AUTHOR QUERY FORM

Journal: LFS ELSEVIER Article Number: 12953	Please e-mail or fax your responses and any corrections to: E-mail: <u>corrections.esil@elsevier.spitech.com</u> Fax: +1 619 699 6721
-------------------------------------------------------------	---------------------------------------------------------------------------------------------------------------------------------------------

Dear Author,

Any queries or remarks that have arisen during the processing of your manuscript are listed below and highlighted by flags in the proof. Please check your proof carefully and mark all corrections at the appropriate place in the proof (e.g., by using onscreen annotation in the PDF file) or compile them in a separate list.

For correction or revision of any artwork, please consult http://www.elsevier.com/artworkinstructions.

Any queries or remarks that have arisen during the processing of your manuscript are listed below and highlighted by flags in the proof. Click on the 'Q' link to go to the location in the proof.

Location	on Query / Remark: <u>click on the Q link to go</u>			
in article	cle Please insert your reply or correction at the corresponding line in the pr			
Q1	Figures 1 and 6 contain poor quality of text. Please check and provide replacement as deemed necessary.			

Thank you for your assistance.

Life Sciences xxx (2011) xxx-xxx



3

6

7

8

11

1

1

2

2

2

 $\frac{2}{2}$

2

2

50

49

51

Contents lists available at ScienceDirect

Life Sciences



journal homepage: www.elsevier.com/locate/lifescie

Study of the reversal effect of NF449 on neuromuscular blockade induced by d-tubocurarine

Tzu-Rong Su^a, Yu-Shiang Hung^e, Shiang Suo Huang^b, Hsing Hui Su^b, Ching-Chyuan Su^a, George Hsiao^c, Yi-Hung Chen^d, Min-Jon Lin^{e,f,*}

^a Tian-Sheng Memorial Hospital, Tong-Kang, Pintong, Taiwan

^b Department of Pharmacology and Institute of Medicine, College of Medicine, Chung Shan Medical University, Taichung, Taiwan

^c Department and Graduate Institute of Pharmacology, College of Medicine, Taipei Medical University, Taipei, Taiwan

^d Graduate Institute of Acupuncture Science, China Medical University, Taichung, Taiwan

^e Department of Biomedical Sciences, Chung Shan Medical University, Taichung, Taiwan

10 ^f Department of Medical Research, Chung Shan Medical University Hospital, Taichung, Taiwan

ARTICLE INFO

Article history:
Received 5 December 2010
Accepted 22 March 2011
Available online xxxx
Keywords:
NF449
Suramin analogues
Neuromuscular blockade reversal agents
Nicotinic acetylcholine receptor
D-tubocurarine
Purinoceptors

ABSTRACT

Aims: The aim of this study was to investigate the mechanism for the reversal effect of NF449 (a suramin 27 analogue) on the neuromuscular block induced by D-tubocurarine (D-TC). 28

Main methods: Nerve-stimulated muscle contractions and end-plate potentials were performed in mouse29phrenic nerve-diaphragm preparations. Acetylcholine (ACh)-induced muscle contractions were performed in30the chick biventer cervicis preparations. Presynaptic nerve terminal waveform recordings were performed in31mouse triangularis sterni preparations.32

Key findings: Amongst the suramin analogues in this study, only the NF449 and suramin were able to reverse 33 the blockade effect produced by D-TC on nerve-stimulated muscle contractions. Each of these suramin 34 analogues (NF007, NF023, NF279 and NF449) alone has no significant effect on the amplitude of nerve- 35 stimulated muscle contractions. NF449 and suramin also showed the antagonising effects on the inhibition of 36 end-plate potentials induced by D-TC. Furthermore, pre-treatment with NF449 can antagonise the inhibition 37 of D-TC in ACh-induced contractions of chick biventer cervicis muscle. NF449 produced a greater rightward 38 shift of the dose-response inhibition curve for D-TC than did suramin. Because other purinergic 2X (P2X) 39 receptor antagonists, NF023 and NF279, do not have the reverse effects on the neuromuscular blockade of 40 D-TC, the effect of NF449 seems irrelevant to inhibition of P2X receptors.

Significance: These data suggest that NF449 was able to compete with the binding of D-TC on the nicotinic ACh 42 receptors, and the effect of NF449 was more potent than suramin in reducing the inhibition of D-TC. The 43 structure of NF449 may provide useful information for designing potent antidotes against neuromuscular 44 toxins.

