Genetic polymorphisms of DNA double strand break gene Ku70 and gastric cancer in Taiwan

Mei-Due Yang^a, Hwei-Chung Wang^a, Wen-Shin Chang^{b,c}, Chia-Wen Tsai^{b,c}, and Da-Tian Bau^{b,c,d}

^a Department of General Surgery, China Medical University Hospital, Taichung, Taiwan,

R.*O*.*C*.

^b Terry Fox Cancer Research Lab, China Medical University Hospital, Taichung, Taiwan,

R.*O*.*C*.

^c Graduate Institute of Basic Medical Science, China Medical University, Taichung,

Taiwan, R.O.C.

^d Institute of Clinical Medical Science, China Medical University, Taichung, Taiwan,

R.O.C.

Correspondence to: Da-Tian Bau, Terry Fox Cancer Research Lab, Department of Medical Research, China Medical University Hospital, 2 Yuh-Der Road, Taichung, 404 Taiwan, R.O.C. Tel: +886422053366 Ext 3312, Fax: +886422053366 Ext 1511, e-mail: datian@mail.cmuh.org.tw; artbau2@gmail.com

Emails of other authors: MD Yang: <u>d5218ymd@yahoo.com.tw</u> WC Wang: <u>whcym64@yahoo.com.tw</u> WS Chang: <u>halittlemelon@hotmail.com</u>

CW Tsai: wenwen816@gmail.com

Keywords Ku70· Polymorphism· Gastric cancer· Carcinogenesis

Abstract

Background and aim The DNA repair gene Ku70, an important member of non-homologous end-joining repair system, is thought to play an important role in the repairing of DNA double strand breaks. It is known that defects in double strand break repair capacity can lead to irreversible genomic instability. However, the polymorphic variants of *Ku70*, have never been reported about their association with gastric cancer susceptibility.

Methods In this hospital-based case-control study, the associations of *Ku70* promoter T-991C (rs5751129), promoter G-57C (rs2267437), promoter A-31G (rs132770), and intron 3 (rs132774) polymorphisms with gastric cancer risk in a Taiwanese population were investigated. In total, 136 patients with gastric cancer and 560 age- and gender-matched healthy controls recruited from the China Medical Hospital in Taiwan were genotyped.

Results As for *Ku70* promoter T-991C, the ORs after adjusted by age and gender of the people carrying TC and CC genotypes were 2.41 (95% CI=1.53-3.88) and 3.21 (95% CI=0.96-9.41) respectively, compared to those carrying TT wild-type genotype. The *P* for trend was significant (*P*<0.0001). In the dominant model (TC plus CC versus TT), the association between *Ku70* promoter T-991C polymorphism and the risk for gastric cancer was also significant (adjusted OR=2.48, 95% CI=1.74-3.92).

When stratified by age and gender, the association was restricted to those at the age of 55 or elder of age (TC vs TT: adjusted OR=2.52, 95% CI=1.37-4.68, P=0.0139) and male (TC vs TT: adjusted OR=2.58, 95% CI=1.33-4.47, P=0.0085). As for the other three polymorphisms, there was no difference between both groups in the distributions of their genotype frequencies.

Conclusion In conclusion, the *Ku70* promoter T-991C (rs5751129), but not the *Ku70* promoter C-57G (rs2267437), promoter A-31G (rs132770) or intron 3 (rs132774), is associated with gastric cancer susceptibility. This polymorphism may be a novel useful marker for gastric carcinogenesis.

Background

Gastric cancer is the fourth most common cancer over the world and affects approximately 900,000 individuals every year [1]. Although the identification of *Helicobacter pylori* has revolutionized the understanding of its epidemiology and pathogenesis, the initiation etiology and genomic contributing factors of gastric cancer are still largely unknown [2]. Human DNA repair mechanisms protect the genome from various insults caused by endogenous and environmental agents [3], and mutations or defects in the DNA repairing system are thought to be essential for tumorigenesis [4, 5]. Therefore, it is logical to suspect that genetic variants of DNA repair genes might contribute to gastric cancer pathogenesis.

