

Manuscript Number: ARCHIVES-PMR-D-10-00769R2

Title: Spinal Cord Mechanism Involving the Remote Effects of Dry Needling on the Irritability of Myofascial Trigger Spots in Rabbit Skeletal Muscle

Article Type: Original Article

Keywords: Neural pathway; Dry needling; Myofascial trigger spot; Electromyography; Remote effect

Corresponding Author: Dr. Chang-Zern Hong, M.D.

Corresponding Author's Institution: University of California Irvine

First Author: Yueh-Ling Hsieh, PT, PhD

Order of Authors: Yueh-Ling Hsieh, PT, PhD; Li-Wei Chou, MD, MSc; Yie-San Joe, MD; Chang-Zern Hong, M.D.

Abstract: Objective: To elucidate the neural mechanisms underlying the remote effects produced by dry needling rabbit skeletal muscle myofascial trigger spots (MTrSs) via analyses of their endplate noise (EPN) recordings.

Design: Experimental animal controlled trial.

Setting: An animal laboratory of a university.

Animals: Ninety-six male New Zealand rabbits (body weight: 2.5-3.0 kg, age: 16-20 weeks).

Intervention: Animals received no intervention for neural interruption in Group I, transection of the tibial nerve in Group II, transection of L5 and L6 spinal cord in Group III, and transection of the T1 and T2 spinal cord in Group IV. Each group was further divided into 4 subgroups: animals received ipsilateral dry needling (IDN), contralateral dry needling (CDN), ipsilateral sham needling (ISN), or contralateral sham needling (CSN), of gastrocnemius (GAS) MTrSs.

Main Outcome Measures: EPN amplitudes of biceps femoris (BF) MTrSs.

Results: BF MTrS mean EPN amplitudes significantly increased ( $P < .05$ ) initially after GAS verum needling, but reduced to a level significantly lower ( $P < .05$ ) than the pre-needling level in Groups I and IV with IDN or CDN, and in Group II with CDN (but not IDN). No significant EPN amplitude changes were observed in BF MTrS in Group III or in the control animals receiving superficial needling (sham).

Conclusion: This remote effect of dry needling depends on an intact afferent pathway from the stimulating site to the spinal cord and a normal spinal cord function at the levels corresponding to the innervation of the proximally affected muscle.

November 14, 2010

RE: **ARCHIVES-PMR-D-10-00769R1**

**Dear Dr. Basford:**

Thank you very much again for further reviewing our manuscript entitled "**Spinal Cord Mechanism Involving the Remote Effects of Dry Needling on the Irritability of Myofascial Trigger Spots in Rabbit Skeletal Muscle**". I deeply appreciate the suggestion about editing and English correction. Dr. Hsieh and I have tried our best to make revision following the comments and suggestions from you and the reviewer.

Please find the following items accompanied with this submission:

1. A list of our **responses to reviewer's comments** - highlighted in yellow.
2. A **marked copy of the revised manuscript** - highlighted on the revised portion in yellow.
3. A **clean copy of the revised manuscript**.
4. Two copies of new **Figure 2** (black and white)- pdf and power-point files.
5. Four copies (one for each author) of the **signed "Disclosure Statements and Copyright Assignment form"**

We are looking forward to hearing from you about the final decision. Thank you again.

With regards

**Chang-Zern Hong, M.D.**

Research Professor

Hung-Kuang University

Sa-Lu, Tai-Chung

TAIWAN

and

Recalled Professor

University of California Irvine

Irvine, CA

## RESPONSES TO REVIEWERS COMMENTS

-Highlighted in yellow

Ms. Ref. No.: **ARCHIVES-PMR-D-10-00769R1**

Title: **Spinal Cord Mechanism Involving the Remote Effects of Dry Needling on the Irritability of Myofascial Trigger Spots in Rabbit Skeletal Muscle**

### Comments from Editor-in-Chief

Presentation. While your revisions have led to an improved paper, there remain a number of areas where it is difficult to follow your thoughts. This is most likely in part due to the changes you have made. A number of specific suggestions appear below, but I again suggest that the revised paper undergo a final editing with attention to word flow and clarity.

Abstract. Objective. Delete the first six words and begin with "To.."

#### **Response:**

We have modified this sentence according to the reviewer's suggestion (see **Comments from Reviewer** below).

It has been changed into "To elucidate the neural mechanisms underlying the remote effects produced by dry needling rabbit skeletal muscle myofascial trigger spots (MTrSs) via analyses of their endplate noise (EPN) recordings".

Lines 55-57. This material in the first sentence is difficult to follow and may be more detailed than necessary. Would the substitution of something on the order of "Myofascial pain is one of the most common sources of musculoskeletal pain and has as its hallmark the presence of taut bands and small hyperirritable regions referred to as myofascial trigger points (MTrPs).

#### **Response:**

We have corrected this sentence according to your suggestion.

Line 63. Perhaps "Dry needling has a well established role in the treatment of myofascial pain.

#### **Response:**

We have modified this sentence according to your suggestion.

Line 66. Delete "either..located."

#### **Response:**

We have modified this portion according to the reviewer's suggestion (see **Comments**

from Reviewer below).

It has been changed into “Clinical studies have demonstrated that dry needling MTrPs at the pain site suppresses their activity resulting in pain reduction, but also that those MTrPs can also be suppressed through dry needling distant MTrPs that are anatomically proximal or distal to the site of clinical pain.<sup>8-12</sup> Acupuncture studies have demonstrated that the Traditional Chinese Medicine principle that pain conditions can be improved by stimulating acupuncture points distant from the site of pain.<sup>4,13-14</sup>”.

Line 111. "Carefully" might be deleted.

**Response:**

We have deleted the word "Carefully".

Lines 452-465. See reviewer comments about deleting this section.

**Response:**

**We try to emphasize the importance of this special technique (to elicit LTRs). If the experiment is not carried with this technique, we would not be able to find the remote effects of needling. Furthermore, both LTRs and neural control of needling are related to the spinal cord mechanism. The spinal cord mechanism of dry needling can change EPN activity and achieve the remote needling effect. Therefore, we prefer to keep this section.**

Figure 2. Please convert to black and white.

**Response:**

We have converted this figure 2 into black and white.

A Disclosure Statements and Copyright Assignment form **will be emailed to you shortly**. We require ALL coauthors to complete, sign, and submit the form at this time. These forms replace those provided at original submission. Please return the forms by EES upload or fax. Editors must have these documents in hand for each member of the author group before proceeding with manuscript evaluation. Again, if you submitted these forms online, you must resubmit them to the Editorial Office now.

**Response:**

**Four copies (one for each author) of the signed “Disclosure Statements and Copyright Assignment form” have been sent accompanied with this submission.**

When you submit the revised manuscript, please include, in a file separate from your cover letter, an itemized response to each of the suggested revisions and any other changes made. Use consecutive line numbering and cite line numbers for each change. In addition, highlight each change in the revised manuscript. You must return the revised manuscript by **Nov 30 2010 12:00AM**.

**Response:**

**We have sent the following items accompanied with this submission:**

1. A **marked copy of the revised manuscript** - highlighted on the revised portion in yellow.
2. A **clean copy of the revised manuscript**.
3. Two copies of new **Figure 2** (black and white)- pdf and power-point files.
4. Four copies (one for each author) of the **signed "Disclosure Statements and Copyright Assignment form"**

## **Reviewer comment excerpts:**

### **General Issues**

Is a MTrS really any different than a MTrP? Perhaps MTrP use across the paper might be less confusing for the reader if they are clinically equivalent and this could just be notated in the abstract

**Response:**

We used this term for all animal-study papers published previously, since late Dr. David Simons insisted that we could not measure the subjective pain in animal (like human) and should use the term "MTrS" for animal. As far as we can define the term clearly (as listed in the "Acronyms" in this paper), the reader should not be confused.

### **Abstract**

In the Objective section, "The purpose of this study is to investigate the possible neural pathway for the remote effects of dry needling based on the assessment of endplate noise (EPN) recorded for the myofascial trigger spot (MTrS) in rabbit skeletal muscle" might be better worded "The purpose of this study is to elucidate the neural mechanisms underlying the remote effects produced by dry needling rabbit skeletal muscle myofascial trigger spots (MTrSs) via analyses of their endplate noise (EPN) recordings."

**Response:**

We have made a revision according to your suggestion.

In the Intervention section line 24, "on the MTrS of the gastrocnemius" would be better worded "of gastrocnemius (GAS) MTrSs"

**Response:**

We have made a revision according to your suggestion.

In the Main Outcome Measures section lines 25-26, "Amplitudes of EPN on the MTrS region of biceps femoris" would be better worded "EPN amplitudes of biceps femoris (BF)

MTrSs"

**Response:**

We have made a revision according to your suggestion.

In the Results section, "Mean EPN amplitudes significantly increased ( $P < .05$ ) initially, but reduced to a level significantly lower ( $P < .05$ ) than the pre-needling level in Group I with IDN or CDN, Group II with CDN 29 (but not IDN), and Group IV with IDN or CDN, but no such changes were observed in Group III. There were no significant changes ( $P > .05$ ) in EPN amplitudes in all control animals" could be more succinctly and perhaps better worded as "BF MTrS mean EPN amplitudes significantly increased ( $P < .05$ ) initially after GAS verum needling, but reduced to a level significantly lower ( $P < .05$ ) than the pre-needling level in Groups I and IV with IDN or CDN, and in Group II with CDN (but not IDN). No significant EPN amplitude changes were observed in BF MTrS in Group III or in the control animals receiving superficial needling (sham)."

**Response:**

We have revised the above two sentences according to your suggestion.

In Conclusions section, line 34, "level corresponding to the innervations of the proximally affected muscle" would be better worded "levels corresponding to the innervation of the proximally affected muscle"

**Response:**

We had modified it as your suggestion.

## Introduction

First paragraph, "Myofascial pain is one of the most common musculoskeletal pain associated with myofascial trigger points (MTrPs) which are hyperirritable spots in taut bands of skeletal muscle fibers due to accumulation of unique hypersensitive loci.<sup>1,2</sup> Clinically, an MTrP is characterized with a typical referred pain pattern and a local twitch response (LTR) in response to snapping palpation" could be better worded and is not quite accurate as LTR are no longer considered a diagnostic criterion to diagnose MTrPs. A more accurate alternative wording might be "Myofascial pain syndrome, a common source of musculoskeletal pain, is associated with hypersensitive loci in taut bands of skeletal muscle fibers which are termed myofascial trigger points (MTrPs).<sup>1,2</sup> Clinically, a given MTrP has a characteristic referred pain pattern, and may be associated with a local twitch response (LTR) in response to snapping palpation"

**Response:**

The first sentence "'Myofascial pain is one..... of unique hypersensitive loci.<sup>1,2</sup> " has been changed (as suggested by the Editor-in-Chief) into "Myofascial pain is one of the most

common sources of musculoskeletal pain and has as its hallmark the presence of taut bands and small hyperirritable regions referred to as myofascial trigger points (MTrPs).”.

The second sentence “Clinically, an MTrP is characterized .....to snapping palpation” has also been changed into “Clinically, a given MTrP has a characteristic referred pain pattern, and may be associated with a local twitch response (LTR) in response to snapping palpation”.

line 66, the reference "8,9-12" could just be "8-12" since the references are consecutive

**Response:**

We have made a revision according to your suggestion.

lines 64-66, "In addition to direct needling of the painful MTrP, clinical studies have demonstrated suppressive effect on this MTrP with dry needling at a remote MTrP, either proximally or distally located.<sup>8,9-12</sup> Similar remote effectiveness in pain control has also been documented in acupuncture therapy.<sup>4, 13, 14</sup>" is somewhat awkwardly worded and might better be worded "Clinical studies have demonstrated that dry needling MTrPs at the pain site suppresses their activity resulting in pain reduction, but also that those MTrPs can also be suppressed through dry needling distant MTrPs that are anatomically proximal or distal to the site of clinical pain.<sup>8-12</sup> Acupuncture studies have demonstrated that the Traditional Chinese Medicine principle that pain conditions can be improved by stimulating acupuncture points distant from the site of pain. <sup>4,13-14</sup>"

**Response:**

We have revised these two sentences according to your suggestion.

As per the recommendations for alteration to purpose noted in the abstract, perhaps the last paragraph of the introduction might be better worded "The purpose of this study is to confirm the remote effects of dry needling on trigger points and to elucidate the neural mechanisms underlying the remote effects produced by dry needling rabbit skeletal muscle myofascial trigger spots (MTrSs) via analyses of their endplate noise (EPN) recordings."

**Response:**

We have made a revision based on your suggestion. This sentence has been changed into: “The purpose of this study is to confirm the remote effects of dry needling on trigger points and to elucidate the neural mechanisms underlying the remote effects produced by dry needling rabbit myofascial trigger spots (MTrSs, equivalent to human MTrPs) in the gastrocnemius muscle (GAS)<sup>17,</sup><sup>18</sup> via analyses of EPN recordings from the biceps femoris (BF).”.

## Materials and Methods

Line 82 "age" should be "ages"

**Response:**

We have made a revision according to your suggestion.

Line 83, "cared" should be "cared for"

**Response:**

We have made a revision according to your suggestion.

Line 110, consider using "MTrP" instead of "MTrS" throughout paper

**Response:**

As mentioned above, We used this term for all animal-study papers published previously, since late Dr. David Simons insisted that we could not measure the subjective pain in animal (like human) and should use the term "MTrS" for animal. As far as we can define the term clearly (as listed in the "Acronyms" in this paper), the reader should not be confused.

Line 115, should put the manufacturer data in instead of the "a"

**Response:**

According to the "Instruction for Authors" (listed below), the suppliers should not be put and cited directly in the manuscript. Therefore, we did not make correction on this issue.

Additionally, we have provided more information (mailing address of suppliers) in the "suppliers" listed after "References".

**Supplier: After the References section, provide a Suppliers list with contact information (names and complete mailing addresses) for manufacturers of devices and other nondrug products used directly in a study (ie, do not provide such information for products not directly used in your research but mentioned in studies you cite). Identify equipment and/or materials in text, tables, and legends by superscript lower case letters. List suppliers consecutively in the order they are mentioned in the text.**

Lines 128-9, should mention which manufacturer's needle was used and then similar to correction on line 15 the authors can get rid of "b" superscript

**Response:**

According to the "Instruction for Authors" (listed below), the suppliers should not be put and cited directly in the manuscript. Therefore, we did not make correction on this issue.

Line 163 need to also cross out (eliminate) the "in Group IV animals"

**Response:**

We have deleted these words according to your suggestion.

Line 172, put manufacturer information in and eliminate the "c"

**Response:**

According to the "Instruction for Authors" (listed below), the suppliers should not be put and



cited directly in the manuscript. Therefore, we did not make correction on this issue.

Line 201, how was the needle fixed to the skin

**Response:**

It was carefully and firmly taped on the skin. This sentence has been changed into "Then the needle was fixed in place (**carefully and firmly taped on the skin**) to ensure that this EPN can run continuously on the recording screen with constant amplitudes."