© 2011 Published by Elsevier Inc. 46

48

Introduction

The neuromuscular junction consists of the presynaptic motor nerve terminal and the motor end-plate of skeletal muscle. Acetylcholine (ACh) is synthesized within the presynaptic terminal and stored in vesicles (Huh and Fuhrer, 2002; Fagerlund and Eriksson, 2009). The arrival of an action potential at the presynaptic terminal leads to the release of ACh that then diffuses across the junctional cleft and binds to nicotinic ACh (nACh) receptors at the postsynaptic

0024-3205/\$ - see front matter © 2011 Published by Elsevier Inc. doi:10.1016/j.lfs.2011.03.013

membrane. Some chemical compounds can interrupt the neuromus- 59 cular transmission and cause muscle relaxation. The studies of the 60 structure-activity relationship of agonists and antagonists have 61 revealed how they bind to orthostatic binding sites (Albuquerque 62 et al., 2009; Zouridakis et al., 2009; Azam and McIntosh, 2009). 63 Previous studies showed that suramin not only reversed the effects of 64 neuromuscular blockers (Henning et al., 1993) but also inhibited the 65 prejunctional Ca²⁺ channels (Henning et al., 1996). However, the 66 mechanism of effect of suramin on the neuromuscular junction is still 67 unclear. Suramin is a polysulphonated naphthylurea derivative that is 68 used, as a therapeutic agent in the treatment of human African 69 trypanosomiasis (sleeping sickness) and onchocerciasis (Schneider, 70 1960; Schulz-Key et al., 1985). Recent studies have shown that 71 suramin possesses a variety of biological activities. The compound can 72 inhibit reverse transcriptase and prevent HIV entry into the cell 73 (Jentsch et al., 1987), and it has been used experimentally in the 74

^{*} Corresponding author at: Department of Biomedical Sciences, Chung Shan Medical University, No.110, Sec.1, Jianguo N. Rd., Taichung City 402, Taiwan. Fax: +886 4 23803865.

E-mail address: mjl@csmu.edu.tw (M.-J. Lin).

ARTICLE IN PRESS

T.-R. Su et al. / Life Sciences xxx (2011) xxx-xxx



Q1 Fig. 1. The reverse effect of NF449 on the neuromuscular blockade induced by D-tubocurarine (D-TC) in the mouse phrenic nerve-diaphragm. Pretreatment with NF449 (100 μM) can prevent the block of nerve-stimulated muscle contractions induced by D-TC (5 μM) (A). Pretreatment with vehicle served as a control for the comparison (B). The partial inhibition of nerve-evoked muscle contractions induced by a lower concentration of D-TC (1.2 μM) can also be reversed by the after-treatment with NF449 (C). The blocking effects of D-TC (5 μM) were reversible with washout (D, B). Calibrations: 5 min; 0.5 g.

treatment of cancer (Dhar et al., 2000; Ord et al., 2005; Villalona-

Calero et al., 2008). Suramin is also an antagonist of P2 purinoceptors

77 (Dunn and Blakeley, 1988; Inoue et al., 1991). Previously, we have

78 demonstrated that suramin can reverse the inhibitory action of a

⁷⁹ tripeptide, carbobenzoxy₁Gly-Gly-Arg₁ β -naphthylamide, which acts

both at the postsynaptic nACh receptors and presynaptic autorecep- 80 tors (Lin-Shiau and Lin, 1998). Recently, a series of novel suramin 81 analogues have been developed (Kassack et al., 2004; Ullmann et al., 82 2005) that are specifically designed for antitumor activity (Krejci 83 et al., 2010). Most of these analogues have a symmetrical structure 84

t1.1 Table 1

The blocking effect of D-tubocurarine (D-TC) on the nerve-evoked muscle contractions of mouse phrenic nerve-diaphragm pretreated with suramin and suramin analogues (NF007, NF023, NF279 and NF449). Suramin and suramin analogues were pretreated for 15–25 min prior to the application of D-TC (5 μM).