DNA double strand breaks (DSBs) are repaired by the DNA DSB repair system [6, 7], which consists of two subpathways, homologous recombination (HR) and nonhomologous end-joining (NHEJ) [8]. In humans, NHEJ is the predominant repair system. At present, several proteins involved in the NHEJ pathway have been identified; namely, the ligase IV and its associated protein XRCC4, the three components of the DNA dependent protein kinase (DNA-PK) complex, Ku70, Ku80, and the catalytic subunit PKcs [9]. Genetic variation in DNA repair genes has been postulated as an important contributor to the aetiology of gastric cancer [10, 11]. However, there is seldom information regarding gastric cancer and DNA repair gene polymorphisms. As for NHEJ, genetic polymorphisms influence DNA repair capacity and confer predisposition to several cancers, including skin [12], breast [13-15], bladder [16, 17], and oral cancers [18, 19].

In our previous study, we have found that one polymorphism of *XRCC4* DSB repair gene is associated with gastric cancer susceptibility in Taiwan [20]. In this study, we assumed that the upstream gene *Ku70* in NHEJ, like *XRCC4*, its polymorphisms may also contribute to gastric carcinogenesis. To test this hypothesis, we determined the genotypic frequency of four polymorphisms of the *Ku70* gene at promoter T-991C (rs5751129), promoter G-57C (rs2267437), promoter A-31G (rs132770), and intron 3 (rs132774), using a polymerase chain reaction (PCR)-based restriction fragment length polymorphism (RFLP) method. To the best of our knowledge, this is the first study carried out to evaluate the contribution of *Ku70* polymorphisms in gastric oncology all over the world.

Methods

Study population and sample collection

One hundred and thirty six patients diagnosed with gastric cancer were recruited at the outpatient clinics of general surgery between 2005-2008 at the China Medical University Hospital, Taichung, Taiwan, Republic of China. The mean age of the gastric cancer patients and the controls were 51.4 (range = 38 to 79, SD = 9.3) and 52.2 (range = 41 to 77, SD = 7.6) years, respectively. All patients voluntarily participated, completed a self-administered questionnaire and provided peripheral blood samples. Five hundred and sixty non-gastric cancer healthy people as controls were selected by matching for age and gender after initial random sampling from the Health Examination Cohort of the hospital. Our study was approved by the Institutional Review Board of the China Medical University Hospital and written-informed consent was obtained from all participants.

Genotyping conditions

Genomic DNA was prepared from peripheral blood leucocytes using a QIAamp Blood Mini Kit (Blossom, Taipei, Taiwan). The PCR cycling conditions were: one cycle at 94°C for 5 min; 35 cycles of 94°C for 30 sec, 55°C for 30 sec, and 72°C for 30 sec, and a final extension at 72°C for 10 min. Pairs of PCR primer sequences and restriction enzyme for each DNA product are all listed in Table 1. Then, 10 μ l of product was loaded into a 3% agarose gel containing ethidium bromide for electrophoresis. Ten percent of the samples both in control and patient groups were analyzed of their genotypes by PCR direct sequencing (Genomics BioSci & Tech Co., Taipei) and the consistency is 100%.

Statistical analyses

To ensure that the controls used were representative of the general population and to exclude the possibility of genotyping error, the deviation of the genotype frequencies of Ku70 single nucleotide polymorphisms in the control subjects from those expected under the Hardy-Weinberg equilibrium was assessed using the goodness-of-fit test. Pearson's Chi-square test or Fisher's exact test (when the expected number in any cell was less than five) was used to compare the distribution of the Ku70 genotypes between cases and controls. Cancer risk associated with the genotypes was estimated as odds ratio (ORs) and 95% confidence intervals (CIs) using unconditional logistic regression. Data was recognized as significant when the statistical *P*-value was less than 0.05.