Lines 210-213, "Five randomly selected samples of EPN recordings (10 ms each) were taken before, during, and 3 min after the completion of the needling treatment for all groups, and also taken before and 30 minutes after surgery for Group II animals, and every 30 minutes up to 120 minutes after surgery for Group III and IV animals" would be better worded "Five randomly selected samples of EPN recordings (10 ms each) were taken before, during, and 3 min after the completion of the needling treatment for all groups; before and 30 minutes after surgery for Group II animals; and every 30 minutes up to 120 minutes after surgery for Group III and IV animals."

**Response:**

We have revised the above statements according to your suggestion.

Line 246, put manufacturer in and eliminate the "d"

**Response:**

According to the "Instruction for Authors" (listed below), the suppliers should not be put and cited directly in the manuscript. Therefore, we did not make correction on this issue.

Line 265, should also remove "treated differently"

**Response:**

We had removed these two words according to your suggestion.

Line 276-280 sentence-not sure it adds much new information that isn't already presented in prior sentence and Figure 3, and same for sentence lines 282-4

**Response:**

We would like to emphasize the important findings of "the initial increase followed by the subsequent decrease in EPN amplitudes after IDN or CDN, but not after ISN or CSN", although these findings could be identified in Figure 3. Therefore, we prefer to keep these statements.

Information in sentence lines 284-7 could have been incorporated in lines 279-80 to make this result section more succinct

**Response:**

Based on your suggestion, these sentences have been merged as listed below:

"There were significant differences in EPN amplitudes recorded either during or after needling between IDN and ISN, or between CDN and CSN subgroups (Bonferroni post-hoc test,  $P < .05$ ), but not between IDN and IDN subgroups (Bonferroni post-hoc test,  $P > .05$ )".

Lines 324-327 sentence-again seems to be repeating same information as prior sentences in a slightly different way

**Response:**

These statements have been merged to the previous sentence as listed below:

" There were significant differences in EPN amplitudes among those recorded before, during, and after needling (repeated measures ANOVA,  $F = 80.77$ ,  $P < .05$ ) in CDN subgroup (**similar to the changes in Group I, Bonferroni post-hoc test,  $P < .05$** ), but not in IDN subgroup (repeated measures ANOVA,  $F = 2.89$ ,  $P > .05$ ).".

Information in lines 334-336 could have been incorporated in the 329-332 sentence to make this results section more succinct

**Response:**

These statements have been merged to the previous sentence as listed below:

"There were significant differences in mean EPN amplitudes between CDN and its comparable CSN subgroups, **but not between IDN and ISN subgroups (Bonferroni post-hoc test,  $P > .05$ )**, at the time during (Bonferroni post-hoc test,  $P < .05$ ) and after (Bonferroni post-hoc test,  $P < .05$ ) needling"

Lines 360-363 sentence-I don't think it is necessary information to report and does not add to the results

**Response:**

Regarding the result of "During the 2 h period after transection, the mean EPN amplitudes were significantly lower than the pre-transected levels", **we have made discussion about this issue in the section of " Electrophysiological Findings of the Remote Effect after Interruption of Certain Neural Circuits " and considered that this could be related to spinal shock. Therefore, we prefer to keep this.**

Lines 376-381 - sentence information could be merged to be more succinct

**Response:**

These statements have been merged as listed below:

"There were no significant differences in each time-dependent alterations of EPN amplitude between IDN and ISN subgroups, **between IDN and CDN subgroups**, or between CDN and CSN

subgroups (Bonferroni post-hoc test, all  $P > .05$ ).".

Lines 399-406 sentences the information could be merged to reflect the EPN amplitudes normalized 2 hours post cord transaction to be similar for all groups

**Response:**

These statements have been merged as listed below:

". . . but recovered to pre-transection level after 90 min (Bonferroni post-hoc tests, all  $P > .05$ ), and normalized to be similar for all subgroups by 120 min after transaction (two-way ANOVA,  $F=0.09$ ,  $P > .05$ ).".

Line 421-423-is this information necessary

**Response:**

This sentence has been deleted.

Lines 423-425 information could be incorporated in lines 416-417

**Response:**

This statement has been merged to the previous sentence as listed below:

". . . but there were no significant differences in these changes between IDN and CDN subgroups (Bonferroni post-hoc test,  $P > .05$ ). "

Lines 452-465-not sure how much this information adds to purpose of paper, consider deleting

**Response:**

**We try to emphasize the importance of this special technique (to elicit LTRs). If the experiment is not carried with this technique, we would not be able to find the remote effects of needling. Furthermore, both LTRs and neural control of needling are related to the spinal cord mechanism. The spinal cord mechanism of dry needling can change EPN activity and achieve the remote needling effect. Therefore, we prefer to keep this section.**

Lines 467-468-don't need to put subtitle in, and same for lines 481-482 and 514

**Response:**

We have deleted those subtitles per your suggestion.

ORIGINAL ARTICLE

**Spinal Cord Mechanism Involving the Remote Effects of Dry Needling on the Irritability of Myofascial Trigger Spots in Rabbit Skeletal Muscle**

**Yueh-Ling Hsieh, PT, PhD, Li-Wei Chou, MD, MSc, Yie-San Joe, MD,**

**Chang-Zern Hong, MD**

*Chou LW contributed equally to the first author in this work.*

From the Department of Physical Therapy and Graduate Institute of Rehabilitation Science (Hsieh, Chou), and School of Chinese Medicine, College of Chinese Medicine (Chou), China Medical University, Taichung; Department of Physical Medicine and Rehabilitation, China Medical University Hospital, Taichung (Chou); Department of Physical Medicine and Rehabilitation, Cheng Ching Hospital, Taichung (Joe); College of Life Science, National Chung Hsing University, Taichung (Joe); Department of Physical Therapy, Hungkuang University, Taichung (Hong), Taiwan.

**Word counts: Abstract : 245, Text: 4088.**

**Correspondence and reprints availability from: Chang-Zern Hong, M.D.,**

Department of Physical Therapy, Hungkuang University, 34 Chung-Chie Road, Shalu,  
Taichung, Taiwan.

E-mail: [johnczhong@yahoo.com](mailto:johnczhong@yahoo.com).

**Running title: NEURAL MECHANISM FOR REMOTE EFFECT OF DRY  
NEEDLING**

This study was supported by a grant from National Science Council (NSC  
99-2314-B241-001) and China Medical University (CMU97-188), Taiwan.

No commercial party having a direct financial interest in the results of the research  
supporting this article has or will confer a benefit upon the authors or upon any  
organization with which the authors are associated.

**Devices**

- a. **Thermometer:** Physitemp Instruments, Inc. 154, Huron Avenue, Clifton, New  
Jersey, USA.
- b. **Digital EMG machine:** Neuro-EMG-Micro; Neurosoft, 5, Voronin Str, Ivanovo,  
Russia.

1 *ORIGINAL ARTICLE*

2 **Spinal Cord Mechanism Involving the Remote Effects of Dry Needling on the**

3 **Irritability of Myofascial Trigger Spots in Rabbit Skeletal Muscle**

4

5

6 **Running title: NEURAL MECHANISM FOR REMOTE EFFECT OF DRY**

7 **NEEDLING**

8

9 **ABSTRACT**

10

11 **Objective:** The purpose of this study is to investigate the possible neural pathway  
12 for the remote effects of dry needling based on the assessment of endplate noise (EPN)  
13 recorded for the myofascial trigger spot (MTrS) in rabbit skeletal muscle.  
14 To elucidate the neural mechanisms underlying the remote effects produced by dry  
15 needling rabbit skeletal muscle myofascial trigger spots (MTrSs) via analyses of their  
16 endplate noise (EPN) recordings.

17 **Design:** Experimental animal controlled trial.

18 **Setting:** An animal laboratory of a university.

19 **Animals:** Ninety-six male New Zealand rabbits (body weight: 2.5–3.0 kg, age: 16–20  
20 weeks).

21 **Intervention:** Animals received no intervention for neural interruption in Group I,  
22 transection of the tibial nerve in Group II, transection of L5 and L6 spinal cord in  
23 Group III, and transection of the T1 and T2 spinal cord in Group IV. Each group was  
24 further divided into 4 subgroups: animals received ipsilateral dry needling (IDN),  
25 contralateral dry needling (CDN), ipsilateral sham needling (ISN), or contralateral  
26 sham needling (CSN), on the MTrS of the gastrocnemius of gastrocnemius (GAS)  
27 MTrSs.

28 **Main Outcome Measures:** Amplitudes of EPN amplitudes on the MTrS region of

29 biceps femoris (BF) MTrSs. EPN amplitudes of biceps femoris (BF) MTrSs.

30 **Results:** Mean EPN amplitudes significantly increased ( $P < .05$ ) initially, but reduced

31 to a level significantly lower ( $P < .05$ ) than the pre-needling level in Group I with IDN

32 or CDN, Group II with CDN (but not IDN), and Group IV with IDN or CDN, but no

33 such changes were observed in Group III. There were no significant changes ( $P > .05$ )

34 in EPN amplitudes in all control animals. BF MTrS mean EPN amplitudes

35 significantly increased ( $P < .05$ ) initially after GAS verum needling, but reduced to a

36 level significantly lower ( $P < .05$ ) than the pre-needling level in Groups I and IV with

37 IDN or CDN, and in Group II with CDN (but not IDN). No significant EPN

38 amplitude changes were observed in BF MTrS in Group III or in the control animals

39 receiving superficial needling (sham).

40 **Conclusion:** This remote effect of dry needling depends on an intact afferent pathway

41 from the stimulating site to the spinal cord and a normal spinal cord function at the

42 levels corresponding to the innervations of the proximally affected muscle.

43

44 **Key Words:** Neural pathway; Dry needling; Myofascial trigger spot;

45 Electromyography; Remote effect.

46



47 **DEFINITION OF ACRONYMS**

48 BF = biceps femoris muscle;

49 CDN = contralateral dry needling;

50 CSN = contralateral sham needling;

51 EMG = electromyographic;

52 EPN = endplate noise;

53 GAS = gastrocnemius muscle;

54 IDN = ipsilateral dry needling;

55 ISN = ipsilateral sham needling;

56 LTR = local twitch response;

57 MTrP = myofascial trigger point (human);

58 MTrS = myofascial trigger spot (rabbit)

59

## INTRODUCTION

60

61 Myofascial pain is one of the most common musculoskeletal pain associated  
62 with myofascial trigger points (MTrPs) which are hyperirritable spots in taut bands of  
63 skeletal muscle fibers due to accumulation of unique hypersensitive loci. Myofascial  
64 pain is one of the most common sources of musculoskeletal pain and has as its  
65 hallmark the presence of taut bands and small hyperirritable regions referred to as  
66 myofascial trigger points (MTrPs).<sup>1,2</sup> Clinically, an MTrP is characterized with a  
67 typical referred pain pattern and a local twitch response (LTR) in response to snapping  
68 palpation. Clinically, a given MTrP has a characteristic referred pain pattern, and may  
69 be associated with a local twitch response (LTR) in response to snapping palpation.<sup>2</sup>  
70 In the MTrP region, electromyographic (EMG) activity of endplate noise (EPN) can  
71 be recorded, and both prevalence and amplitude of EPN can be used as indicators to  
72 assess the irritability of MTrP.<sup>2-5</sup>

73 Dry needling of MTrP to alleviate myofascial pain has long been established  
74 and widely used in treating patients. Dry needling has a well-established role in the  
75 treatment of myofascial pain.<sup>6-9</sup> In addition to direct needling of the painful MTrP,  
76 clinical studies have demonstrated suppressive effect on this MTrP with dry needling  
77 at a remote MTrP, either proximally or distally located.<sup>8,9-12</sup> Similar remote  
78 effectiveness in pain control has also been documented in acupuncture therapy.<sup>4, 13-14</sup>

79 Clinical studies have demonstrated that dry needling MTrPs at the pain site suppresses  
80 their activity resulting in pain reduction, but also that those MTrPs can also be  
81 suppressed through dry needling distant MTrPs that are anatomically proximal or  
82 distal to the site of clinical pain.<sup>8-12</sup> Acupuncture studies have demonstrated that the  
83 Traditional Chinese Medicine principle that pain conditions can be improved by  
84 stimulating acupuncture points distant from the site of pain.<sup>4,13-14</sup> The effects of  
85 acupuncture may also spread to the contralateral side.<sup>15</sup> Studies of acupuncture needle  
86 stimulation in anesthetized animals have identified a wide variety of reflex responses  
87 in remote modification of various organ functions.<sup>16</sup> However, its underlying neuronal  
88 control mechanism remains unclear.

89 ~~The purpose of this study is to confirm the remote effects of dry needling and to~~  
90 ~~investigate the possible neural pathway for the remote effects of dry needling of the~~  
91 ~~myofascial trigger spot (MTrS) in the rabbit gastrocnemius (GAS) muscle based on~~  
92 ~~the assessment of EPN recorded for the MTrS in the biceps femoris (BF) muscle. The~~  
93 ~~animal model with myofascial trigger spots (MTrS, equivalent to human MTrP) has~~  
94 ~~been established previously,<sup>17,18</sup> and was used in our study.~~

95 The purpose of this study is to confirm the remote effects of dry needling on  
96 trigger points and to elucidate the neural mechanisms underlying the remote effects  
97 produced by dry needling rabbit myofascial trigger spots (MTrSs, equivalent to

98 human MTrPs) in the gastrocnemius muscle (GAS)<sup>17, 18</sup> via analyses of EPN

99 recordings from the biceps femoris (BF).

100

## MATERIALS AND METHODS

101

### 102 **Animals**

103 The experiments were performed on adult male New Zealand rabbits (age ages  
104 from 16 to 20 weeks, body weight of 2.5–3.0 kg). Each animal was housed and cared  
105 for following the ethical guidelines of the International Association for Study of Pain  
106 in animals were followed.<sup>19</sup> Effort was made to minimize discomfort and to reduce  
107 the number of animals used. All animal experiments were conducted with the  
108 procedure approved by the Animal Care and Use Committee of a university in  
109 accordance with the Guidelines for Animal Experimentation.

110 Ninety-six rabbits were divided randomly into four groups (fig 1) based on the  
111 procedure performed. Group I (n=24) animals received no surgical intervention (intact  
112 neural pathway), Group II animals (n=24) received transection of tibial nerve in the  
113 electrophysiologically investigated side (peripheral sensory pathway), Group III  
114 animals (n = 24) received transection of L5-L6 spinal cord (BF innervation level), and  
115 Group IV animals (n=24) received transection of T1-T2 spinal cord (supra-segment of  
116 BF innervation). For the EPN amplitude variable, a sample size of 24 subjects in each  
117 group was sufficient to give statistical power of 97.06% with a significance level of P  
118 < .05. Animals in each group were randomly divided further into four subgroups  
119 based on the condition of treatment on GAS: experimental animals with ipsilateral dry

120 needling (IDN, n=8), or contralateral dry needling (CDN, n=8), control animals with  
121 ipsilateral sham needling (ISN, n=4), or contralateral sham needling (CSN, n=4), on  
122 the MTrS of the gastrocnemius. Fewer animals were studied in the control group since  
123 no significant changes were observed in all animals treated with sham needling.  
124 Regarding the assignment of groups or subgroups, animals were selected from the  
125 first available litter, and subsequently from the next litter, and so forth according to  
126 the sequence in a random table.