t1.3	Treatment	п	Time to complete blockade (min)	The percentage of inhibition of amplitude (before/20 min after the application of D-TC)
t1.4	Group 1			
t1.5	Pretreatment with NF007 $(100 \mu\text{M}) + \text{D-TC}$	5	3.2 ± 0.8	Complete blockade [†]
t1.6	D-TC alone	5	2.8 ± 0.5	Complete blockade
t1.7				
t1.8	Group 2			
t1.9	Pretreatment with NF023 $(100 \mu\text{M}) + \text{D-TC}$	4	7.7 ± 0.3	Complete blockade
t1.10	D-TC alone	4	3.4 ± 0.8	Complete blockade
t1.11				
t1.12	Group 3			
t1.13	Pretreatment with NF279 $(100 \mu\text{M}) + \text{D-TC}$	4	4.3 ± 0.6	Complete blockade
t1.14	D-TC alone	4	3.6 ± 0.6	Complete blockade
t1.15				
t1.16	Group 4		405.04*	
t1.17	Pretreatment with Suramin $(100 \mu\text{M}) + \text{D-1C}$	11	18.5 ± 3.1	Complete blockade
t1.18	D-IC alone	11	4.1 ± 0.5	Complete blockade
t1.19	Group F			
t1.20 +1.21	Bretreatment with NE449 (30 μ M) \pm p-TC	4	Without complete blockade [#]	$75.1 \pm 5.09^{*}$
61.21 +1.00	p_{TC} alone	4		$75.1 \pm 5.5\%$
+1.22	b-re alone	4	4.4 <u>+</u> 0.0	complete blockade
t1.25	Group 6			
t1.24	Pretreatment with NF449 $(100 \mu M) + p-TC$	9	Without complete blockade ^{#,*}	$10.7 \pm 6.8\%^*$
t1.26	p-TC alone	9	5.0 ± 0.8	Complete blockade

Data are presented as mean \pm S.E.M.

t1.27 The experimental animals were divided into six groups. The *diaphragm of each* individual animal *was divided* into *two* equal parts, one is pretreatment with the suramin analogue and the other is a treatment for D-TC only (pair study).

t1.29 * P < 0.05 as compared with D-TC alone within each group.

t1.30 [#] Without complete blockade: the nerve-evoked muscle contractions cannot be blocked completely.

t1.31 ^{\dagger} Complete blockade = 100% inhibition of nerve-evoked muscle contractions.

with a central urea bridge except for NF007. Amongst these analogues, 85 86 the structure of NF279 is very similar to suramin. These analogues are polysulphonates with six negative charges for suramin and NF279 and 87 88 eight negative charges for NF449. Therefore, we investigated the effects of suramin analogues NF007, NF023, NF279 and NF449 on the 89 neuromuscular junctions. It turned out that NF449, but not the other 90 analogues, was an effective protecting agent against the inhibition of 91 D-tubocurarine (D-TC) on the nACh receptor, and its effect was more 9293 potent than that of suramin.

94 Materials and methods

95 Mouse phrenic nerve-diaphragm preparations

Mice of the ICR strain (17-22 g) were anaesthetised with carbon 96 dioxide followed by cervical dislocation. Phrenic nerve-diaphragm 97 preparations were isolated and suspended in 10 ml of modified 98 Krebs solution with the compositions (in mM): NaCl 131, KCl 4.8, 99 MgSO₄ 1.2, CaCl₂ 2.5, NaHCO₃ 12.5 and glucose 11. The modified 100 Krebs solution was saturated with carbogen (95% $O_2/5\%$ CO_2) and 101 kept at a constant pH (7.3-7.4). Muscle contractions were elicited 102 by electrical stimulation of the phrenic nerve with supramaximal 103 104 rectangular pulses of 0.05 ms in duration at a frequency of 0.1 Hz and recorded by the isometric transducer on a Gould TA240 105 polygraph. The muscle resting tension was adjusted to 1 g and the 106 preparation was allowed to equilibrate for 20-30 min before 107 starting the experimental protocol. 108

109 Chick biventer cervicis muscle preparations

The preparations were isolated from baby Leghorns, 4–7 days old, 110 111 according to previously described methods (Ginsborg and Warriner, 1960). The organ bath contained 10 ml modified Krebs solution. The 112contraction of the muscle was recorded isometrically with 0.5 g initial 113tension. Each muscle contraction was elicited by the addition of 30 µM 114 ACh to the organ bath for about 1 min followed by the washout of ACh. 115 The interval between applications of ACh was 20 min to prevent 116 desensitisation of the nACh receptors. 117

118 Miniature end-plate potential and end-plate potential recordings

119 Miniature end-plate potential (mepp) and end-plate potential (epp) were measured by an intracellular glass microelectrode with a 120high impedance amplifier (Axoclamp 2B) in bridge mode. The 121 microelectrodes were filled with 3 M KCl resistance of $5-12 \text{ M}\Omega$. 122 Epps were evoked by stimulation of the phrenic nerve using a bipolar 123124suction electrode at a frequency of 0.1 Hz with 0.02 ms supramaximal rectangular pulses (A-M systems model 2100 stimulator), 125and the diaphragm was immobilised by the cut muscle method 126(Barstad and Lilleheil, 1968). The signals of epps, mepps and 127presynaptic currents were digitalised (Digidata 1440A and pClamp 128 12910, Axon Instruments) and stored for later analysis (pClamp-Clampfit 13010, Axon Instruments).

