ginseng components on the differentiation of mouse embryonic stem cells into cardiac-like cells]. *Yakugaku Zasshi* 128(3):461-467; 2008.
  107. Sato, N.; Sanjuan, I. M.; Heke, M.; Uchida, M.; Naef, F.; Brivanlou, A. H. Molecular signature of human embryonic stem cells and its comparison with the mouse. *Dev Biol* 260(2):404-413; 2003.
  108. Shi, A. W.; Wang, X. B.; Lu, F. X.; Zhu, M. M.; Kong, X. Q.; Cao, K. J. Ginsenoside Rg1 promotes endothelial progenitor cell migration and proliferation. *Acta pharmacologica Sinica* 30(3):299-306; 2009.
  109. Shi, Q.; Hao, Q.; Bouissac, J.; Lu, Y.; Tian, S.; Luu, B. Ginsenoside-Rd from Panax notoginseng enhances astrocyte differentiation from neural stem cells. *Life Sci* 76(9):983-995; 2005.
  110. Shi, Y. L.; Wang, W. P. [Biological effects of toosendanin, an active ingredient of herbal vermifuge in Chinese traditional medicine]. *Sheng Li Xue Bao* 58(5):397-406; 2006.
  111. Shin, J. Y.; Song, J. Y.; Yun, Y. S.; Yang, H. O.; Rhee, D. K.; Pyo, S. Immunostimulating effects of acidic polysaccharides extract of Panax ginseng on macrophage function. *Immunopharmacol Immunotoxicol* 24(3):469-482; 2002.

112. Shyu, W. C.; Chen, C. P.; Lin, S. Z.; Lee, Y. J.; Li, H. Efficient tracking of non-iron-labeled mesenchymal stem cells with serial MRI in chronic stroke rats. *Stroke; a journal of cerebral circulation* 38(2):367-374; 2007.
113. Shyu, W. C.; Li, K. W.; Peng, H. F.; Lin, S. Z.; Liu, R. S.; Wang, H. J.; Su, C. Y.; Lee, Y. J.; Li, H. Induction of GAP-43 modulates neuroplasticity in PBSC (CD34+) implanted-Parkinson's model. *Journal of neuroscience research* 87(9):2020-2033; 2009.
114. Shyu, W. C.; Lin, S. Z.; Chiang, M. F.; Su, C. Y.; Li, H. Intracerebral peripheral blood stem cell (CD34+) implantation induces neuroplasticity by enhancing beta1 integrin-mediated angiogenesis in chronic stroke rats. *J Neurosci* 26(13):3444-3453; 2006.
115. Sloley, B. D.; Urichuk, L. J.; Morley, P.; Durkin, J.; Shan, J. J.; Pang, P. K.; Coutts, R. T. Identification of kaempferol as a monoamine oxidase inhibitor and potential Neuroprotectant in extracts of Ginkgo biloba leaves. *J Pharm Pharmacol* 52(4):451-459; 2000.
116. Svendsen, C. N.; Caldwell, M. A.; Shen, J.; ter Borg, M. G.; Rosser, A. E.; Tyers, P.; Karmiol, S.; Dunnett, S. B. Long-term survival of human central nervous system progenitor cells transplanted into a rat model of Parkinson's disease. *Exp Neurol* 148(1):135-146; 1997.
117. Szasz, B. K.; Lenkey, N.; Barth, A. M.; Mike, A.; Somogyvari, Z.; Farkas, O.; Lendvai, B. Converging effects of Ginkgo biloba extract at the level of transmitter release, NMDA and sodium currents and dendritic spikes. *Planta Med* 74(10):1235-1239; 2008.
118. Tohda, C.; Tamura, T.; Matsuyama, S.; Komatsu, K. Promotion of axonal maturation and prevention of memory loss in mice by extracts of Astragalus mongholicus. *Br J Pharmacol* 149(5):532-541; 2006.
119. Torrente, Y.; Polli, E. Mesenchymal stem cell transplantation for neurodegenerative diseases. *Cell transplantation* 17(10-11):1103-1113; 2008.
120. Tsai, H. Y.; Lin, H. Y.; Fong, Y. C.; Wu, J. B.; Chen, Y. F.; Tsuzuki, M.; Tang, C. H. Paeonol inhibits RANKL-induced osteoclastogenesis by inhibiting ERK, p38 and NF-kappaB pathway. *Eur J Pharmacol* 588(1):124-133; 2008.
121. Tsai, N. M.; Chen, Y. L.; Lee, C. C.; Lin, P. C.; Cheng, Y. L.; Chang, W. L.; Lin, S. Z.; Harn, H. J. The natural compound n-butylidenephthalide derived from *Angelica sinensis* inhibits malignant brain tumor growth in vitro and in vivo. *J Neurochem* 99(4):1251-1262; 2006.
122. Tuli, R.; Li, W. J.; Tuan, R. S. Current state of cartilage tissue engineering. *Arthritis Res Ther* 5(5):235-238; 2003.