## 127 **Animal Preparation**

128 Before anesthesia, the tenderest spots (i.e., MTrS) of BF and GAS were  
129 identified by finger pinching. The animal responded to pinch stimulation with  
130 withdrawal of the lower limb, turning its head, screaming, etc, only when the most  
131 painful spot was pinched, and this most tender spot was confirmed as the MTrS.  
132 <sup>2,17,18,20-23</sup> These painful regions were marked on the skin ~~carefully~~ with an indelible  
133 marker and were designated for electrophysiological assessment or dry needling. The  
134 animals were anesthetized with 2% isoflurane in oxygen flow for induction followed  
135 by a 0.5% maintenance dose.<sup>24</sup> Body temperature was monitored by a thermistor  
136 probe of a **thermometer**<sup>a</sup> in the rectum and maintained at approximately 37.5 °C using  
137 a body temperature control system consisting of thermostatically regulated DC current  
138 heating pad and an infrared lamp. The hind limbs of anesthetized rabbits were shaved

139 and cleaned with povidone-iodine solution. The skin of the lateral thigh in one  
140 randomly selected side was incised to expose the BF, which served as an EPN  
141 recording site. The marked spot region in the BF muscle was grasped between two  
142 fingers from behind the muscle and the muscle palpated by gently rubbing (rolling) it  
143 between the fingers to discover a taut band. A taut band felt like a clearly delineated  
144 "rope" of muscle fibers and was roughly 2–3 mm or more in diameter. The fibers of  
145 the taut band were unmistakably firmer in consistency than the surrounding muscle.

#### 146 **Needling of Gastrocnemius Muscle**

147 All needling procedures were performed by the same investigator who was  
148 blinded to the group assignment regarding to surgical intervention on neural pathway.  
149 Dry needling stimulation was performed with a disposable 30G acupuncture needle  
150 (300  $\mu$ m in diameter, 1.5 inches in length) at ipsilateral or contralateral GAS (fig 2).  
151 The technique of dry needling was similar to that suggested by Hong<sup>18, 20, 25-27</sup> with  
152 multiple needle insertions to elicit rabbit-LTRs as much as possible. For needling in  
153 MTrS of GAS, the needle was first inserted through the skin perpendicularly at the  
154 center of the marked spot and advanced slowly and gently into the muscle until the  
155 needle tip touched the bone surface to estimate the thickness of the muscle. The  
156 needle was withdrawn back to the subcutaneous layer, and rapidly moved in and out  
157 for insertion of multiple sites in different directions (in a cone shape with the center at

158 the initial needle insertion of a perpendicular direction, and the angle of the cone  
159 margin was about 20°). For each needle insertion, the needle was advanced into the  
160 depth near the bone surface. Simultaneous needle rotation was performed to facilitate  
161 fast “in-and-out” needle movement as suggested by Chou et al.<sup>4</sup> in order to elicit as  
162 many LTRs as possible. For sham needling, the needle was inserted into the  
163 subcutaneous layer of the marked MTrP region at a depth approximately 1-2 mm from  
164 the skin surface. After insertion, the needle stayed there without further movement.

165

## 166 **Transection Operations**

### 167 **1. Transection of tibial nerve**

168 During anesthesia for the animals in Group II, the incision was made over the  
169 posterior aspect of one thigh ipsilaterally to the EPN recording side. Under the  
170 operating microscope, the sciatic nerve was exposed, and the tibial nerve isolated and  
171 transected at the site about 1 cm from its insertion into the GAS.

### 172 **2. Transection of spinal cord**

173 After completing laminectomy and making a slit in the dorsal portion of the dura  
174 mater, the cord was transected by a knife and then aspirated by suction at about 2 mm  
175 caudal and rostral to the level of transection, at L5-L6 levels of the spinal cord for  
176 animals in Group III, or at T1-T2 levels of the spinal cord for Group IV animals.



177 Gelfoam was placed into the empty vertebral column to seal the empty vertebral  
178 cavity and reduce bleeding. In a previous study<sup>20</sup> and in our preliminary data, about  
179 2½ h after surgery, the rabbits would have almost completely recovered from spinal  
180 shock ~~in Group IV animals~~. The animal would then be ready for the needling study.

181

## 182 **Recording of Endplate Noise**

### 183 **1. Electromyography setting**

184 For EPN assessment, a two-channel digital EMG machine<sup>c</sup> and monopolar  
185 needle electrodes (37 mm disposable Teflon-coated model) were used. The gain was  
186 set at 20µV per division for recordings from both channels. Low-cut frequency filter  
187 was set at 100 Hz and the high-cut at 1,000 Hz. Sweep speed was 10 ms per division.  
188 The search needle for EPN recording was inserted into the MTrS region and  
189 connected to the first channel of the EMG machine. The control needle was inserted  
190 into the non-taut band region near the MTrS in the same muscle and connected to the  
191 second channel. A common reference needle electrode for each channel was placed on  
192 the incised skin and connected to both channels via a y-connector.

### 193 **2. Search for endplate noise**

194 This procedure was performed by an investigator who was blind to the group  
195 assignment. The search needle was inserted into the MTrS region in a direction

196 parallel to the muscle fibers at an angle of approximately  $60^\circ$  to the surface of the  
197 muscle. After initial insertion just short of the depth of the MTrS or to a comparable  
198 depth in the case of control sites, the needle was advanced very slowly with  
199 simultaneous slow rotation to prevent it from 'grabbing' and releasing the tissue  
200 suddenly to advance in a large jump. Each advance was of minimal distance ( $\sim 1$  mm).  
201 When the needle approached an active locus (EPN locus), the continuous distant  
202 electrical activity, i.e., EPN, can be heard. As soon as EPN with amplitude higher than  
203  $10 \mu\text{V}$  could be recorded, the examiner stopped advancing the needle, but minimally  
204 moving the needle gently to different direction, trying to obtain EPN with highest  
205 amplitude. If this was impossible, the needle was advanced to another site until an  
206 EPN with optimal amplitude (usually higher than  $30\mu\text{V}$ ) could be recorded. Then the  
207 needle was fixed in place (carefully and firmly taped on the skin) to ensure that this  
208 EPN can run continuously on the recording screen with constant amplitudes.  
209 Continuous EPN tracing was recorded throughout the entire course of the experiment,  
210 and provided the opportunity for continuous visual observation of EPN changes on  
211 the EMG screen. If the EPN could not be sustained, the searching needle would be  
212 moved to another site until a satisfactory EPN tracing could be obtained. The entire  
213 EPN tracing found in MTrS of BF were recorded for the analysis of amplitude  
214 changes.

### 215 **3. Measurement of the amplitude of endplate noise**

216 Five randomly selected samples of EPN recordings (10 ms each) were taken  
217 before, during, and 3 min after the completion of the needling treatment for all groups;  
218 ~~and also taken~~ before and 30 minutes after surgery for Group II animals; and every 30  
219 minutes up to 120 minutes after surgery for Group III and IV animals. The mean  
220 amplitude of EPN of 5 random samples was analyzed and calculated through the  
221 embedded software in the Neuro-EMG-Micro equipment, and was recorded as the  
222 value for a certain measurement point for each animal.

223

### 224 **Data Analysis**

225 Data of EPN amplitudes in different measurement points for different groups or  
226 subgroups were expressed as the mean  $\pm$  standard error of the mean (SEM) for further  
227 statistical analysis. The Shapiro-Wilk's normality test was conducted to determine  
228 whether the data fit a normal distribution prior to subsequent analyses, and showed all  
229 measures of EPN amplitude were normally distributed. Tests of 'homogeneity' or  
230 'baseline balance' on covariates including body weight, age, and anesthesia condition  
231 were measured and equivalent before the needling treatment in all animals. The  
232 differences in EPN amplitude across measurement points in each group were carried  
233 out using repeated measures ANOVA, and later further analyzed by a Bonferroni

234 post-hoc method. The differences in EPM amplitudes within each of the subgroups  
235 (IDN, CDN, ISN and CSN) and across measurement points (before, during, and after  
236 needling) were analyzed using two-way ANOVA (side × time) followed by a  
237 Bonferroni post-hoc analysis for each group. The differences in EPN amplitude within  
238 measurement point (before, during, and after needling) across subgroups (IDN, CDN,  
239 ISN and CSN) were tested by paired t-test. A p value of <0.05 was considered to be  
240 statistically significant. All data was analyzed using SPSS version 10.0 for Windows.<sup>d</sup>  
241

242

## RESULTS

### 243 **Effects of Dry Needling of Distal MTrS in Intact Rabbits (Group I)**

244 The serial alterations of the mean EPN amplitude before, during, and after dry  
245 needling at ipsilateral and contralateral GAS for Group I are demonstrated in figure 3.  
246 Before needling treatment, there was no significant difference among the four  
247 subgroups ~~treated differently~~ (two-way ANOVA,  $F=0.10$ ,  $P> .05$ ).

248 The mean amplitudes of EPN before, during, and after needling were  
249  $18.20\pm 0.70\mu\text{V}$ ,  $27.71\pm 0.47\mu\text{V}$ , and  $13.15\pm 0.59\mu\text{V}$ , respectively in IDN subgroup, and  
250  $17.96\pm 0.69\mu\text{V}$ ,  $24.66\pm 1.47\mu\text{V}$ , and  $14.01\pm 0.86\mu\text{V}$ , respectively in CDN subgroup. In  
251 either IDN or CDN subgroup, the amplitudes at different times were significantly  
252 different (repeated measures ANOVA:  $F=45.99$  and  $P< .05$  for IDN,  $F=113.98$  and  
253  $P< .05$  for CDN). Compared with the data in the pre-needling level, the EPN  
254 amplitudes were significantly increased during the dry needling (Bonferroni post-hoc  
255 test,  $P< .05$ ), and then significantly decreased to a much lower level after completion  
256 of the needling treatment (Bonferroni post-hoc test,  $P< .05$ ) for either IDN or CDN  
257 subgroup as shown in figure 3. However, these serial alterations of EPN amplitudes  
258 were not found in the comparable subgroup ISN or CSN (repeated measures ANOVA,  
259  $P> .05$ ). There were significant differences in EPN amplitudes recorded either during  
260 or after needling between IDN and ISN, or between CDN and CSN subgroups

261 (Bonferroni post-hoc test,  $P < .05$ ), but not between IDN and IDN subgroups. In  
262 addition, alterations in EPN amplitudes in IDN subgroup were similar to those in CDN  
263 subgroup. The magnitude or time dependent alteration of EPN amplitude after CDN  
264 was not significantly different from that after IDN (Bonferroni post-hoc test,  $P > .05$ ).

265

## 266 **Effects of Dry Needling of Distal MTrS in Rabbits With Tibial Nerve** 267 **Transection (Group II)**

268 The serial alterations of the mean EPN amplitude ( $\pm$ SEM) throughout the entire  
269 experiment in Group II are demonstrated in figure 4. The mean amplitude of EPN had  
270 no significant changes before, during, and after ipsilateral tibial nerve transection (i.e.,  
271 GAS denervation) (repeated measures ANOVA,  $F=0.06$ ,  $P > .05$ ). Before the needling  
272 treatment, there was no significant difference in EPN amplitude among the subgroups  
273 treated with dry or sham needling at ipsilateral or contralateral side (two-way ANOVA,  
274  $F=0.68$ ,  $P > .05$ ).

275 The mean amplitudes ( $\pm$ SEM) of EPN recorded from BF before, during, and after  
276 needling were  $16.72 \pm 0.34 \mu\text{V}$ ,  $16.64 \pm 0.37 \mu\text{V}$ , and  $15.46 \pm 0.50 \mu\text{V}$ , respectively in IDN  
277 subgroup, and were  $16.90 \pm 0.38 \mu\text{V}$ ,  $21.63 \pm 0.91 \mu\text{V}$ , and  $12.40 \pm 0.36 \mu\text{V}$ , respectively in  
278 CDN subgroup. There were significant differences in EPN amplitudes among those  
279 recorded before, during, and after needling (repeated measures ANOVA,  $F= 80.77$ ,

280  $P < .05$ ) in CDN subgroup (similar to the changes in Group I, Bonferroni post-hoc test,  
281  $P < .05$ ), but not in IDN subgroup (repeated measures ANOVA,  $F = 2.89$ ,  $P > .05$ ). In  
282 the CDN subgroup, the EPN amplitudes were increased during needling (Bonferroni  
283 post hoc test,  $P < .05$ ). However, 3 min later and after cessation of needling, it was  
284 reduced significantly to a level less than that before needling (Bonferroni post hoc test,  
285  $P < .05$ ). The EPN amplitudes also had no significant changes after ISN (repeated  
286 measures ANOVA,  $F = 0.10$ ,  $P > .05$ ) or after CSN (repeated measures ANOVA,  
287  $F = 0.02$ ,  $P > .05$ ). There were significant differences in mean EPN amplitudes between  
288 CDN and its comparable CSN subgroups, but not between IDN and ISN subgroups  
289 (Bonferroni post-hoc test,  $P > .05$ ), at the time during (Bonferroni post-hoc test,  $P < .05$ )  
290 and after (Bonferroni post-hoc test,  $P < .05$ ) needling. Moreover, there were significant  
291 differences in the magnitude or time-dependent alterations of EPN amplitude between  
292 CDN and CSN subgroups (Bonferroni post-hoc test,  $P < .05$ ). However, there were no  
293 significant differences between IDN and ISN subgroups at the time course either  
294 during (Bonferroni post hoc test,  $P > .05$ ) or after (Bonferroni post hoc test,  $P > .05$ )  
295 treatment at ipsilateral GAS.

296

297 **Effects of Dry Needling of Distal MTrS in Rabbits With Lumbar**

298 **Cord Transection (Group III)**

299 The serial alterations of the mean EPN amplitude ( $\pm$ SEM) throughout the entire  
300 experiment for Group III are shown in figure 5. There were significant differences in  
301 EPN amplitude among those recorded before, immediately, and 30, 60, 90, and 120  
302 min after L5-L6 transection (repeated measures ANOVA,  $F=29.81$ ,  $P < .05$ ). During  
303 the 2 h period after transection, the mean EPN amplitudes were significantly lower  
304 than the pre-transected levels (Bonferroni post-hoc test, all were  $P < .05$  at 30, 60, 90,  
305 and 120 h). There was no significant difference among the four subgroups treated with  
306 dry or sham needling at the ipsilateral or contralateral side regardless of the time  
307 before (two-way ANOVA,  $F=0.23$ ,  $P > .05$ ), during ( $F=1.45$ ,  $P > .05$ ), or after ( $F=1.72$ ,  
308  $P > .05$ ) needling treatments.