Mouse triangularis sternus preparation and nerve terminal waveform recordings

The thin layer (2-3 muscle layers) of mouse left triangularis 133 sterni intercostal nerve-muscle preparation was isolated according 134to the method described previously (McArdle et al., 1981; Mallart, 1351985). The isolated preparations were pinned out in the Sylgard-136 coated glass chamber 1-2 ml and visualised at 400× magnification 137 by a Zeiss microscope (Axioskop FS plus). Preparations were 138 continuously perfused at a rate of 2-4 ml/min with an oxygenated 139(95% O₂ plus 5% CO₂) modified Krebs solution containing (mM): 140 141 NaCl 131, KCl 4.8, MgSO₄ 1.2, CaCl₂ 2.5, NaHCO₃ 12.5 and glucose 11, pH 7.2_7.4. The intercostal nerves were stimulated by a suction 142 electrode using supramaximal voltage and square-wave pulses of 143 0.02 ms in duration at 0.2 Hz ($A_{\rm T}$ M systems model 2100 stimulator). 144 Presynaptic waveforms were obtained from extracellular recording 145 close to the nerve terminal endings of intercostal nerves with glass 146 microelectrodes filled with 2 M NaCl (7–13 M Ω) placed inside the 147 perineural sheath (Mallart, 1985). Postsynaptic activity was blocked 148 by adding 20–30 μ M D-TC to the bathing medium. The signals of the 149 presynaptic waveforms were displayed and analysed on pClamp 10 150 software. All animal care was performed in accordance with the 151 guidelines of the Committee of the CSMU as previously described 152 (Su et al., 2009). 153

Suramin (8, 8[']₁₇[Carbonyl*bis* [imino-3,1-phenylenecarbonylimino 155 (4-methyl-3,1-phenylene) carbonylimino]] *bis*-1,3,5-naphthalenetri- 156 sulphonic acid hexasodium salt), NF023 (8, 8 (carbonylbis (imino-3,1-157 phenylenecarbonylimino)) bis-(1,3,5-naphthalenetrisulfonicacid)), 158



Fig. 2. The inhibition curves of nerve-evoked muscle contractions induced by D-tubocurarine in the mouse phrenic nerve-diaphragm pretreated with or without NF449 (or suramin). The pretreatment with NF449 100 μ M (or suramin 100 μ M) significantly prevented the decrease in nerve-evoked muscle contractions induced by D-TC (5 μ M) treatment (A). The partial inhibition of muscle contractions induced by 1.2 μ M D-TC were reversed with the after-treatment of NF449 100 μ M (or suramin 100 μ M) (B). The arrow at time 0 min indicates the application of suramin analogues (A) and D-TC (A) and suramin analogues (B), respectively. NF449 caused a greater rightward shift of the concentration-inhibition curve of D-TC than did suramin (C).

154

ARTICLE IN PRESS

T.-R. Su et al. / Life Sciences xxx (2011) xxx-xxx

NF279 (8, 8/_ (carbonylbis (imino-4,1-phenylenecarbonylimino-4,1-159 160 phenylenecarbonylimino)) bis (1,3,5-naphthalenetrisul fonic acid)) and NF449 (4, 4', 4", 4", carbonylbis (imino-5,1,3-benzenetriylbis-161 162(carbonylimino))) tetrakis-benzene-1,3-disulphonic acid octasodium salt) were purchased from Tocris biosciende Inc. (UK). NF007 (8-(3-163 nitrobenzamido)-1,3,5-naphthalenetrisulphonic acid) was purchased 164 from Calbiochem Inc. (USA). D-TC was purchased from Fluka Inc. 165166 (USA).

167 Statistics

The data are given as mean \pm S.E.M. The statistical significance of differences was evaluated using a paired or unpaired Student's *t*-test. When more than one group was compared with one control, significance was evaluated using one-way analysis of variance (ANOVA). Probability values (*P*) less than 0.05 were considered 172 significant.