Results

There were no significant differences between both groups in their age, sex, and other cancer prevalence. The frequencies of the genotypes of the Ku70 promoter T-991C, promoter C-57G, promoter A-31G, and intron 3 polymorphisms in the gastric cancer and control groups are shown in Table 2. The ORs after adjusted by age and gender of the people carrying TC and CC genotypes were 2.41 (95% CI=1.53-3.88) and 3.21 (95% CI=0.96-9.41) respectively, compared to those carrying TT wild-type genotype. The P for trend was significant (P<0.0001). In the dominant model (TC plus CC versus TT), the association between Ku70 promoter T-991C polymorphism and the risk for gastric cancer was also significant (adjusted OR=2.48, 95% CI=1.74-3.92). On the contrary, as for the Ku70 promoter C-57G, promoter A-31G, and intron 3 polymorphisms, the distributions of these polymorphisms were in Hardy-Weinberg equilibrium but there was no difference between gastric cancer and control groups in the distribution in the genotype frequency at these SNP sites (Table 2).

We have further evaluated the genetic association between gastric cancer in different age range and gender by performing the stratification analysis of the Ku70 promoter T-991C genotypes (Table 3). As for those people under the age of 55, the distribution of the Ku70promoter T-991C genotypes was not significantly different between case and control groups. Interestingly, an increased risk was found in those people at the age of 55 or elder who carried TC genotype, compared with TT genotype (adjusted OR=2.52, 95% CI=1.37-4.68, P=0.0139) (Table 3). When data were stratified by gender, the significantly increased risk was found in those male carrying TC genotype, compared with TT genotype (adjusted OR=2.58, 95% CI=1.33-4.47, P=0.0085). There was no difference found in female (Table 3).

Discussion

The present study is the first one to investigate the role of Ku70 gene polymorphisms, which has never been reported to be associated with gastric cancer risk. Our study revealed that the Ku70 promoter T-991C polymorphism was associated with the susceptibility to gastric cancer, but the C-57G, A-31G or intron 3 polymorphisms were not (Table 2), especially in those at the age of 55 or elder, and in male (Table 3). In 2003, Fu et al have reported that, Ku70 rs2267437 polymorphism (which was called C-61G in their paper) was associated with breast cancer risk [21]. It is reasonable that we did not find a significant association of this SNP with gastric cancer as they did in this SNP with breast cancer in Taiwan population. Instead, we found that the genotype of Ku70 rs2267437 was associated with gastric cacner. Since Ku70 rs2267437 located in the promoter region, its different genotypes may determine the differential expression level of Ku70 protein in the gastric tissue. Thus, the genotypes of *Ku70* promoter *T-991C* (rs5751129) may determine the expression levels of Ku70 in different stages, regulate the DNA repair capacity and overall genome stability, and play a role in gastric carcinogenesis. Supporting the idea above, Hu et al reported that the expression of TERT and Ku70 proteins was significantly up-regulated in both precancerous lesion and gastric cancer tissues and was significantly higher in gastric cancer tissues than that in precancerous lesions [22]. In the future, the correlation between Ku70 rs5751129 genotype and phenotype still needs further investigation. In addition to be associated with oral cancer

[18], *Ku70* rs5751129 polymorphism was also found to be associated with two-side pterygium pathology, which are caused by an uncontrolled cell proliferation like that of a tumor [23].

We have taken into consideration and tried our best to conquer the limitations of this study. First, for the possibility that some findings of further sub-grouping are attributed to chance because of the limited numbers of patients, we just provided the main finding with these 136 gastric patients and 560 (more than four-fold of the number of the patients) non-cancer controls, which is very convincing and reliable with multiple-checked significances (P < 0.05 both in genotype and allelic frequencies, and those obvious odds ratios and 95% CIs). Second, the false positive findings are statistically explained by hidden population stratification. But in this study, it seems unlikely that population stratification is a major concern, for all the cases and the controls recruited in this study were drawn from the same Taiwanese ethnic group and the Taiwanese population has relatively homogenous genetic background [24]. Therefore, the potential confounding effect of population stratification for genotyping data is not a major concern. Third, the possible selection bias was taken into consideration and reduced to a lowest level by frequency matching on age and gender between the cases and controls. Forth, from another angle, the frequencies of Ku70 polymorphisms variant alleles were similar to those reported in the NCBI website in the Asian population studies, for example C allele frequencies of Ku70 promoter T-991C are 5.7% in our control group and 4.2~8.9% for Asian populations in NCBI, which suggest no selection bias for the subject's enrolments in terms of genotypes. Therefore, the need for the present results of Ku70 promoter T-991C to be verified in further larger gastric patient population is not so urgent. Of course, further studies in larger population and other functional repair assays should be performed in the future for further findings, particularly those among subgroups by clinical and environmental factors, and clarify the mechanisms involved.