309 The mean amplitudes ( $\pm$ SEM) of EPN before, during, and after needling were  
310  $11.56 \pm 0.36 \mu\text{V}$ ,  $11.47 \pm 0.43 \mu\text{V}$ , and  $11.28 \pm 0.47 \mu\text{V}$ , respectively in IDN subgroup, and  
311 were  $11.67 \pm 0.45 \mu\text{V}$ ,  $12.32 \pm 0.46 \mu\text{V}$ , and  $12.33 \pm 0.46 \mu\text{V}$ , respectively in CDN subgroup.  
312 There was no significant difference in EPN amplitude among those recorded before,  
313 during, and after IDN (repeated measures ANOVA,  $F=0.63$ ,  $P > .05$ ), CDN (repeated  
314 measures ANOVA,  $F=1.17$ ,  $P > .05$ ), ISN treatment (repeated measures ANOVA,  
315  $F=0.23$ ,  $P > .05$ ), or CSN (repeated measures ANOVA,  $F=0.52$ ,  $P > .05$ ). There were  
316 no significant differences in each time-dependent alterations of EPN amplitude  
317 between IDN and ISN subgroups, between IDN and CDN subgroups, or between CDN



318 and CSN subgroups (Bonferroni post-hoc test, all  $P > .05$ ). There were also no  
319 significant differences in each time dependent alteration of EPN amplitude between  
320 IDN subgroup and CDN subgroup (Bonferroni post hoc test, all were  $P > .05$  at each  
321 recording time).

322

### 323 **Effects of Dry Needling of Distal MTrS in Rabbits With Thoracic** 324 **Cord Transection (Group IV)**

325 The serial alterations of the mean EPN amplitude ( $\pm$ SEM) throughout the entire  
326 experiment for Group IV are shown in figure 6. The EPN amplitude recorded at 30  
327 and 60 min after T1-T2 transection was significantly reduced when compared to the  
328 pre-transection level (Bonferroni post-hoc test,  $P < .05$ ), but recovered to  
329 pre-transection level after 90 min (Bonferroni post-hoc tests, all  $P > .05$ ), and  
330 normalized to be similar for all subgroups by 120 min after transaction (two-way  
331 ANOVA,  $F=0.09$ ,  $P > .05$ ). The EPN amplitude recorded at 90 and 120 min recovered  
332 to the level with no significant differences from pre-transection level (Bonferroni  
333 post hoc tests, all were  $P > .05$  at 90 and 120 min recording time). Before needling  
334 treatment, there was no significant difference among all four subgroups treated with  
335 dry or sham needling at the ipsilateral or contralateral side (two way ANOVA,  $F=0.09$ ,  
336  $P > .05$ ).

337

338       The mean amplitudes ( $\pm$ SEM) of EPN recorded from BF before, during, and after  
339 needling were  $18.17\pm 0.36\mu\text{V}$ ,  $26.88\pm 0.43\mu\text{V}$ , and  $15.74\pm 0.26\mu\text{V}$ , respectively in IDN  
340 subgroup, and were  $18.28\pm 0.45\mu\text{V}$ ,  $27.72\pm 0.47\mu\text{V}$ , and  $16.20\pm 0.22\mu\text{V}$  respectively in  
341 CDN subgroup. There were significant differences in EPN amplitudes among those  
342 recorded before, during, and after IDN (repeated measures ANOVA,  $F= 264.29$ ,  
343  $P < .05$ ) or CDN (repeated measures ANOVA,  $F= 243.11$ ,  $P < .05$ ). The mean EPN  
344 amplitudes were significantly increased during IDN or CDN (Bonferroni post-hoc test,  
345  $P < .05$ ), but after cessation of needling, they reduced significantly to a level less than  
346 that before needling (Bonferroni post-hoc test,  $P < .05$ ), but there were no significant  
347 differences in these changes between IDN and CDN subgroups (Bonferroni post-hoc  
348 test,  $P > .05$ ). These serial alterations of EPN amplitudes were not found in either ISN  
349 (repeated measures ANOVA,  $F=1.02$ ,  $P > .05$ ) or CSN subgroup (repeated measures  
350 ANOVA,  $F=2.02$ ,  $P > .05$ ). There were significant differences between dry needling  
351 and its comparable sham needling subgroups (Bonferroni post hoc test,  $P < .05$ ).  
352 Moreover, there were no significant differences in the magnitude or time dependent  
353 alterations of EPN amplitude between CDN and IDN subgroups (Bonferroni post hoc  
354 test,  $P < .05$ ).

355

## DISCUSSION

356

357 To our knowledge, the present study is the first animal study to investigate the  
358 neural mechanism of the remote effects of dry needling. In this study, we found that  
359 an intact afferent nerve from the remote stimulation site and normal spinal cord  
360 segments corresponding to the innervation of the affected proximal muscle are  
361 essential for the remote effect from either ipsilateral or contralateral stimulation.

### 362 **Technical Issues on Dry Needling**

363 The dry needling used in this study is a technique of MTrP injection with  
364 multiple high-speed needle insertions into different sensitive loci in an MTrP region  
365 suggested by Hong.<sup>25-27</sup> High speed needling can provide high-pressure stimulation to  
366 the sensitive loci in the MTrP region to elicit LTRs. It is essential to elicit LTRs  
367 during needling of an MTrP in order to obtain immediate and complete pain relief.<sup>6,</sup>  
368 <sup>25-29</sup> Dry needling at the MTrS was effective in diminishing spontaneous electrical  
369 activity (i.e., EPN) of MTrS of rabbit skeletal muscle if LTRs were elicited.<sup>30</sup> After  
370 several LTRs had been elicited by the needling of an MTrS of rabbit skeletal muscle,  
371 no more LTRs could be elicited from the same region<sup>18</sup> and the irritability of the  
372 MTrS could be suppressed.<sup>30</sup> Needling-elicited LTRs are involuntary discharges of  
373 muscle fiber mediated through the nervous system and integrated at the spinal cord  
374 level.<sup>18,20</sup> Therefore, it is important to apply this needling technique to achieve the

375 best needling effect or remote needling effect for the study on the neural mechanism.

376

### 377 **Electrophysiological Confirmation of the Remote Effect in Normal** 378 **Neural Circuits**

379 Changes in the EPN amplitude in the MTrS were found during and after dry  
380 needling at the distal MTrSs in animals with intact neural circuits (Group I). It appears  
381 that either IDN or CDN to the distal MTrS could initially increase the irritability of  
382 the proximal MTrS, followed by a suppression effect after cessation of needling.  
383 Fernandez-Camero, et al.<sup>31</sup> also found an increase in spontaneous electrical activity at  
384 an MTrP region during a persistent noxious stimulation at another distant MTrP,  
385 followed by a suppression of electrophysiological irritability after cessation of  
386 needling. The two findings above strongly support clinical observations related to the  
387 interaction between one MTrP and another MTrP located in the region of the referred  
388 pain (referred zone) of that MTrP.<sup>2,6,9-12,26-29</sup>

### 389 **Electrophysiological Findings of the Remote Effect after Interruption** 390 **of Certain Neural Circuits**

391 In the study on Group II rabbits with ipsilateral denervation of GAS, the remote  
392 effect disappeared after IDN, but persisted after CDN. These results demonstrated the  
393 importance of an intact afferent pathway to the spinal cord in the remote modulation

394 of EPN amplitudes. After destruction of spinal cord corresponding to the level of BF  
395 (Group III), the remote effect disappeared after either IDN or CDN treatment. This  
396 finding suggested the existence of intraspinal connections between the GAS afferents  
397 and BF spinal interneurons. The partial suppression of the EPN amplitude after spinal  
398 cord transection at L5-L6 is possibly related to the influence of the spinal shock. After  
399 interruption of upper motor neuron and supra-sensory connections (Group IV), the  
400 remote effect persisted, either with IDN or CDN, but smaller than that in intact  
401 animals (Group I). It may imply the possible influences from supraspinal centers, such  
402 as descending pain inhibitory systems. These influences are anticipated to be minimal  
403 after transection at higher spinal levels. Loss of inputs to this system would weaken  
404 the inhibition on the pain level,<sup>32</sup> which could lead to the EPN amplitude being less  
405 suppressed. Therefore, the recruitment of the diffuse noxious inhibitory control  
406 (DNIC) system may be also elicited by dry needling treatment on regions remote to  
407 the stimulation site.

#### 408 **Possible Neural Control for the Remote Effect of Dry Needling**

409 The neural pathway for the remote effect appears to be a spinal reflex, probably  
410 similar to that mediating the referred pain<sup>26, 29</sup> and local twitch response.<sup>18, 20, 26, 29</sup>  
411 Hong has hypothesized a corresponding “MTrP circuit” for each MTrP, which can  
412 modulate the pain, referred pain, and local twitch response elicited by stimulating the

413 MTrP.<sup>6, 29, 33</sup> The neural connection in the spinal cord responsible for this remote

414 effectiveness is probably similar to that for the referred pain.<sup>26, 29</sup>

415 The initial increase in EPN with remote dry needling followed by a reduction in

416 EPN after local twitch responses are elicited indicating inactivation of the remote

417 MTrS. Strong stimulation from continuous dry needling of an MTrS can activate the

418 sensitized nociceptors and generate strong impulses propagating to the spinal cord to

419 activate the corresponding motoneurons (including those in the same segment

420 corresponding to the needling muscle and other segments corresponding to the remote

421 muscles) to fire reflexively, thereby causing increased EPN amplitude in MTrSs not

422 only at the needling muscle but also at other remote muscles. These strongly activated

423 motoneurons are also controlled by recurrent inhibitions. As the firing rate of

424 motoneuron increases, the amount of recurrent inhibitions will also increase,

425 subsequently limiting and suppressing the firing rate of the efferents. In this way,

426 these impulses elicited by dry needling eventually breaks the vicious cycle of the

427 neural circuits (i.e. MTrP circuits<sup>29, 33</sup>) responsible for MTrSs through spatial and

428 temporal interactions in the spinal cord. Thus the EPN amplitude is suppressed after

429 dry needling. Possibly, there are certain neural connections among the inhibitory

430 interneuron and descending pain control system in the spinal cord that can modulate

431 the irritability of MTrPs when a remote painful dry needling stimulation is applied

432 (fig 7). The initial increase in EPN is consistent with suppression of the DNIC system,  
433 and the subsequent reduction in EPN is consistent with activation or enhancement of  
434 the DNIC system. Therefore, the physiological basis for the remote effects of dry  
435 needling may be related to an inactivation of MTrS<sup>29, 33</sup> and activation of DNIC<sup>34, 35</sup>  
436 induced by noxious stimulation applied at the painful region (such as trigger point  
437 needling) or at a remote site (such as in remote dry needling). This is probably the  
438 mechanism of remote pain control by dry needling which is similar to  
439 hyperstimulation analgesia in acupuncture.<sup>32,36</sup>

440

#### 441 **Limitations of the Study**

442 The difficulty in confirming the correlation between the alterations of EPN  
443 amplitude and pain intensity in rabbit may be criticized. However, a conclusion based  
444 on the human study<sup>4</sup> may be reasonably applied on rabbits, because there are plenty of  
445 similarities between the human MTrP and rabbit MTrS.<sup>18, 26, 28, 29</sup> Lack of follow-up  
446 assessments for the long-term remote effect is another deficiency of this study.  
447 However, we rarely see the long-term effects of dry needling if the underlying  
448 pathology of MTrP activation is not treated appropriately.<sup>6, 26, 27, 28, 33</sup> Another problem  
449 is that out sham needling (similar to superficial dry needling) may not be appropriate  
450 as a control. Fortunately, we see no significant changes after treatment with sham

451 needling. However, this could be related to the small sample size. In our clinical  
452 practice, we have observed much less effectiveness of superficial needling than that of  
453 deep dry needling with our multiple quick insertion technique. In addition,  
454 considering the individual differences in the motoneuron excitability and the  
455 supraspinal control of spinal inhibitory interneurons, just based on  
456 electrophysiological study, we are unable to distinguish the relative contribution from  
457 each inhibitory mechanism for motoneuronal excitability to the changes in MTrP  
458 irritability due to remote dry needling. All the above factors should be taken into  
459 consideration for data interpretation.



## **CONCLUSION**

We have demonstrated that an intact afferent from the stimulating site to the spinal cord and a normal function of spinal cord corresponding the innervation of the remotely affected muscles are essential for this remote effectiveness. This study may help in the understanding of the mechanism for beneficial effects of dry needling at remote MTrPs for myofascial pain control.

## **Acknowledgements**

The authors thank the Ms Pin-Wen Tu and Pei-Ling Chang for their skillful technical assistances with the EMG recordings and data analyses.

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## Suppliers

- a. Physitemp Instruments, Inc. 154, Huron Avenue, Clifton, New Jersey, USA.
- b. Yu-Kuang Industrial Co., Ltd., 5F-6, NO. 20, Lane 609, Sec.5, Chung-shing Rd.  
San-Chung City, Taipei, Taiwan.
- c. Neuro-EMG-Micro; Neurosoft, 5, Voronin Str, Ivanovo, Russia.
- d. Statistical Package for the Social Sciences version 10.0 for Windows; SPSS Inc.  
Headquarters, 233 S. Wacker Dr, 11th Fl, Chicago, IL 60606.

## Figure Legends

**Fig 1.** Study flow diagram. BF: biceps femoris; Contra: contralateral; EPN: endplate noise; GAS: gastrocnemius; Ipsi: ipsilateral; MTrS: myofascial trigger spot.

**Fig 2.** Sites of EPN recording or dry needling for all animals, and area receiving surgical transection for animals in Groups II, III, and IV.

**Fig 3.** A series of changes in the EPN amplitude measured at MTrS of biceps femoris before, during, and after dry needling manipulation at gastrocnemius (GAS) in the Group I. (A) Time course of EPN amplitude. (B) Sample recordings of EPN responses in two rabbits of Group I.

†:  $P < .05$ , showed significant differences among the four subgroups. \*:  $P < .05$

showed the significant differences compared to the values at pre-needling level in subgroups with dry needling at ipsilateral and contralateral GAS.

**Fig 4.** A series of changes in the EPN amplitude measured at MTrS of biceps femoris before and after tibial nerve transection surgery, and then, before, during and after dry needling manipulation at gastrocnemius (GAS) in Group II. (A) Time course of EPN



amplitude. (B) Sample recordings of EPN responses in two rabbits from Group II.

†:  $P < .05$ , showed significant differences among the four subgroups. \*:  $P < .05$ ,

showed the significant differences compared with the values at pre-needling level in

subgroups with dry needling at ipsilateral and contralateral GAS.

**Fig 5.** A series of changes in the EPN amplitude measured at MTrS of biceps femoris

before and 30–120 min after lumbar transection surgery, as well as before, during, and

after dry needling manipulation at gastrocnemius (GAS) in Group III. (A) Time

course of EPN amplitude. (B) Sample recordings of EPN responses in two rabbits

from Group III.

#:  $P < .05$ , showed the significant differences when compared the values at

pre-transected levels.

**Fig 6.** A series of changes in the EPN amplitude measured at MTrS of biceps femoris

before and 30–120 min after thoracic transection surgery, as well as before, during,

and after dry needling manipulation at gastrocnemius (GAS) in Group IV. (A) Time

course of EPN amplitude. (B) Sample recordings of EPN responses in two rabbits

from Group IV.

†:  $P < .05$ , showed significant differences among the four subgroups. \*:  $P < .05$ ,

showed the significant differences compared with the values at pre-needling level in subgroups with dry needling at ipsilateral and contralateral GAS; #:  $P < .05$ , showed significant differences when compared with the values at pre- transected levels.

**Fig 7.** Schematic drawing of the proposed neural mechanisms for remote effect on proximal MTrS in response to dry needling at distal MTrS.