123. Van Hoof, D.; Heck, A. J.; Krijgsveld, J.; Mummery, C. L. Proteomics and human embryonic stem cells. *Stem Cell Res* 1(3):169-182; 2008.
124. Wang, C. L.; Shi, D. Z.; Yin, H. J. [Effect of panax quinquefolius saponin on angiogenesis and expressions of VEGF and bFGF in myocardium of rats with acute myocardial infarction]. *Zhongguo Zhong xi yi jie he za zhi Zhongguo Zhongxiyi jiehe zazhi = Chinese journal of integrated traditional and Western medicine / Zhongguo Zhong xi yi jie he xue hui, Zhongguo Zhong yi yan jiu yuan zhu ban* 27(4):331-334; 2007.
125. Wang, J.; Wang, X.; Jiang, S.; Yuan, S.; Lin, P.; Zhang, J.; Lu, Y.; Wang, Q.; Xiong, Z.; Wu, Y. and others. Growth inhibition and induction of apoptosis and differentiation of tanshinone IIA in human glioma cells. *J Neurooncol* 82(1):11-21; 2007.
126. Wang, N. Y.; Lu, C. J.; Chen, X. H. [Study on effect of ginsenoside Rg1 in promoting myocardial vascular endothelial cell regeneration through induction on bone marrow stem cell's migration and differentiation in rabbits of myocardial infarction]. *Zhongguo Zhong xi yi jie he za zhi Zhongguo Zhongxiyi jiehe zazhi = Chinese journal of integrated traditional and Western medicine / Zhongguo Zhong xi yi jie he xue hui, Zhongguo Zhong yi yan jiu yuan zhu ban* 25(10):916-919; 2005.
127. Wang, S. L.; Li, Y. B.; Wang, Y. P.; Feng, M. [Effect of TSPG on proliferation and differentiation of human embryonic neural stem cell into dopaminergic neuron]. *Zhongguo Zhong Yao Za Zhi* 32(13):1310-1313; 2007.
128. Watanabe, H.; Ohta, H.; Imamura, L.; Asakura, W.; Matoba, Y.; Matsumoto, K. Effect of Panax ginseng on age-related changes in the spontaneous motor activity and dopaminergic nervous system in the rat. *Jpn J Pharmacol* 55(1):51-56; 1991.
129. Wei, S. Y.; Zhang, P. X.; Yang, D. M.; Zhang, H. B.; Jiang, B. G. [Traditional Chinese medicine and formulas of improving peripheral nerve regeneration]. *Zhongguo Zhong Yao Za Zhi* 33(17):2069-2072; 2008.
130. Wen, C.; Xu, H.; Huang, Q. F. [Effect of drugs for promoting blood circulation on blood lipids and inflammatory reaction of atherosclerotic plaques in ApoE gene deficiency mice]. *Zhongguo Zhong Xi Yi Jie He Za Zhi* 25(4):345-349; 2005.
131. Wislet-Gendebien, S.; Hans, G.; Leprince, P.; Rigo, J. M.; Moonen, G.; Rogister, B. Plasticity of cultured mesenchymal stem cells: switch from nestin-positive to excitable neuron-like phenotype. *Stem Cells* 23(3):392-402; 2005.
132. Wong, R. W.; Rabie, A. B. Traditional Chinese medicines and bone

- formation--a review. *J Oral Maxillofac Surg* 64(5):828-837; 2006.
133. Wu, W.; Kuang, P.; Li, Z. Protective effect of radix *Salviae miltiorrhizae* on apoptosis of neurons during focal cerebral ischemia and reperfusion injury. *J Tradit Chin Med* 17(3):220-225; 1997.
  134. Yamaguchi, K.; Shinohara, C.; Kojima, S.; Sodeoka, M.; Tsuji, T. (2E,6R)-8-hydroxy-2,6-dimethyl-2-octenoic acid, a novel anti-osteoporotic monoterpene, isolated from *Cistanche salsa*. *Biosci Biotechnol Biochem* 63(4):731-735; 1999.
  135. Yandava, B. D.; Billingham, L. L.; Snyder, E. Y. "Global" cell replacement is feasible via neural stem cell transplantation: evidence from the dysmyelinated shiverer mouse brain. *Proc Natl Acad Sci U S A* 96(12):7029-7034; 1999.
  136. Yang, B. H.; Zhu, L. Q.; Zhang, J. Z.; Niu, F. L.; Cui, W. [Effects of *Astragalus membranaceus* and *Panax notoginseng* on the transformation of bone marrow stem cells and proliferation of EPC in vitro]. *Zhongguo Zhong yao za zhi = Zhongguo zhongyao zazhi = China journal of Chinese materia medica* 30(22):1761-1763; 2005.
  137. Yang, H. M.; Shin, H. K.; Kang, Y. H.; Kim, J. K. *Cuscuta chinensis* extract promotes osteoblast differentiation and mineralization in human osteoblast-like MG-63 cells. *J Med Food* 12(1):85-92; 2009.