In this study, the genotype distribution of the C allele at Ku70 promoter T-991C was significantly higher in the gastric cancer group (12.5%) than in the control group (5.7%). It was also found that patients carrying heterozygous TC for Ku70 promoter T-991C had a 2.41-fold higher risk of gastric cancer (Table 2). Combination of heterozygous and homozygous (TC or CC) was almost at the same level (OR=2.48) for gastric cancer risk (Table 2). All these data suggested that the C allele at Ku70 promoter T-991C was a novel and important biomarker for gastric cancer. As for the Ku70 promoter C-57G, A-31G or intron 3 polymorphisms, our results indicated that their genetic differences were not associated with gastric cancer risk in Taiwanese population (Table 2).

Conclusions

In conclusion, this is the first report to investigate the association between Ku70 gene polymorphisms and gastric cancer. Our findings suggested that Ku70 promoter T-991C, but not Ku70 promoter C-57G, promoter A-31G or intron 3 polymorphisms, was associated with higher susceptibility to gastric cancer. The Ku70 promoter T-991C polymorphism might become a potential biomarker for the gastric oncology prediction and this paper may also provide a valuable insight into the gastric carcinogenesis.

Abbreviations

Double strand breaks (DSBs); homologous recombination (HR); nonhomologous end-joining (NHEJ); polymerase chain reaction (PCR); restriction fragment length polymorphism (RFLP); odds ratio (ORs); confidence intervals (CIs).

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

MD Yang participated in the collection of the samples and clinical data. HC Wang participated in the literature reviewing. WS Chang carried out the PCR-RFLP and sequencing genotyping. CW Tsai performed the statistical analysis. DT Bau designed the study and

coordinated the whole team work. All authors read and approved the final manuscript.

Acknowledgements

This study was supported by research grants from the China Medical University and Hospital (DMR-99-041), Terry Fox Cancer Research Foundation and National Science Council (NSC 98-2320-B-039-010-MY3).

References

- 1. Steward BW. WHO: World Cancer Report 2003. IARC Press, Lyon, 2004.
- 2. Fuchs CS, Mayer RJ. Gastric carcinoma. N Engl J Med. 1995;333:32-41.
- 3. Sugimura T, Kumimoto H, Tohnai I, Fukui T, Matsuo K, Tsurusako S, Mitsudo K, Ueda M, Yajima K, Ishizaki K. Gene-environment interaction involved in oral carcinogenesis: molecular epidemiological study for metabolic and DNA repair gene polymorphisms. J Oral Pathol Med. 2006;35:11-18.
- Miller KL, Karagas MR, Kraft P, Hunter DJ, Catalano PJ, Byler SH, Nelson HH. XPA, haplotypes, and risk of basal and squamous cell carcinoma. Carcinogenesis. 2006;27:1670-1675.
- Vogelstein B, Alberts B, Shine K. Genetics. Please don't call it cloning! Science.
 2002;295:1237.
- Wood RD, Mitchell M, Sgouros J, Lindahl T. Human DNA repair genes. Science. 2001;291:1284-1289.
- 7. Yu Z, Chen J, Ford BN, Brackley ME, Glickman BW. Human DNA repair systems: an overview. Environ Mol Mutagen. 1999;33:3-20.
- Khanna KK, Jackson SP. DNA double-strand breaks: signaling, repair and the cancer connection. Nat Genet. 2001;27:247-254.
- 9. Jackson SP. Sensing and repairing DNA double-strand breaks. Carcinogenesis.

2002;23:687-696.