1. Strong irritation to nociceptors in the MTrS by dry needling at gastrocnemius. 2.

Afferent input from gastrocnemius to dorsal horn (L6-S2 sensory neuron) probably in

the MTrS circuit. 3. Ascending projection to upper (L5-L6) sensory neurons probably

in another MTrS circuit (in dorsal horn). 4. Impulse via interneuron to L5-L6

motoneuron (anterior horn) corresponding to biceps femoris. 5. Increase efferent

output to neuromuscular junction in the biceps femoris. 6. Increase EPN amplitude. 7.

Strongly activated motoneuron also activates inhibitory interneuron to increase

recurrent inhibition on firing rate. 8. Suppress efferent output from motoneuron. 9.

Depress the EPN amplitude. 10. The excitability of inhibitory interneuron can also be

influenced by descending inputs, thereby altering the overall excitability of the

motoneuron pool and efferent as well as the irritability of MTrS at biceps femoris.

1 *ORIGINAL ARTICLE*

2 **Spinal Cord Mechanism Involving the Remote Effects of Dry Needling on the**

3 **Irritability of Myofascial Trigger Spots in Rabbit Skeletal Muscle**

4

5

6 **Running title: NEURAL MECHANISM FOR REMOTE EFFECT OF DRY**

7 **NEEDLING**

8

9 **ABSTRACT**

10

11 **Objective:** To elucidate the neural mechanisms underlying the remote effects  
12 produced by dry needling rabbit skeletal muscle myofascial trigger spots (MTrSs) via  
13 analyses of their endplate noise (EPN) recordings.

14 **Design:** Experimental animal controlled trial.

15 **Setting:** An animal laboratory of a university.

16 **Animals:** Ninety-six male New Zealand rabbits (body weight: 2.5–3.0 kg, age: 16–20  
17 weeks).

18 **Intervention:** Animals received no intervention for neural interruption in Group I,  
19 transection of the tibial nerve in Group II, transection of L5 and L6 spinal cord in  
20 Group III, and transection of the T1 and T2 spinal cord in Group IV. Each group was  
21 further divided into 4 subgroups: animals received ipsilateral dry needling (IDN),  
22 contralateral dry needling (CDN), ipsilateral sham needling (ISN), or contralateral  
23 sham needling (CSN), of gastrocnemius (GAS) MTrSs.

24 **Main Outcome Measures:** EPN amplitudes of biceps femoris (BF) MTrSs.

25 **Results:** BF MTrS mean EPN amplitudes significantly increased ( $P < .05$ ) initially  
26 after GAS verum needling, but reduced to a level significantly lower ( $P < .05$ ) than the  
27 pre-needling level in Groups I and IV with IDN or CDN, and in Group II with CDN

28 (but not IDN). No significant EPN amplitude changes were observed in BF MTrS in

29 Group III or in the control animals receiving superficial needling (sham).

30 **Conclusion:** This remote effect of dry needling depends on an intact afferent pathway

31 from the stimulating site to the spinal cord and a normal spinal cord function at the

32 levels corresponding to the innervation of the proximally affected muscle.

33

34 **Key Words:** Neural pathway; Dry needling; Myofascial trigger spot;

35 Electromyography; Remote effect.

36

37 **DEFINITION OF ACRONYMS**

38 BF = biceps femoris muscle;

39 CDN = contralateral dry needling;

40 CSN = contralateral sham needling;

41 EMG = electromyographic;

42 EPN = endplate noise;

43 GAS = gastrocnemius muscle;

44 IDN = ipsilateral dry needling;

45 ISN = ipsilateral sham needling;

46 LTR = local twitch response;

47 MTrP = myofascial trigger point (human);

48 MTrS = myofascial trigger spot (rabbit)

49

50

## INTRODUCTION

51 Myofascial pain is one of the most common sources of musculoskeletal pain and  
52 has as its hallmark the presence of taut bands and small hyperirritable regions referred  
53 to as myofascial trigger points (MTrPs).<sup>1,2</sup> Clinically, a given MTrP has a  
54 characteristic referred pain pattern, and may be associated with a local twitch  
55 response (LTR) in response to snapping palpation.<sup>2</sup> In the MTrP region,  
56 electromyographic (EMG) activity of endplate noise (EPN) can be recorded, and both  
57 prevalence and amplitude of EPN can be used as indicators to assess the irritability of  
58 MTrP.<sup>2-5</sup>

59 Dry needling has a well-established role in the treatment of myofascial pain.<sup>6-9</sup>  
60 Clinical studies have demonstrated that dry needling MTrPs at the pain site suppresses  
61 their activity resulting in pain reduction, but also that those MTrPs can also be  
62 suppressed through dry needling distant MTrPs that are anatomically proximal or  
63 distal to the site of clinical pain.<sup>8-12</sup> Acupuncture studies have demonstrated that the  
64 Traditional Chinese Medicine principle that pain conditions can be improved by  
65 stimulating acupuncture points distant from the site of pain.<sup>4,13-14</sup> The effects of  
66 acupuncture may also spread to the contralateral side.<sup>15</sup> Studies of acupuncture needle  
67 stimulation in anesthetized animals have identified a wide variety of reflex responses  
68 in remote modification of various organ functions.<sup>16</sup> However, its underlying neuronal

69 control mechanism remains unclear.

70 The purpose of this study is to confirm the remote effects of dry needling on

71 trigger points and to elucidate the neural mechanisms underlying the remote effects

72 produced by dry needling rabbit myofascial trigger spots (MTrSs, equivalent to

73 human MTrPs) in the gastrocnemius muscle (GAS)<sup>17, 18</sup> via analyses of EPN

74 recordings from the biceps femoris (BF).

75



## MATERIALS AND METHODS

### Animals

The experiments were performed on adult male New Zealand rabbits (ages from 16 to 20 weeks, body weight of 2.5–3.0 kg). Each animal was housed and cared for following the ethical guidelines of the International Association for Study of Pain in animals were followed.<sup>19</sup> Effort was made to minimize discomfort and to reduce the number of animals used. All animal experiments were conducted with the procedure approved by the Animal Care and Use Committee of a university in accordance with the Guidelines for Animal Experimentation.

Ninety-six rabbits were divided randomly into four groups (fig 1) based on the procedure performed. Group I (n=24) animals received no surgical intervention (intact neural pathway), Group II animals (n=24) received transection of tibial nerve in the electrophysiologically investigated side (peripheral sensory pathway), Group III animals (n = 24) received transection of L5-L6 spinal cord (BF innervation level), and Group IV animals (n=24) received transection of T1-T2 spinal cord (supra-segment of BF innervation). For the EPN amplitude variable, a sample size of 24 subjects in each group was sufficient to give statistical power of 97.06% with a significance level of  $P < .05$ . Animals in each group were randomly divided further into four subgroups based on the condition of treatment on GAS: experimental animals with ipsilateral dry

95 needling (IDN, n=8), or contralateral dry needling (CDN, n=8), control animals with  
96 ipsilateral sham needling (ISN, n=4), or contralateral sham needling (CSN, n=4), on  
97 the MTrS of the gastrocnemius. Fewer animals were studied in the control group since  
98 no significant changes were observed in all animals treated with sham needling.  
99 Regarding the assignment of groups or subgroups, animals were selected from the  
100 first available litter, and subsequently from the next litter, and so forth according to  
101 the sequence in a random table.

## 102 **Animal Preparation**

103 Before anesthesia, the tenderest spots (i.e., MTrS) of BF and GAS were  
104 identified by finger pinching. The animal responded to pinch stimulation with  
105 withdrawal of the lower limb, turning its head, screaming, etc, only when the most  
106 painful spot was pinched, and this most tender spot was confirmed as the MTrS.  
107 <sup>2,17,18,20-23</sup> These painful regions were marked on the skin with an indelible marker and  
108 were designated for electrophysiological assessment or dry needling. The animals  
109 were anesthetized with 2% isoflurane in oxygen flow for induction followed by a  
110 0.5% maintenance dose.<sup>24</sup> Body temperature was monitored by a thermistor probe of a  
111 thermometer<sup>a</sup> in the rectum and maintained at approximately 37.5 °C using a body  
112 temperature control system consisting of thermostatically regulated DC current  
113 heating pad and an infrared lamp. The hind limbs of anesthetized rabbits were shaved

114 and cleaned with povidone-iodine solution. The skin of the lateral thigh in one  
115 randomly selected side was incised to expose the BF, which served as an EPN  
116 recording site. The marked spot region in the BF muscle was grasped between two  
117 fingers from behind the muscle and the muscle palpated by gently rubbing (rolling) it  
118 between the fingers to discover a taut band. A taut band felt like a clearly delineated  
119 "rope" of muscle fibers and was roughly 2–3 mm or more in diameter. The fibers of  
120 the taut band were unmistakably firmer in consistency than the surrounding muscle.

### 121 **Needling of Gastrocnemius Muscle**

122 All needling procedures were performed by the same investigator who was  
123 blinded to the group assignment regarding to surgical intervention on neural pathway.  
124 Dry needling stimulation was performed with a disposable 30G acupuncture needle  
125 (300  $\mu$ m in diameter, 1.5 inches in length)<sup>b</sup> at ipsilateral or contralateral GAS (fig 2).  
126 The technique of dry needling was similar to that suggested by Hong<sup>18, 20, 25-27</sup> with  
127 multiple needle insertions to elicit rabbit-LTRs as much as possible. For needling in  
128 MTrS of GAS, the needle was first inserted through the skin perpendicularly at the  
129 center of the marked spot and advanced slowly and gently into the muscle until the  
130 needle tip touched the bone surface to estimate the thickness of the muscle. The  
131 needle was withdrawn back to the subcutaneous layer, and rapidly moved in and out  
132 for insertion of multiple sites in different directions (in a cone shape with the center at

133 the initial needle insertion of a perpendicular direction, and the angle of the cone  
134 margin was about 20°). For each needle insertion, the needle was advanced into the  
135 depth near the bone surface. Simultaneous needle rotation was performed to facilitate  
136 fast “in-and-out” needle movement as suggested by Chou et al.<sup>4</sup> in order to elicit as  
137 many LTRs as possible. For sham needling, the needle was inserted into the  
138 subcutaneous layer of the marked MTrP region at a depth approximately 1-2 mm from  
139 the skin surface. After insertion, the needle stayed there without further movement.

140

## 141 **Transection Operations**

### 142 **1. Transection of tibial nerve**

143 During anesthesia for the animals in Group II, the incision was made over the  
144 posterior aspect of one thigh ipsilaterally to the EPN recording side. Under the  
145 operating microscope, the sciatic nerve was exposed, and the tibial nerve isolated and  
146 transected at the site about 1 cm from its insertion into the GAS.

### 147 **2. Transection of spinal cord**

148 After completing laminectomy and making a slit in the dorsal portion of the dura  
149 mater, the cord was transected by a knife and then aspirated by suction at about 2 mm  
150 caudal and rostral to the level of transection, at L5-L6 levels of the spinal cord for  
151 animals in Group III, or at T1-T2 levels of the spinal cord for Group IV animals.

152 Gelfoam was placed into the empty vertebral column to seal the empty vertebral  
153 cavity and reduce bleeding. In a previous study<sup>20</sup> and in our preliminary data, about  
154 2½ h after surgery, the rabbits would have almost completely recovered from spinal  
155 shock. The animal would then be ready for the needling study.

156

## 157 **Recording of Endplate Noise**

### 158 **1. Electromyography setting**

159 For EPN assessment, a two-channel digital EMG machine<sup>c</sup> and monopolar  
160 needle electrodes (37 mm disposable Teflon-coated model) were used. The gain was  
161 set at 20µV per division for recordings from both channels. Low-cut frequency filter  
162 was set at 100 Hz and the high-cut at 1,000 Hz. Sweep speed was 10 ms per division.  
163 The search needle for EPN recording was inserted into the MTrS region and  
164 connected to the first channel of the EMG machine. The control needle was inserted  
165 into the non-taut band region near the MTrS in the same muscle and connected to the  
166 second channel. A common reference needle electrode for each channel was placed on  
167 the incised skin and connected to both channels via a y-connector.

### 168 **2. Search for endplate noise**

169 This procedure was performed by an investigator who was blind to the group  
170 assignment. The search needle was inserted into the MTrS region in a direction

171 parallel to the muscle fibers at an angle of approximately  $60^\circ$  to the surface of the  
172 muscle. After initial insertion just short of the depth of the MTrS or to a comparable  
173 depth in the case of control sites, the needle was advanced very slowly with  
174 simultaneous slow rotation to prevent it from 'grabbing' and releasing the tissue  
175 suddenly to advance in a large jump. Each advance was of minimal distance ( $\sim 1$  mm).  
176 When the needle approached an active locus (EPN locus), the continuous distant  
177 electrical activity, i.e., EPN, can be heard. As soon as EPN with amplitude higher than  
178  $10 \mu\text{V}$  could be recorded, the examiner stopped advancing the needle, but minimally  
179 moving the needle gently to different direction, trying to obtain EPN with highest  
180 amplitude. If this was impossible, the needle was advanced to another site until an  
181 EPN with optimal amplitude (usually higher than  $30\mu\text{V}$ ) could be recorded. Then the  
182 needle was fixed in place (carefully and firmly taped on the skin) to ensure that this  
183 EPN can run continuously on the recording screen with constant amplitudes.  
184 Continuous EPN tracing was recorded throughout the entire course of the experiment,  
185 and provided the opportunity for continuous visual observation of EPN changes on  
186 the EMG screen. If the EPN could not be sustained, the searching needle would be  
187 moved to another site until a satisfactory EPN tracing could be obtained. The entire  
188 EPN tracing found in MTrS of BF were recorded for the analysis of amplitude  
189 changes.

190 **3. Measurement of the amplitude of endplate noise**

191 Five randomly selected samples of EPN recordings (10 ms each) were taken  
192 before, during, and 3 min after the completion of the needling treatment for all groups;  
193 before and 30 minutes after surgery for Group II animals; and every 30 minutes up to  
194 120 minutes after surgery for Group III and IV animals. The mean amplitude of EPN  
195 of 5 random samples was analyzed and calculated through the embedded software in  
196 the Neuro-EMG-Micro equipment, and was recorded as the value for a certain  
197 measurement point for each animal.

198

199 **Data Analysis**

200 Data of EPN amplitudes in different measurement points for different groups or  
201 subgroups were expressed as the mean  $\pm$  standard error of the mean (SEM) for further  
202 statistical analysis. The Shapiro-Wilk's normality test was conducted to determine  
203 whether the data fit a normal distribution prior to subsequent analyses, and showed all  
204 measures of EPN amplitude were normally distributed. Tests of 'homogeneity' or  
205 'baseline balance' on covariates including body weight, age, and anesthesia condition  
206 were measured and equivalent before the needling treatment in all animals. The  
207 differences in EPN amplitude across measurement points in each group were carried  
208 out using repeated measures ANOVA, and later further analyzed by a Bonferroni

209 post-hoc method. The differences in EPM amplitudes within each of the subgroups  
210 (IDN, CDN, ISN and CSN) and across measurement points (before, during, and after  
211 needling) were analyzed using two-way ANOVA (side  $\times$  time) followed by a  
212 Bonferroni post-hoc analysis for each group. The differences in EPN amplitude within  
213 measurement point (before, during, and after needling) across subgroups (IDN, CDN,  
214 ISN and CSN) were tested by paired t-test. A p value of  $<0.05$  was considered to be  
215 statistically significant. All data was analyzed using SPSS version 10.0 for Windows.<sup>d</sup>  
216



217

## RESULTS

### 218 **Effects of Dry Needling of Distal MTrS in Intact Rabbits (Group I)**

219 The serial alterations of the mean EPN amplitude before, during, and after dry  
220 needling at ipsilateral and contralateral GAS for Group I are demonstrated in figure 3.  
221 Before needling treatment, there was no significant difference among the four  
222 subgroups (two-way ANOVA,  $F=0.10$ ,  $P> .05$ ).