174

175

Results

Effects of NF449 on nerve-stimulated muscle contractions

As compared to the use of vehicle alone (Fig. 1B), pretreatment 176 with NF449 (100 μ M) can prevent the block of muscle contraction by 177 D-TC (Fig. 1A). Another experiment showed that the partial inhibition 178 of muscle contraction caused by 1.2 μ M D-TC (Fig. 1C and D) can be 179 reversed by subsequent treatment with NF449 (Fig. 1C). All effects of 180 the blockade were reversible by washout. Pretreatment with other 181 suramin analogues, NF007, NF023 and NF279, were unable to prevent 182 the block of nerve-evoked muscle contractions induced by D-TC 183



Fig. 3. Effect of pretreatment with NF449 on the inhibition of end-plate potentials (epp) or miniature end-plate potentials (mepp) induced by D-TC. (A) Representative epp traces recorded from mouse phrenic nerve diaphragm preparation. (B) The summarised plots of the D-TC-inhibition curves are shown for D-TC alone, NF449 and suramin pretreatment. The arrow indicates the application of NF449 or suramin or vehicle. Calibrations: 4 ms; 2 mV. (C) The recording traces showed the mepp amplitudes by addition of D-TC alone, and pretreatment with suramin or NF449 followed by addition of D-TC. The statistical bar chart showed the reversal effect of suramin and NF449 on the inhibition of mepp amplitudes induced by D-TC. Calibrations: 10 s; 1 mV.

T.-R. Su et al. / Life Sciences xxx (2011) xxx-xxx



Fig. 4. Effects of the pretreatment with NF449 on the muscle contractions of chick biventer cervicis induced by ACh (30μ M). The representative traces of muscle contractions induced by ACh are shown in (A). ACh-induced muscle contractions caused by the application of ACh (30μ M) and then washout after the muscle contractions reached the maximal response. The upright arrow and down arrow indicate the application of ACh and washout (W), respectively. The effect of NF449 or suramin alone on the ACh-induced muscle contractions is summarised in (B). Pretreatment with NF449 or suramin on ACh-induced contractions in the presence D-TC is summarised in (C).* *P*<0.05 as compared with D-TC alone. Calibrations: 10 s; 1 g.

(Table 1). The time course of inhibition curves of D-TC when pretreated or after-treated with NF449 are shown in Fig. 2A and B, respectively. Fig. 2C shows the concentration—inhibition curve of D-TC. The IC₅₀ (inhibition concentration 50%) of D-TC alone was about 1.2 μ M. Suramin and NF449 significantly shifted the curves to the right (IC₅₀ from ~1.2 to ~4 μ M for suramin and from ~1.2 to ~8 μ M for NF449).

191 Effects of NF449 on the end-plate potential (epp) and miniature end-192 plate potential (mepp)

193Application of D-TC (5 µM) to the extracellular solution produced a complete blockade of epps within 5 min (Fig. 3A). The time course of 194inhibition curves showed that pretreatment with either NF449 or 195suramin significantly reduced the percent inhibition of the epps induced 196 by D-TC (Fig. 3B). The percent inhibitions of epps for D-TC 197198alone, pretreatment with suramin and NF449 were 0%, $43.9 \pm 10.1\%$ 199and $61.4 \pm 8.9\%$, respectively at 10 min after the application of D-TC. Similarly, NF449 and suramin significantly reduced the inhibition of 200 mepps induced by D-TC (data not shown). D-TC alone produced a 201complete inhibition of mepps after 5 min of incubation. The 202percent inhibitions of mepps (amplitude of before/after the D-TC) 203were $50.2 \pm 4.1\%$ for suramin and $66.1 \pm 5.2\%$ for NF449. NF449 and 204 suramin did not change the frequency of mepps (NF449: 0.95 ± 0.08 Hz; 205 suramin: 0.88 ± 0.07 Hz; and control: 0.97 ± 0.08 Hz). 206

207 Effect of NF449 on the ACh-induced contraction in the chick biventer 208 cervicis muscle

The contractions of chick biventer cervicis muscle (4–7 days old) can be induced by the addition of ACh (Fig. 4A). The time interval between each application was 20 min to prevent the nicotinic 211 receptors from desensitising. Only the muscle preparations with less 212 than 5% amplitude variance between two applications of ACh 213



Fig. 5. Effects of NF449 and suramin on the presynaptic terminal waveforms. Treatment with NF449 (A, 100 μ M) and suramin (B, 100 μ M) did not affect the presynaptic Na⁺ and K⁺ current waveforms. The K⁺ channel blocker, 4-aminopyridine (100 μ M), showed inhibition of the K⁺ current waveform as a comparison (C). Calibrations: 2 ms; 2 mV.