- Gonzalez CA, Sala N, Capella G. Genetic susceptibility and gastric cancer risk. Int J Cancer. 2002;100:249-260.
- 11. Goode EL, Ulrich CM, Potter JD. Polymorphisms in DNA repair genes and associations with cancer risk. Cancer Epidemiol Biomarkers Prev. 2002;11:1513-1530.
- Han J, Colditz GA, Samson LD, Hunter DJ. Polymorphisms in DNA double-strand break repair genes and skin cancer risk. Cancer Res. 2004;64:3009-3013.
- 13. Bau DT, Fu YP, Chen ST, Cheng TC, Yu JC, Wu PE, Shen CY. Breast cancer risk and the DNA double-strand break end-joining capacity of nonhomologous end-joining genes are affected by BRCA1. Cancer Res. 2004;64:5013-5019.
- 14. Bau DT, Mau YC, Ding SL, Wu PE, Shen CY. DNA double-strand break repair capacity and risk of breast cancer. Carcinogenesis. 2007;28:1726-1730.
- 15. Chiu CF, Wang HC, Wang CH, Wang CL, Lin CC, Shen CY, Chiang SY, Bau DT. A new single nucleotide polymorphism in XRCC4 gene is associated with breast cancer susceptibility in Taiwanese patients. Anticancer Res. 2008;28:267-270.
- Chang CH, Chang CL, Tsai CW, Wu HC, Chiu CF, Wang RF, Liu CS, Lin CC, Bau DT. Significant association of an XRCC4 single nucleotide polymorphism with bladder cancer susceptibility in Taiwan. Anticancer Res. 2009;29:1777-1782.
- 17. Chang CH, Wang RF, Tsai RY, Wu HC, Wang CH, Tsai CW, Chang CL, Tsou YA, Liu

CS, Bau DT. Significant association of XPD codon 312 single nucleotide polymorphism with bladder cancer susceptibility in Taiwan. Anticancer Res. 2009;29:3903-3907.

- 18. Bau DT, Tseng HC, Wang CH, Chiu CF, Hua CH, Wu CN, Liang SY, Wang CL, Tsai CW, Tsai MH. Oral cancer and genetic polymorphism of DNA double strand break gene Ku70 in Taiwan. Oral Oncol. 2008;44:1047-1051.
- 19. Chiu CF, Tsai MH, Tseng HC, Wang CL, Wang CH, Wu CN, Lin CC, Bau DT. A novel single nucleotide polymorphism in XRCC4 gene is associated with oral cancer susceptibility in Taiwanese patients. Oral Oncol. 2008;44:898-902.
- 20. Chiu CF, Wang CH, Wang CL, Lin CC, Hsu NY, Weng JR, Bau DT. A novel single nucleotide polymorphism in XRCC4 gene is associated with gastric cancer susceptibility in Taiwan. Ann Surg Oncol. 2008;15:514-518.
- 21. Fu YP, Yu JC, Cheng TC, Lou MA, Hsu GC, Wu CY, Chen ST, Wu HS, Wu PE, Shen CY. Breast cancer risk associated with genotypic polymorphism of the nonhomologous end-joining genes: a multigenic study on cancer susceptibility. Cancer Res. 2003;63:2440-2446.
- 22. Hu H, Zhang Y, Zou M, Yang S, Liang XQ. Expression of TRF1, TRF2, TIN2, TERT, KU70, and BRCA1 proteins is associated with telomere shortening and may contribute to multistage carcinogenesis of gastric cancer J Cancer Res Clin Oncol.

2010;136:1407-1414.

- 23. Tsai YY, Bau DT, Chiang CC, Cheng YW, Tseng SH, Tsai FJ. Pterygium and genetic polymorphism of DNA double strand break repair gene Ku70. Mol Vis. 2007;13:1436-1440.
- 24. Yang HC, Lin CH, Hsu CL, Hung SI, Wu JY, Pan WH, Chen YT, Fann CS. A comparison of major histocompatibility complex SNPs in Han Chinese residing in Taiwan and Caucasians. J Biomed Sci. 2006;13:489-498.

Polymorphism	Primers sequences (5' to 3')	Restriction	SNP	DNA fragment
(location)		enzyme	sequence	size (bp)
promoter T-991C	F: AACTCATGGACCCACGGTTGTGA	Dpn II	Т	301
(rs5751129)	R: CAACTTAAATACAGGAATGTCTTG		С	200 + 101
promoter G-57C	F: AACTCATGGACCCACGGTTGTGA	Hae II	С	298
(rs2267437)	R: CAACTTAAATACAGGAATGTCTTG		G	195 + 103
promoter A-31G	F: TACAGTCCTGACGTAGAAG	Mnl I	G	226
(rs132770)	R: AAGCGACCAACTTGGACAGA		А	146 + 80
intron 3	F: GTATACTTACTGCATTCTGG	Msc I	G	160
(rs132774)	R: CATAAGTGCTCAGTACCTAT		С	114 + 46

 Table 1. The primer sequences and restriction fragment length polymorphism conditions for Ku70 gene polymorphisms.