223 The mean amplitudes of EPN before, during, and after needling were  
224  $18.20\pm 0.70\mu\text{V}$ ,  $27.71\pm 0.47\mu\text{V}$ , and  $13.15\pm 0.59\mu\text{V}$ , respectively in IDN subgroup, and  
225  $17.96\pm 0.69\mu\text{V}$ ,  $24.66\pm 1.47\mu\text{V}$ , and  $14.01\pm 0.86\mu\text{V}$ , respectively in CDN subgroup. In  
226 either IDN or CDN subgroup, the amplitudes at different times were significantly  
227 different (repeated measures ANOVA:  $F=45.99$  and  $P< .05$  for IDN,  $F=113.98$  and  
228  $P< .05$  for CDN). Compared with the data in the pre-needling level, the EPN  
229 amplitudes were significantly increased during the dry needling (Bonferroni post-hoc  
230 test,  $P< .05$ ), and then significantly decreased to a much lower level after completion  
231 of the needling treatment (Bonferroni post-hoc test,  $P< .05$ ) for either IDN or CDN  
232 subgroup as shown in figure 3. However, these serial alterations of EPN amplitudes  
233 were not found in the comparable subgroup ISN or CSN (repeated measures ANOVA,  
234  $P> .05$ ). There were significant differences in EPN amplitudes recorded either during  
235 or after needling between IDN and ISN, or between CDN and CSN subgroups

236 (Bonferroni post-hoc test,  $P < .05$ ), but not between IDN and IDN subgroups

237 (Bonferroni post-hoc test,  $P > .05$ ).

238

## 239 **Effects of Dry Needling of Distal MTrS in Rabbits With Tibial Nerve**

### 240 **Transection (Group II)**

241 The serial alterations of the mean EPN amplitude ( $\pm$ SEM) throughout the entire  
242 experiment in Group II are demonstrated in figure 4. The mean amplitude of EPN had  
243 no significant changes before, during, and after ipsilateral tibial nerve transection (i.e.,  
244 GAS denervation) (repeated measures ANOVA,  $F=0.06$ ,  $P > .05$ ). Before the needling  
245 treatment, there was no significant difference in EPN amplitude among the subgroups  
246 treated with dry or sham needling at ipsilateral or contralateral side (two-way ANOVA,  
247  $F=0.68$ ,  $P > .05$ ).

248 The mean amplitudes ( $\pm$ SEM) of EPN recorded from BF before, during, and after  
249 needling were  $16.72 \pm 0.34 \mu\text{V}$ ,  $16.64 \pm 0.37 \mu\text{V}$ , and  $15.46 \pm 0.50 \mu\text{V}$ , respectively in IDN  
250 subgroup, and were  $16.90 \pm 0.38 \mu\text{V}$ ,  $21.63 \pm 0.91 \mu\text{V}$ , and  $12.40 \pm 0.36 \mu\text{V}$ , respectively in  
251 CDN subgroup. There were significant differences in EPN amplitudes among those  
252 recorded before, during, and after needling (repeated measures ANOVA,  $F= 80.77$ ,  
253  $P < .05$ ) in CDN subgroup (similar to the changes in Group I, Bonferroni post-hoc test,  
254  $P < .05$ ), but not in IDN subgroup (repeated measures ANOVA,  $F= 2.89$ ,  $P > .05$ ).

255 There were significant differences in mean EPN amplitudes between CDN and its  
256 comparable CSN subgroups, but not between IDN and ISN subgroups (Bonferroni  
257 post-hoc test,  $P > .05$ ), at the time during (Bonferroni post-hoc test,  $P < .05$ ) and after  
258 (Bonferroni post-hoc test,  $P < .05$ ) needling. Moreover, there were significant  
259 differences in the magnitude or time-dependent alterations of EPN amplitude between  
260 CDN and CSN subgroups (Bonferroni post-hoc test,  $P < .05$ ).

261

## 262 **Effects of Dry Needling of Distal MTrS in Rabbits With Lumbar**

### 263 **Cord Transection (Group III)**

264 The serial alterations of the mean EPN amplitude ( $\pm$ SEM) throughout the entire  
265 experiment for Group III are shown in figure 5. There were significant differences in  
266 EPN amplitude among those recorded before, immediately, and 30, 60, 90, and 120  
267 min after L5-L6 transection (repeated measures ANOVA,  $F=29.81$ ,  $P < .05$ ). During  
268 the 2 h period after transection, the mean EPN amplitudes were significantly lower  
269 than the pre-transected levels (Bonferroni post-hoc test, all were  $P < .05$  at 30, 60, 90,  
270 and 120 h). There was no significant difference among the four subgroups treated with  
271 dry or sham needling at the ipsilateral or contralateral side regardless of the time  
272 before (two-way ANOVA,  $F=0.23$ ,  $P > .05$ ), during ( $F=1.45$ ,  $P > .05$ ), or after ( $F=1.72$ ,  
273  $P > .05$ ) needling treatments.

274 The mean amplitudes ( $\pm$ SEM) of EPN before, during, and after needling were  
275  $11.56\pm 0.36\mu\text{V}$ ,  $11.47\pm 0.43\mu\text{V}$ , and  $11.28\pm 0.47\mu\text{V}$ , respectively in IDN subgroup, and  
276 were  $11.67\pm 0.45\mu\text{V}$ ,  $12.32\pm 0.46\mu\text{V}$ , and  $12.33\pm 0.46\mu\text{V}$ , respectively in CDN subgroup.  
277 There was no significant difference in EPN amplitude among those recorded before,  
278 during, and after IDN (repeated measures ANOVA,  $F=0.63$ ,  $P> .05$ ), CDN (repeated  
279 measures ANOVA,  $F=1.17$ ,  $P> .05$ ), ISN treatment (repeated measures ANOVA,  
280  $F=0.23$ ,  $P> .05$ ), or CSN (repeated measures ANOVA,  $F=0.52$ ,  $P> .05$ ). There were  
281 no significant differences in each time-dependent alterations of EPN amplitude  
282 between IDN and ISN subgroups, between IDN and CDN subgroups, or between CDN  
283 and CSN subgroups (Bonferroni post-hoc test, all  $P> .05$ ).

284

## 285 **Effects of Dry Needling of Distal MTrS in Rabbits With Thoracic**

### 286 **Cord Transection (Group IV)**

287 The serial alterations of the mean EPN amplitude ( $\pm$ SEM) throughout the entire  
288 experiment for Group IV are shown in figure 6. The EPN amplitude recorded at 30  
289 and 60 min after T1-T2 transection was significantly reduced when compared to the  
290 pre-transection level (Bonferroni post-hoc test,  $P< .05$ ), but recovered to  
291 pre-transection level after 90 min (Bonferroni post-hoc tests, all  $P> .05$ ), and  
292 normalized to be similar for all subgroups by 120 min after transaction (two-way

293 ANOVA,  $F=0.09$ ,  $P> .05$ ).

294

295       The mean amplitudes ( $\pm$ SEM) of EPN recorded from BF before, during, and after  
296 needling were  $18.17\pm 0.36\mu\text{V}$ ,  $26.88\pm 0.43\mu\text{V}$ , and  $15.74\pm 0.26\mu\text{V}$ , respectively in IDN  
297 subgroup, and were  $18.28\pm 0.45\mu\text{V}$ ,  $27.72\pm 0.47\mu\text{V}$ , and  $16.20\pm 0.22\mu\text{V}$  respectively in  
298 CDN subgroup. There were significant differences in EPN amplitudes among those  
299 recorded before, during, and after IDN (repeated measures ANOVA,  $F= 264.29$ ,  
300  $P< .05$ ) or CDN (repeated measures ANOVA,  $F= 243.11$ ,  $P< .05$ ). The mean EPN  
301 amplitudes were significantly increased during IDN or CDN (Bonferroni post-hoc test,  
302  $P< .05$ ), but after cessation of needling, they reduced significantly to a level less than  
303 that before needling (Bonferroni post-hoc test,  $P< .05$ ), but there were no significant  
304 differences in these changes between IDN and CDN subgroups (Bonferroni post-hoc  
305 test,  $P> .05$ ). These serial alterations of EPN amplitudes were not found in either ISN  
306 (repeated measures ANOVA,  $F=1.02$ ,  $P> .05$ ) or CSN subgroup (repeated measures  
307 ANOVA,  $F=2.02$ ,  $P> .05$ ).

308

## DISCUSSION

309

310 To our knowledge, the present study is the first animal study to investigate the  
311 neural mechanism of the remote effects of dry needling. In this study, we found that  
312 an intact afferent nerve from the remote stimulation site and normal spinal cord  
313 segments corresponding to the innervation of the affected proximal muscle are  
314 essential for the remote effect from either ipsilateral or contralateral stimulation.

315 The dry needling used in this study is a technique of MTrP injection with  
316 multiple high-speed needle insertions into different sensitive loci in an MTrP region  
317 suggested by Hong.<sup>25-27</sup> High speed needling can provide high-pressure stimulation to  
318 the sensitive loci in the MTrP region to elicit LTRs. It is essential to elicit LTRs  
319 during needling of an MTrP in order to obtain immediate and complete pain relief.<sup>6,</sup>  
320 <sup>25-29</sup> Dry needling at the MTrS was effective in diminishing spontaneous electrical  
321 activity (i.e., EPN) of MTrS of rabbit skeletal muscle if LTRs were elicited.<sup>30</sup> After  
322 several LTRs had been elicited by the needling of an MTrS of rabbit skeletal muscle,  
323 no more LTRs could be elicited from the same region<sup>18</sup> and the irritability of the  
324 MTrS could be suppressed.<sup>30</sup> Needling-elicited LTRs are involuntary discharges of  
325 muscle fiber mediated through the nervous system and integrated at the spinal cord  
326 level.<sup>18,20</sup> Therefore, it is important to apply this needling technique to achieve the  
327 best needling effect or remote needling effect for the study on the neural mechanism.

328 Changes in the EPN amplitude in the MTrS were found during and after dry  
329 needling at the distal MTrSs in animals with intact neural circuits (Group I). It appears  
330 that either IDN or CDN to the distal MTrS could initially increase the irritability of  
331 the proximal MTrS, followed by a suppression effect after cessation of needling.  
332 Fernandez-Camero, et al.<sup>31</sup> also found an increase in spontaneous electrical activity at  
333 an MTrP region during a persistent noxious stimulation at another distant MTrP,  
334 followed by a suppression of electrophysiological irritability after cessation of  
335 needling. The two findings above strongly support clinical observations related to the  
336 interaction between one MTrP and another MTrP located in the region of the referred  
337 pain (referred zone) of that MTrP.<sup>2,6,9-12,26-29</sup>

338 In the study on Group II rabbits with ipsilateral denervation of GAS, the remote  
339 effect disappeared after IDN, but persisted after CDN. These results demonstrated the  
340 importance of an intact afferent pathway to the spinal cord in the remote modulation  
341 of EPN amplitudes. After destruction of spinal cord corresponding to the level of BF  
342 (Group III), the remote effect disappeared after either IDN or CDN treatment. This  
343 finding suggested the existence of intraspinal connections between the GAS afferents  
344 and BF spinal interneurons. The partial suppression of the EPN amplitude after spinal  
345 cord transection at L5-L6 is possibly related to the influence of the spinal shock. After  
346 interruption of upper motor neuron and supra-sensory connections (Group IV), the

347 remote effect persisted, either with IDN or CDN, but smaller than that in intact  
348 animals (Group I). It may imply the possible influences from supraspinal centers, such  
349 as descending pain inhibitory systems. These influences are anticipated to be minimal  
350 after transection at higher spinal levels. Loss of inputs to this system would weaken  
351 the inhibition on the pain level,<sup>32</sup> which could lead to the EPN amplitude being less  
352 suppressed. Therefore, the recruitment of the diffuse noxious inhibitory control  
353 (DNIC) system may be also elicited by dry needling treatment on regions remote to  
354 the stimulation site.

355       The neural pathway for the remote effect appears to be a spinal reflex, probably  
356 similar to that mediating the referred pain<sup>26, 29</sup> and local twitch response.<sup>18, 20, 26, 29</sup>  
357 Hong has hypothesized a corresponding “MTrP circuit” for each MTrP, which can  
358 modulate the pain, referred pain, and local twitch response elicited by stimulating the  
359 MTrP.<sup>6, 29, 33</sup> The neural connection in the spinal cord responsible for this remote  
360 effectiveness is probably similar to that for the referred pain.<sup>26, 29</sup>

361       The initial increase in EPN with remote dry needling followed by a reduction in  
362 EPN after local twitch responses are elicited indicating inactivation of the remote  
363 MTrS. Strong stimulation from continuous dry needling of an MTrS can activate the  
364 sensitized nociceptors and generate strong impulses propagating to the spinal cord to  
365 activate the corresponding motoneurons (including those in the same segment



366 corresponding to the needling muscle and other segments corresponding to the remote  
367 muscles) to fire reflexively, thereby causing increased EPN amplitude in MTrSs not  
368 only at the needling muscle but also at other remote muscles. These strongly activated  
369 motoneurons are also controlled by recurrent inhibitions. As the firing rate of  
370 motoneuron increases, the amount of recurrent inhibitions will also increase,  
371 subsequently limiting and suppressing the firing rate of the efferents. In this way,  
372 these impulses elicited by dry needling eventually breaks the vicious cycle of the  
373 neural circuits (i.e. MTrP circuits<sup>29,33</sup>) responsible for MTrSs through spatial and  
374 temporal interactions in the spinal cord. Thus the EPN amplitude is suppressed after  
375 dry needling. Possibly, there are certain neural connections among the inhibitory  
376 interneuron and descending pain control system in the spinal cord that can modulate  
377 the irritability of MTrPs when a remote painful dry needling stimulation is applied  
378 (fig 7). The initial increase in EPN is consistent with suppression of the DNIC system,  
379 and the subsequent reduction in EPN is consistent with activation or enhancement of  
380 the DNIC system. Therefore, the physiological basis for the remote effects of dry  
381 needling may be related to an inactivation of MTrS<sup>29,33</sup> and activation of DNIC<sup>34,35</sup>  
382 induced by noxious stimulation applied at the painful region (such as trigger point  
383 needling) or at a remote site (such as in remote dry needling). This is probably the  
384 mechanism of remote pain control by dry needling which is similar to

385 hyperstimulation analgesia in acupuncture.<sup>32,36</sup>

386

### 387 **Limitations of the Study**

388       The difficulty in confirming the correlation between the alterations of EPN  
389 amplitude and pain intensity in rabbit may be criticized. However, a conclusion based  
390 on the human study<sup>4</sup> may be reasonably applied on rabbits, because there are plenty of  
391 similarities between the human MTrP and rabbit MTrS.<sup>18, 26, 28, 29</sup> Lack of follow-up  
392 assessments for the long-term remote effect is another deficiency of this study.  
393 However, we rarely see the long-term effects of dry needling if the underlying  
394 pathology of MTrP activation is not treated appropriately.<sup>6, 26, 27, 28, 33</sup> Another problem  
395 is that out sham needling (similar to superficial dry needling) may not be appropriate  
396 as a control. Fortunately, we see no significant changes after treatment with sham  
397 needling. However, this could be related to the small sample size. In our clinical  
398 practice, we have observed much less effectiveness of superficial needling than that of  
399 deep dry needling with our multiple quick insertion technique. In addition,  
400 considering the individual differences in the motoneuron excitability and the  
401 supraspinal control of spinal inhibitory interneurons, just based on  
402 electrophysiological study, we are unable to distinguish the relative contribution from  
403 each inhibitory mechanism for motoneuronal excitability to the changes in MTrP

- 404 irritability due to remote dry needling. All the above factors should be taken into
- 405 consideration for data interpretation.