ARTICLE IN PRESS

T.-R. Su et al. / Life Sciences xxx (2011) xxx-xxx

 $(30 \,\mu\text{M})$ were used in the further experiments. The ACh-induced 214 215 contraction was significantly inhibited by the addition of D-TC (5 μ M; D-TC alone, 49.8 \pm 3.3% of control). Treatment with either 216 217NF449 or suramin alone produces only slight changes in the amplitudes of ACh-induced contraction (Fig. 4B). The pretreatment 218 with either NF449 or suramin significantly reduced the inhibition of 219the ACh-induced contractions by D-TC (Fig. 4C; suramin: $67.8 \pm 3.4\%$ 220 of control; NF 449: 93.3 \pm 7.6% of control; *P*<0.05 as compared with 221 222 D-TC alone).

223 Effects of NF449 on the presynaptic terminal waveforms

To determine whether NF449 and suramin can affect the presynaptic currents, we examined the presynaptic terminal waveforms with the application of NF449 or suramin. The results showed that NF449 (Fig. 5A) and suramin (Fig. 5B) did not affect the presynaptic Na⁺ and K⁺ waveforms. The effect of 4-aminopyridne (50 μ M), a K⁺ channel blocker, acts as the positive control for this comparison (Fig. 5C).

231 Discussion

In the present study, we demonstrated that NF449 prevents the
 neuromuscular blockade induced by D-TC via nicotinic ACh (nACh)
 receptors. NF449 can significantly reduce the inhibition of D-TC on the

amplitudes of nerve-stimulated muscle contractions, end-plate 235 potentials (epps) and ACh-induced contractions. Furthermore, 236 NF449 did not affect the presynaptic Na⁺ and K⁺ waveforms. All 237 the evidence indicated that the effect of NF449 was that it could 238 compete with the binding of D-TC to the nACh receptors. 239

It has been reported that NF449 is a selective P2X purinergic 240 receptor antagonist (Braun et al., 2001; Kassack et al., 2004), 241 fibroblast growth factor receptor 3 (FGFR3) inhibitor (Gunosewoyo 242 and Kassiou, 2010; Krejci et al., 2010) and Gs protein inhibitor 243 (Hohenegger et al., 1998). Furthermore, suramin and suramin 244 analogue NF023 act as direct antagonists of heterotrimeric G 245 proteins by blocking of the rate-limiting step of G protein activation 246 (Beindl et al., 1996). Because other P2X receptor antagonists 247 (NF007, NF023 and NF279) (Van Rhee et al., 1994; Damer et al., 248 1998; Soto et al., 1999), FGFR inhibitors (NF279) (Krejci et al., 2010) 249 and G protein uncouplers (NF023) (Beindl et al., 1996) cannot 250 reverse the inhibitory effect induced by D-TC (Table 1), the effect of 251 NF449 on the neuromuscular junction is not likely related to its 252 effects on P2X, FGFR3 receptors or G protein.

In this study, NF449 protects the cholinergic receptor from the 254 action of D-TC and this suggests that the pharmacological competition 255 between these agents reflects a common binding site on the receptor 256 protein (Miledi and Potter, 1971). The structure of suramin analogues 257 and D-TC are shown in Fig. 6. NF449, NF023 and NF279 exhibit close 258 structural similarity with suramin that belongs to a group of 259



Fig. 6. The structures of NF449, NF007, NF023, NF279, suramin and ntubocurarine. NF007: 8-(3-nitrobenzamido)-1,3,5-naphthalenetrisulphonic acid; NF023: 8,8 -[carbonylbis (imino-3,1-phenylenecarbonylimino)]bis-1,3,5-naphthalenetrisulphonic acid; NF279: 8,8'-[carbonylbis(imino-4,1-phenylenecarbonylimino-4,1-phenylenecarbonylimino)]bis-1,3,5-naphthalenetrisulphonic acid; NF449: 4,4',4'',4'''-(carbonylbis (imino-5,1,3-benzenetriylbis-(carbonylimino)))tetrakis-benzene-1,3-disulphonic acid octasodium salt; Suramin: 8,8'-[Carbonylbis [imino-3,1-phenylenecarbonylimino(4-methyl-3,1-phenylene)carbonylimino]]bis-1,3,5-naphthalenetrisulphonic acid hexasodium salt;