  138. Yang, L. Y.; Huang, T. H.; Ma, L. Bone marrow stromal cells express neural phenotypes in vitro and migrate in brain after transplantation in vivo. *Biomed Environ Sci* 19(5):329-335; 2006.
  139. Yang, L. Y.; Zheng, J. K.; Wang, C. Y.; Xu, M. D. [Stromal cells from human Wharton's jelly differentiate into neural cells]. *Sichuan Da Xue Xue Bao Yi Xue Ban* 36(1):13-16; 2005.
  140. Yang, Q. Y.; Zhao, L. C. [Effect of yiqi wenyang huoxue recipe on bone marrow stem cells' mobilization and its influence on heart function in patients with myocardial infarction]. *Zhongguo Zhong xi yi jie he za zhi Zhongguo Zhongxiyi jiehe zazhi = Chinese journal of integrated traditional and Western medicine / Zhongguo Zhong xi yi jie he xue hui, Zhongguo Zhong yi yan jiu yuan zhu ban* 28(12):1078-1081; 2008.
  141. Yang, S. F.; Wu, Q.; Sun, A. S.; Huang, X. N.; Shi, J. S. Protective effect and mechanism of *Ginkgo biloba* leaf extracts for Parkinson disease induced by 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine. *Acta Pharmacol Sin* 22(12):1089-1093; 2001.
  142. Yoo, K. J.; Li, R. K.; Weisel, R. D.; Mickle, D. A.; Jia, Z. Q.; Kim, E. J.; Tomita, S.; Yau, T. M. Heart cell transplantation improves heart function in dilated cardiomyopathic hamsters. *Circulation* 102(19 Suppl 3):III204-209;

- 2000.
143. Yu, M. S.; Wong, A. Y.; So, K. F.; Fang, J. N.; Yuen, W. H.; Chang, R. C. New polysaccharide from *Nerium indicum* protects neurons via stress kinase signaling pathway. *Brain Res* 1153:221-230; 2007.
  144. Yu, S.; Chen, K.; Li, S.; Zhang, K. In vitro and in vivo studies of the effect of a Chinese herb medicine on osteoclastic bone resorption. *Chin J Dent Res* 2(1):7-11; 1999.
  145. Yun, Y. J.; Lee, B.; Hahm, D. H.; Kang, S. K.; Han, S. M.; Lee, H. J.; Pyun, K. H.; Shim, I. Neuroprotective effect of palmul-chongmyeong-tang on ischemia-induced learning and memory deficits in the rat. *Biol Pharm Bull* 30(2):337-342; 2007.
  146. Zhang, J. F.; Sun, G. S.; Zhao, G. H. [Observation on therapeutic effect of herbs-partitioned moxibustion on Parkinson disease of 54 cases]. *Zhongguo Zhen Jiu* 25(9):610-612; 2005.
  147. Zhang, Q. J.; Zhang, Y. Y.; Huang, W. Y. [Traditional Chinese medicine in treatment of Parkinson's disease]. *Zhong Xi Yi Jie He Xue Bao* 2(1):75-77; 2004.
  148. Zhang, Y.; Leung, P. C.; Che, C. T.; Chow, H. K.; Wu, C. F.; Wong, M. S. Improvement of bone properties and enhancement of mineralization by ethanol extract of *Fructus Ligustri Lucidi*. *Br J Nutr* 99(3):494-502; 2008.
  149. Zhao, G.; Jiang, Z. H.; Zheng, X. W.; Zang, S. Y.; Guo, L. H. Dopamine transporter inhibitory and antiparkinsonian effect of common flowering quince extract. *Pharmacol Biochem Behav* 90(3):363-371; 2008.
  150. Zhao, G.; Li, S.; Qin, G. W.; Fei, J.; Guo, L. H. Inhibitive effects of *Fructus Psoraleae* extract on dopamine transporter and noradrenaline transporter. *J Ethnopharmacol* 112(3):498-506; 2007.
  151. Zhao, G.; Qin, G. W.; Wang, J.; Chu, W. J.; Guo, L. H. Functional activation of monoamine transporters by luteolin and apigenin isolated from the fruit of *Perilla frutescens* (L.) Britt. *Neurochem Int* 56(1):168-176.
  152. Zhao, H. W.; Li, X. Y. Ginkgolide A, B, and huperzine A inhibit nitric oxide-induced neurotoxicity. *Int Immunopharmacol* 2(11):1551-1556; 2002.
  153. Zhu, W. W.; Liu, Z. L.; Xu, H. W.; Chu, W. Z.; Ye, Q. Y.; Xie, A. M.; Chen, L.; Li, J. R. [Effect of the oil from *ganoderma lucidum* spores on pathological changes in the substantia nigra and behaviors of MPTP-treated mice]. *Di Yi Jun Yi Da Xue Xue Bao* 25(6):667-671; 2005.