## **CONCLUSION**

We have demonstrated that an intact afferent from the stimulating site to the spinal cord and a normal function of spinal cord corresponding the innervation of the remotely affected muscles are essential for this remote effectiveness. This study may help in the understanding of the mechanism for beneficial effects of dry needling at remote MTrPs for myofascial pain control.

## **Acknowledgements**

The authors thank the Ms Pin-Wen Tu and Pei-Ling Chang for their skillful technical assistances with the EMG recordings and data analyses.

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## **Suppliers**

- a. Physitemp Instruments, Inc. 154, Huron Avenue, Clifton, New Jersey, USA.
- b. Yu-Kuang Industrial Co., Ltd., 5F-6, NO. 20, Lane 609, Sec.5, Chung-shing Rd.  
San-Chung City, Taipei, Taiwan.
- c. Neuro-EMG-Micro; Neurosoft, 5, Voronin Str, Ivanovo, Russia.
- d. Statistical Package for the Social Sciences version 10.0 for Windows; SPSS Inc.  
Headquarters, 233 S. Wacker Dr, 11th Fl, Chicago, IL 60606.

## Figure Legends

**Fig 1.** Study flow diagram. BF: biceps femoris; Contra: contralateral; EPN: endplate noise; GAS: gastrocnemius; Ipsi: ipsilateral; MTrS: myofascial trigger spot.

**Fig 2.** Sites of EPN recording, dry needling for all animals, and area receiving surgical transection for animals in Groups II, III, and IV.

**Fig 3.** A series of changes in the EPN amplitude measured at MTrS of biceps femoris before, during, and after dry needling manipulation at gastrocnemius (GAS) in the Group I. (A) Time course of EPN amplitude. (B) Sample recordings of EPN responses in two rabbits of Group I.

†:  $P < .05$ , showed significant differences among the four subgroups. \*:  $P < .05$

showed the significant differences compared to the values at pre-needling level in subgroups with dry needling at ipsilateral and contralateral GAS.

**Fig 4.** A series of changes in the EPN amplitude measured at MTrS of biceps femoris before and after tibial nerve transection surgery, and then, before, during and after dry needling manipulation at gastrocnemius (GAS) in Group II. (A) Time course of EPN

amplitude. (B) Sample recordings of EPN responses in two rabbits from Group II.

†:  $P < .05$ , showed significant differences among the four subgroups. \*:  $P < .05$ ,

showed the significant differences compared with the values at pre-needling level in

subgroups with dry needling at ipsilateral and contralateral GAS.

**Fig 5.** A series of changes in the EPN amplitude measured at MTrS of biceps femoris

before and 30–120 min after lumbar transection surgery, as well as before, during, and

after dry needling manipulation at gastrocnemius (GAS) in Group III. (A) Time

course of EPN amplitude. (B) Sample recordings of EPN responses in two rabbits

from Group III.

#:  $P < .05$ , showed the significant differences when compared the values at

pre-transected levels.

**Fig 6.** A series of changes in the EPN amplitude measured at MTrS of biceps femoris

before and 30–120 min after thoracic transection surgery, as well as before, during,

and after dry needling manipulation at gastrocnemius (GAS) in Group IV. (A) Time

course of EPN amplitude. (B) Sample recordings of EPN responses in two rabbits

from Group IV.

†:  $P < .05$ , showed significant differences among the four subgroups. \*:  $P < .05$ ,

showed the significant differences compared with the values at pre-needling level in subgroups with dry needling at ipsilateral and contralateral GAS; #:  $P < .05$ , showed significant differences when compared with the values at pre- transected levels.

**Fig 7.** Schematic drawing of the proposed neural mechanisms for remote effect on proximal MTrS in response to dry needling at distal MTrS.

1. Strong irritation to nociceptors in the MTrS by dry needling at gastrocnemius. 2.

Afferent input from gastrocnemius to dorsal horn (L6-S2 sensory neuron) probably in

the MTrS circuit. 3. Ascending projection to upper (L5-L6) sensory neurons probably

in another MTrS circuit (in dorsal horn). 4. Impulse via interneuron to L5-L6

motoneuron (anterior horn) corresponding to biceps femoris. 5. Increase efferent

output to neuromuscular junction in the biceps femoris. 6. Increase EPN amplitude. 7.

Strongly activated motoneuron also activates inhibitory interneuron to increase

recurrent inhibition on firing rate. 8. Suppress efferent output from motoneuron. 9.

Depress the EPN amplitude. 10. The excitability of inhibitory interneuron can also be

influenced by descending inputs, thereby altering the overall excitability of the

motoneuron pool and efferent as well as the irritability of MTrS at biceps femoris.

Figure 1

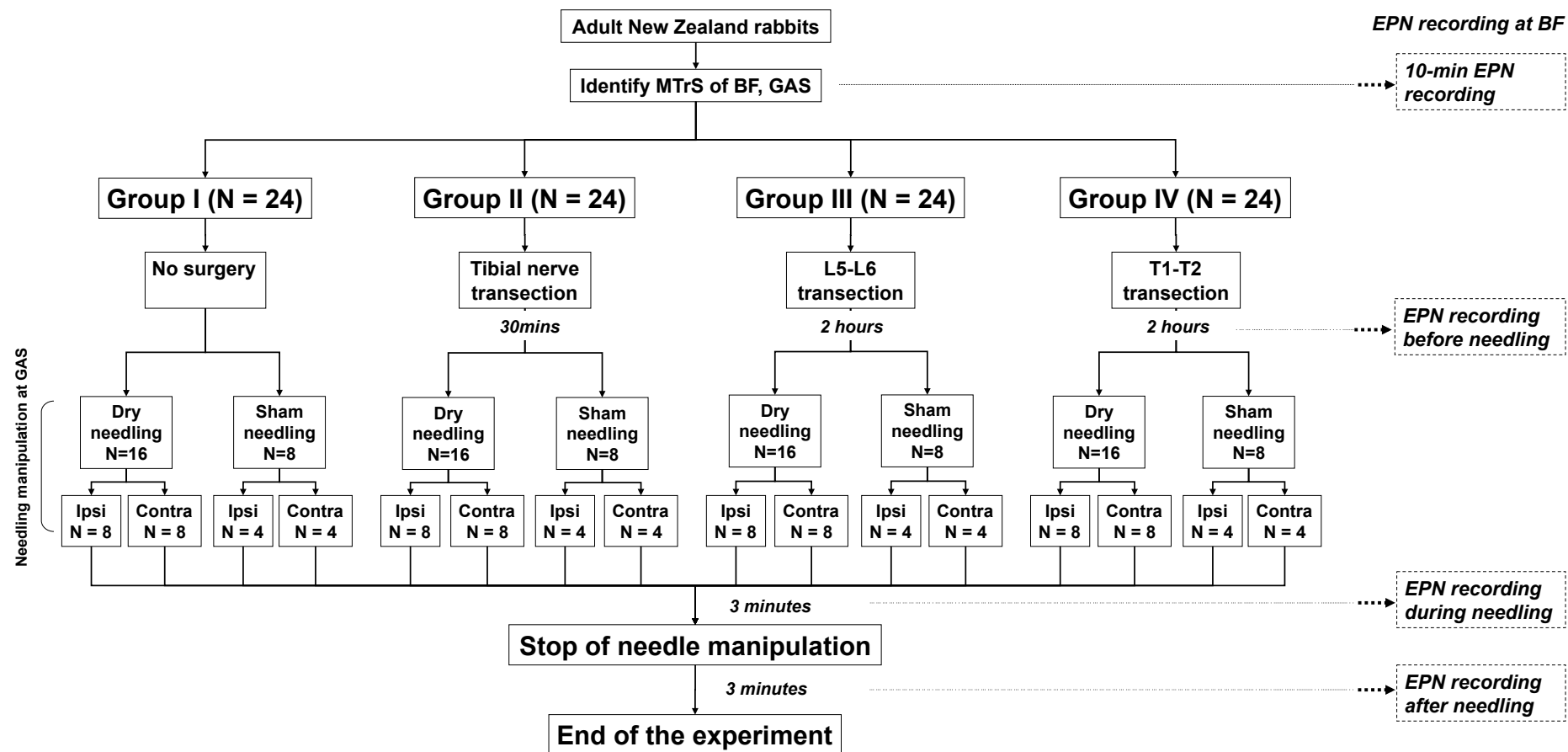
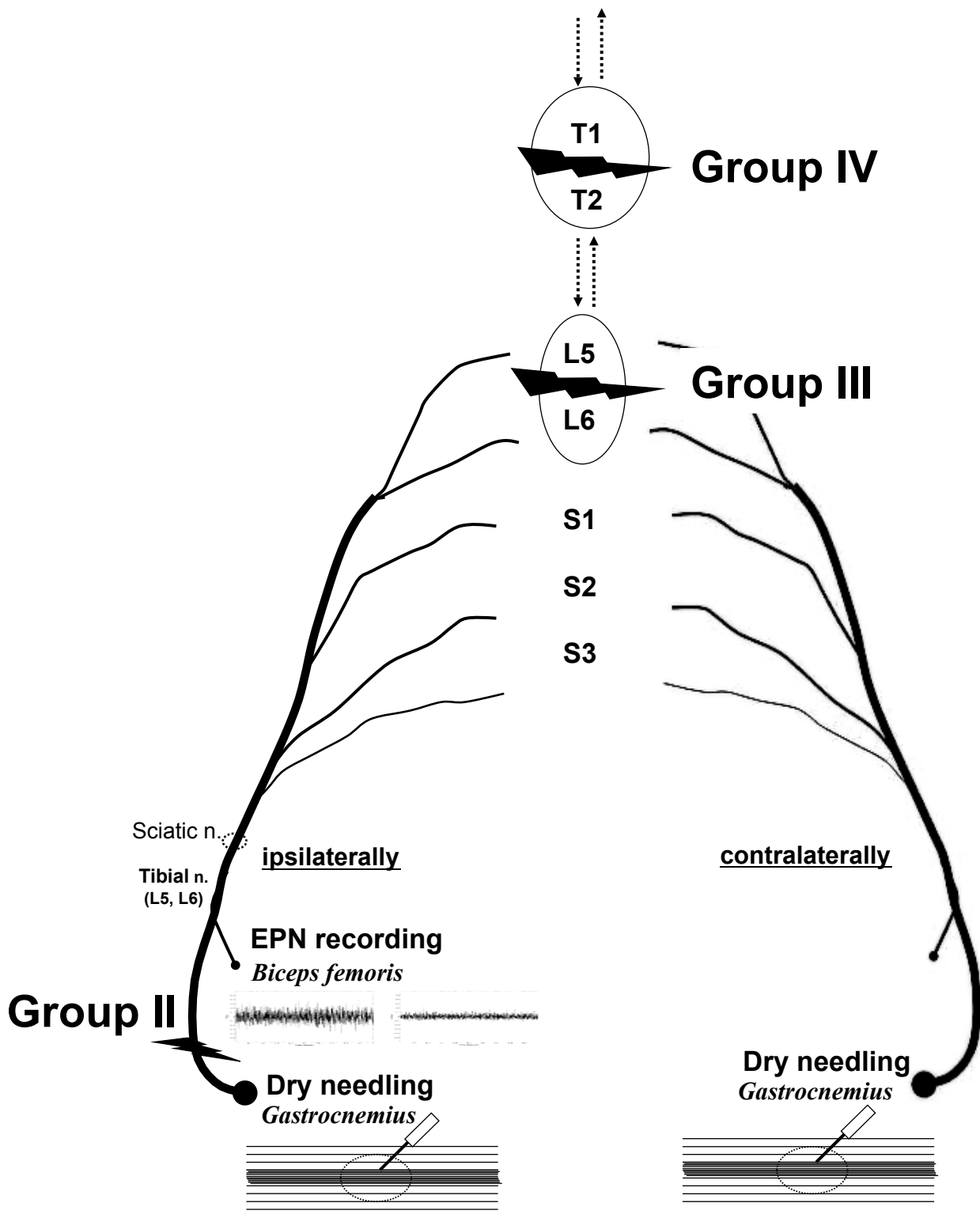
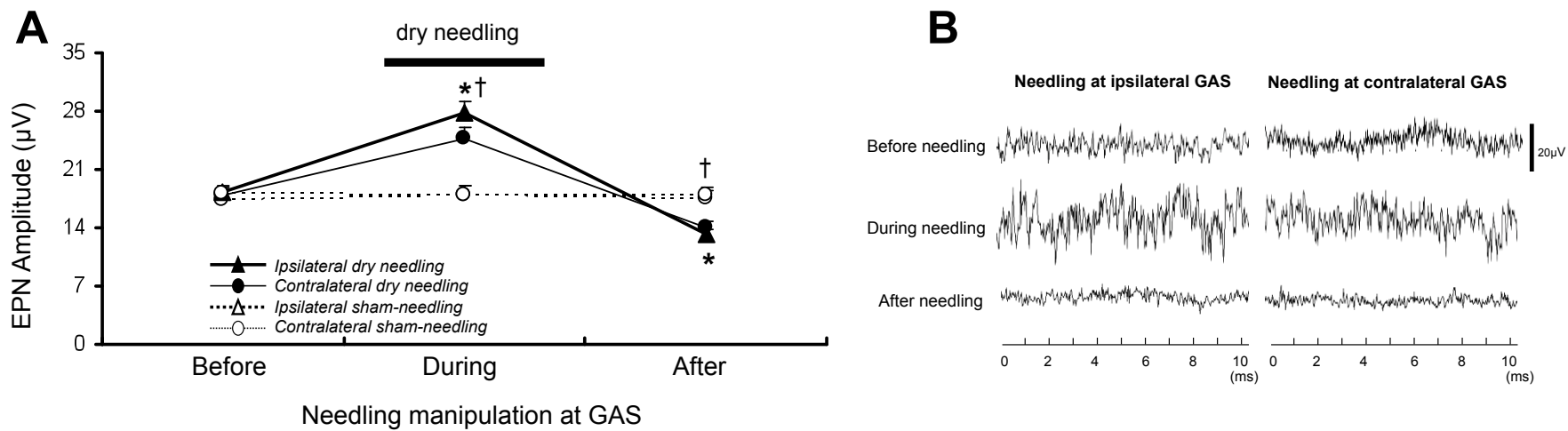


Fig 1. Study flow diagram. BF: biceps femoris; Contra: contralateral; EPN: endplate noise; GAS: gastrocnemius; Ipsi: ipsilateral; MTrS: myofascial trigger spot.