*F and R indicate forward and reverse primers, respectively.

	Case (%)	Control (%)	Adjusted OR ^a (95% CI)	<i>P</i> -value
promoter T-991C (rs5751129)				
TT	106 (77.9)	502 (89.6)	1.00 (ref)	
ТС	26 (19.1)	52 (9.3)	2.41 (1.53-3.88)	0.0020
CC	4 (3.0)	6 (1.1)	3.21 (0.96-9.41)	0.0837
<i>P</i> for trend			2.87 (1.37-5.02)	<0.0001
(TC+CC) vs TT			2.48 (1.74-3.92)	0.0005
CC vs (TT+TC)			2.67 (0.71-9.69)	0.1114
Promoter G-57C (rs2267437)				
CC	95 (69.9)	383 (68.4)	1.00 (ref)	
CG	37 (27.2)	167 (29.8)	0.91 (0.48-1.38)	0.6722
GG	4 (2.9)	10 (1.8)	1.56 (0.55-4.97)	0.4954
<i>P</i> for trend			0.93 (0.53-3.34)	0.6164
(CG+GG) vs CC			0.98 (0.63-1.52)	0.8367
GG vs (CC+CG)			1.52 (0.58-5.01)	0.4917
promoter A-31G (rs132770)				
GG	109 (80.1)	459 (82.0)	1.00 (ref)	
GA	19 (14.0)	73 (13.0)	1.16 (0.71-1.93)	0.7763

Table 2. The associations between Ku70 polymorphisms and gastric cancer risk

AA	8 (5.9)	28 (5.0)	1.25 (0.51-2.68)	0.6642
<i>P</i> for trend			1.18 (0.68-2.34)	0.9065
(GA+AA) vs GG			1.14 (0.73-1.76)	0.6227
AA vs (GG+GA)			1.03 (0.37-3.01)	0.6673
intron 3 (rs132774)				
GG	113 (83.1)	459 (82.0)	1.00 (ref)	
GC	23 (16.9)	101 (18.0)	0.91 (0.60-1.73)	0.8040
CC	0 (0.0)	0 (0.0)		

^a Adjusted by age and gender; the line with ORs that significantly differ from 1.00 are bolded

		Case (%)	Control (%)	Adjusted OR ^a (95% CI)	<i>P</i> -value
Age					
< 55					
	TT	52 (80.0)	238 (90.8)	1.00 (ref)	
	TC	11 (16.9)	22 (8.4)	2.18 (0.97-4.88)	0.0600
	CC	2 (3.1)	2 (0.8)	4.31 (0.69-27.33)	0.1551
≥ 55					
	TT	54 (76.1)	264 (88.6)	1.00 (ref)	
	TC	15 (21.1)	30 (10.1)	2.52 (1.37-4.68)	0.0139
	CC	2 (2.8)	4 (1.3)	2.61 (0.51-11.62)	0.2776
Gender					
Male					
	TT	69 (76.7)	336 (89.1)	1.00 (ref)	
	TC	18 (20.0)	36 (9.6)	2.58 (1.33-4.47)	0.0085
	CC	3 (3.3)	5 (1.3)	2.85 (0.71-11.36)	0.1477
Fema	le				

Table 3. Distribution of Ku70 promoter T-991C genotypes stratified by age and gender in the gastric cancer and

control groups

22

TT	37 (80.4)	166 (90.7)	1.00 (ref)	
TC	8 (17.4)	16 (8.7)	2.13 (0.79-6.03)	0.1019
CC	1 (2.2)	1 (0.6)	4.07 (0.31-69.45)	0.3371

^a Adjusted by age and gender when appropriate; the line with ORs that significantly differ from 1.00

are bolded