**Fig 2.** Sites of EPN recording, dry needling for all animals, and area receiving surgical transection for animals in Groups II, III, and IV.

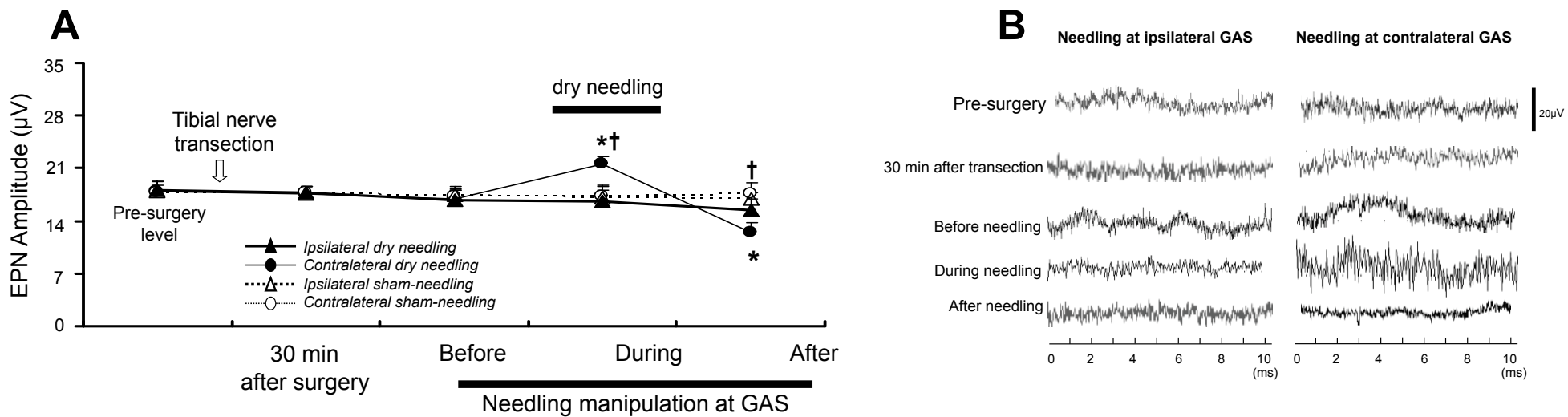
Figure 3



**Fig 3.** A series of changes in the EPN amplitude measured at MTrS of biceps femoris before, during, and after dry needling manipulation at gastrocnemius (GAS) in the Group I. (A) Time course of EPN amplitude. (B) Sample recordings of EPN responses in two rabbits of Group I. †:  $P < 0.05$ , showed significant differences among the four subgroups. \*:  $P < 0.05$  showed the significant differences compared to the values at pre-needling level in subgroups with dry needling at ipsilateral and contralateral GAS.



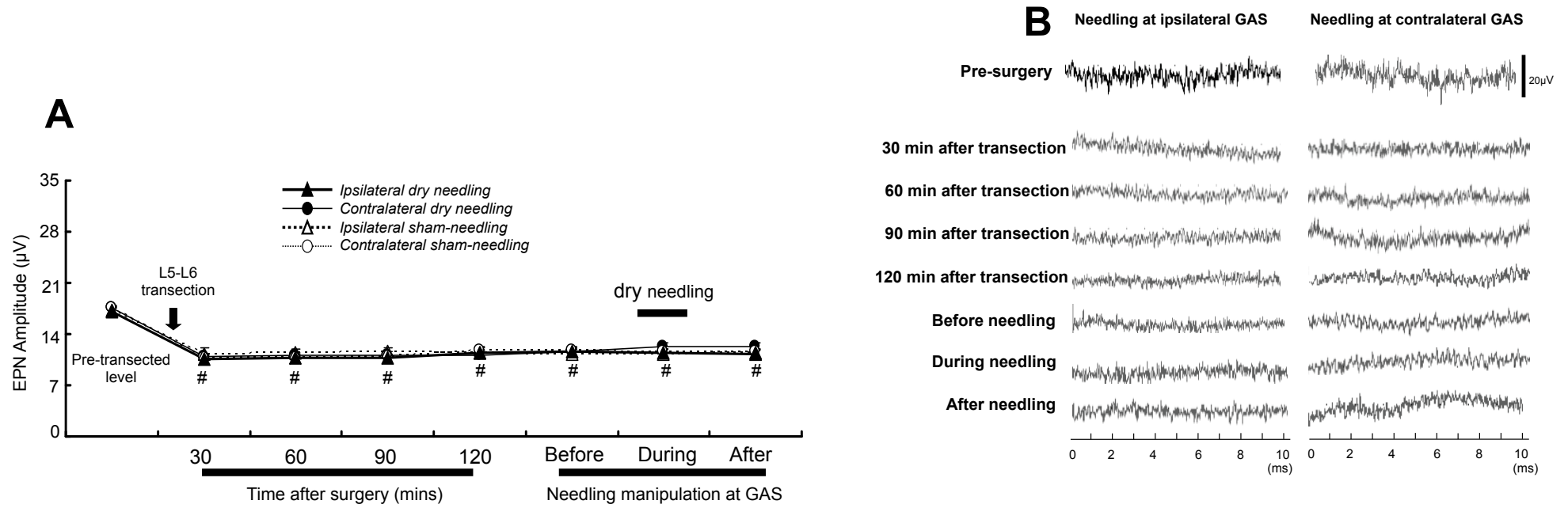
Figure 4



**Fig 4.** A series of changes in the EPN amplitude measured at MTrS of biceps femoris before and after tibial nerve transection surgery, and then, before, during and after dry needling manipulation at gastrocnemius (GAS) in Group II. (A) Time course of EPN amplitude. (B) Sample recordings of EPN responses in two rabbits from Group II.

†:  $P < 0.05$ , showed significant differences among the four subgroups. \*:  $P < 0.05$ , showed the significant differences compared with the values at pre-needling level in subgroups with dry needling at ipsilateral and contralateral GAS.

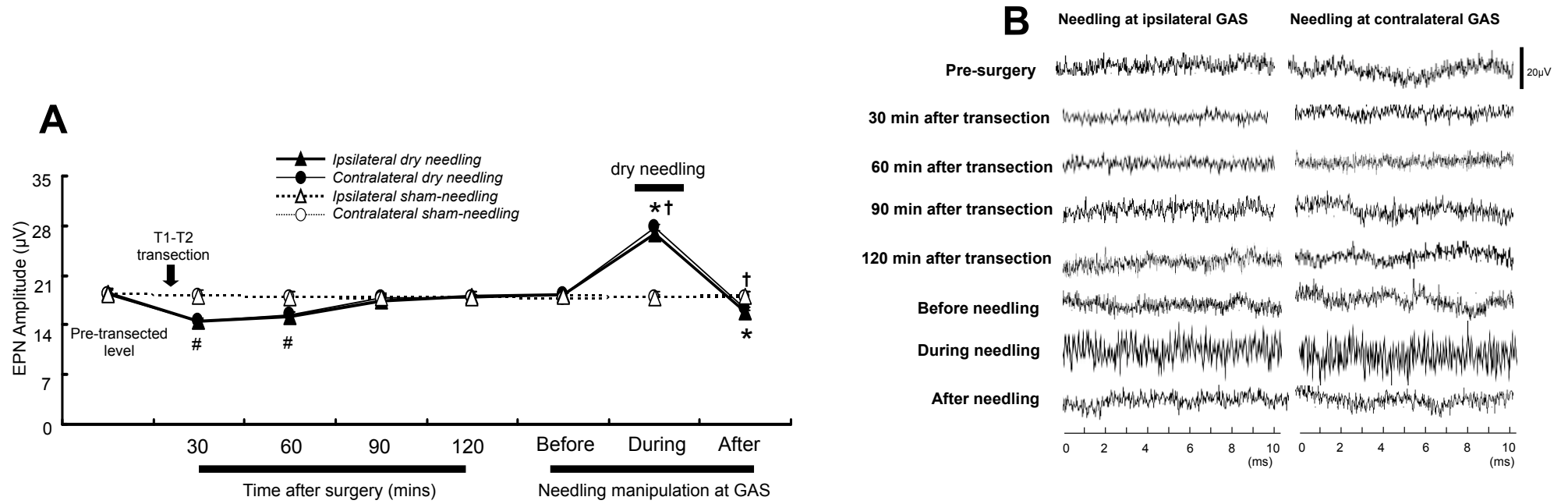
Figure 5



**Fig 5.** A series of changes in the EPN amplitude measured at MTrS of biceps femoris before and 30–120 min after lumbar transection surgery, as well as before, during, and after dry needling manipulation at gastrocnemius (GAS) in Group III. (A) Time course of EPN amplitude. (B) Sample recordings of EPN responses in two rabbits from Group III.

#:  $P < 0.05$ , showed the significant differences when compared the values at pre-transected levels.

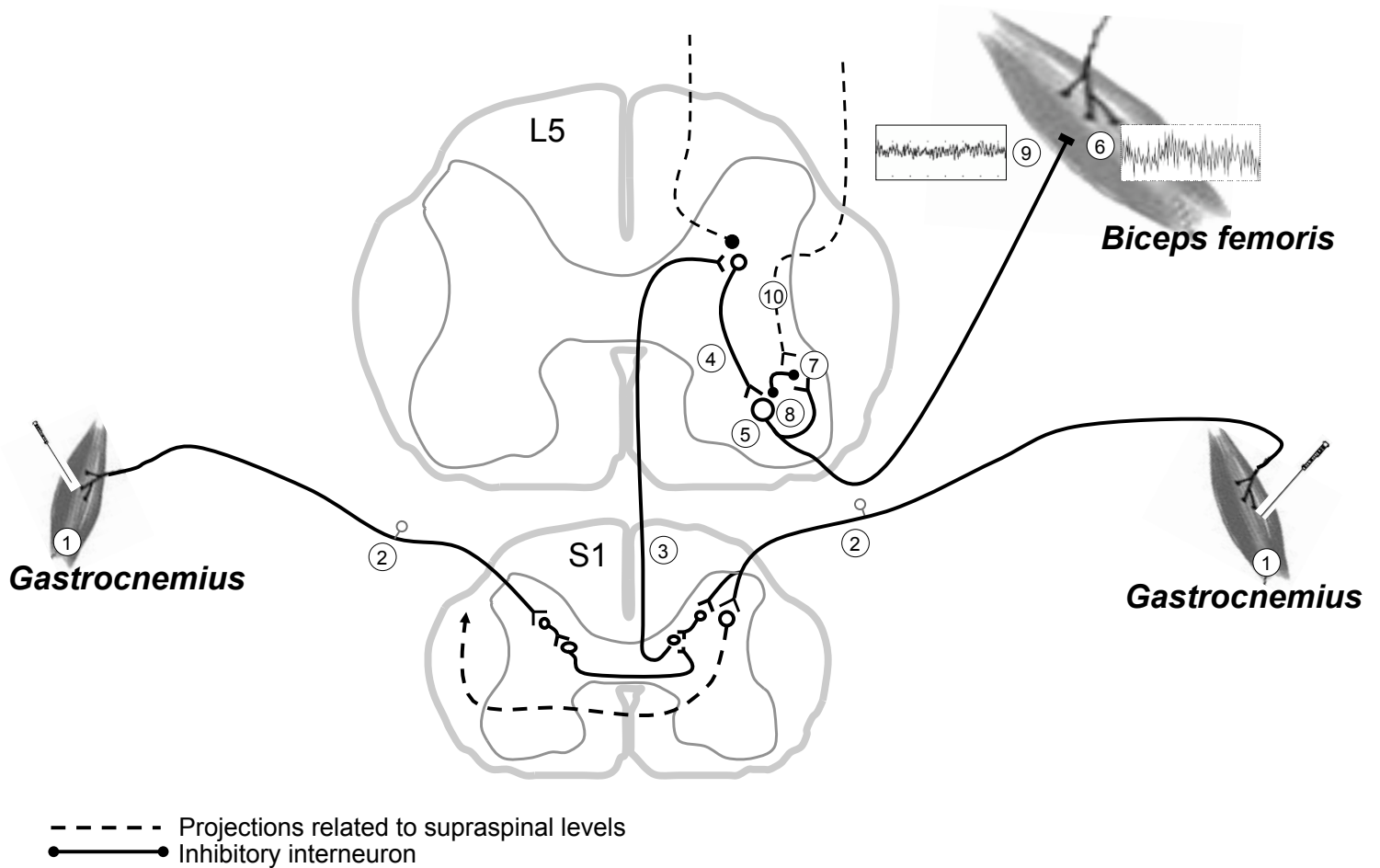
Figure 6



**Fig 6.** A series of changes in the EPN amplitude measured at MTrS of biceps femoris before and 30–120 min after thoracic transection surgery, as well as before, during, and after dry needling manipulation at gastrocnemius (GAS) in Group IV. (A) Time course of EPN amplitude. (B) Sample recordings of EPN responses in two rabbits from Group IV.

†:  $P < 0.05$ , showed significant differences among the four subgroups. \*:  $P < 0.05$ , showed the significant differences compared with the values at pre-needling level in subgroups with dry needling at ipsilateral and contralateral GAS; #:  $P < 0.05$ , showed significant differences when compared with the values at pre-transected levels.

Figure 7



**Fig 7.** Schematic drawing of the proposed neural mechanisms for remote effect on proximal MTrS in response to dry needling at distal MTrS.  
1. Strong irritation to nociceptors in the MTrS by dry needling at gastrocnemius. 2. Afferent input from gastrocnemius to dorsal horn (L6-S2 sensory neuron) probably in the MTrS circuit. 3. Ascending projection to upper (L5-L6) sensory neurons probably in another MTrS circuit (in dorsal horn). 4. Impulse via interneuron to L5-L6 motoneuron (anterior horn) corresponding to biceps femoris. 5. Increase efferent output to neuromuscular junction in the biceps femoris. 6. Increase EPN amplitude. 7. Strongly activated motoneuron also activates inhibitory interneuron to increase recurrent inhibition on firing rate. 8. Suppress efferent output from motoneuron. 9. Depress the EPN amplitude. 10. The excitability of inhibitory interneuron can also be influenced by descending inputs, thereby altering the overall excitability of the motoneuron pool and efferent as well as the irritability of MTrS at biceps femoris.

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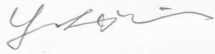
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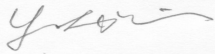
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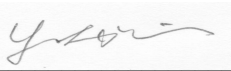
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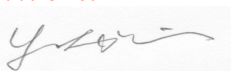
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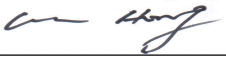
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