T.-R. Su et al. / Life Sciences xxx (2011) xxx-xxx

polysulphonated napthylurea derivatives (Kassack et al., 2004). How 260 does NF449 act on the nicotinic ACh receptor but NF007, NF023 and 261 262 NF279 do not? Further investigation is needed to find the information 263concerning the structural determinants of ligand-receptor interactions (Horti et al., 2010). The structure of NF279 is the most similar to 264suramin, both are symmetrical polysulphonated naphthylamine 265derivatives of urea. The only slight difference between suramin and 266 NF279 is that NF279 lacks two methyl groups located on the benzene 267268moiety as compared with suramin. It has been reported that NF279 is a novel and potent antagonist selective for P2X₁ (Rettinger et al., 2692702000). However, suramin, but not NF279, prolonged the blocking time 271caused by D-TC. Another suramin analogue, NF449, is also a potent 272antagonist selective for P2X₁ (Hülsmann et al., 2003). In contrast to 273 NF279, the treatment with NF449 can significantly prevent the blockade of nerve-evoked muscle contractions induced by D-TC. 274NF449 caused approximately a 5-fold (~1.2 µM to 8 µM) rightward 275 shift in the concentration-inhibition curve of D-TC (Fig. 2), and its 276 effect on the neuromuscular junction is more potent than that of 277suramin. Based on the structure-activity relationship of NF449 at P2 278receptors, any deletion or change of position of the sulphonic acid 279groups, or replacing the central urea bond by the terephthalic acid 280bisamides reduced the potency at P2X₁ by at least 90% (Inoue et al., 281282 1991). We don't know whether the groups of the central urea and 283 sulphonic acid are important for the effect of NF449 on the nACh receptor. NF449 and suramin have a symmetrical structure with a 284central urea group and numbers of negatively charged sulphonate 285groups on the benzene or naphthyl rings. Based on the observation 286287that the asymmetrical structure of NF007 does not have an effect on the neuromuscular junction, the symmetrical polysulphonated naph-288 thylamine is likely important for the protecting effect on the nACh 289receptor by preventing neuromuscular block induced by D-TC. 290

A previous report showed that a stimulatory effect of suramin on the growth factor receptor indicated suramin is both a partial agonist and competitive inhibitor for the high affinity NGF receptor (Gill et al., 1996). Because NF449 and suramin have a slight inhibition of ACh-induced muscle contractions in chick biventer cervicis muscle, NF449 and suramin could possibly act as partial antagonists for the nACh receptor.

297 In summary, NF449 can prevent the blockade of neuromuscular transmission induced by D-TC. The effect of NF449 is to compete with the 298binding of D-TC on the nACh receptor site, and its effect is more potent 299than that of suramin. Even though this study provides evidence from 300 animal experiments, the findings seem to have a potential clinical 301 relevance. Nondepolarising neuromuscular blockers (muscle relaxants) 302 are widely used as an adjunct for general anaesthesia or an anticonvul-303 sant in clinical practice. It is suggested that NF449 may be useful in the 304 305 treatment of muscle relaxants (nACh receptor blockers) overdose. The 306 chemical structure of NF449 may provide important information in developing more effective antidotes against neuromuscular toxins. 307

308 Conflict of interest statement

309 None.

310 Acknowledgements

These studies were supported by grants from the National Science Council of Taiwan (NSC 97-2320-B-040-002-MY3) and Chung Shan Medical University (CSMU–TSMH-098-002).

314 References

- 315
 Albuquerque EX, Pereira EF, Alkondon M, Rogers SW. Mammalian nicotinic acetylcho

 316
 line receptors: from structure to function. Physiol Rev 2009;89(1):73-120.
- Azam L, McIntosh JM. Alpha-conotoxins as pharmacological probes of nicotinic acetylcholine receptors. Acta Pharmacol Sin 2009;30(6):771–83.

Barstad JA, Lilleheil G. Transversely cut diaphragm preparation from rat, an adjuvant 319 tool in the study of the physiology and pbarmacology of the myoneural junction. 320 Arch Int Pharmacodyn Ther 1968;175(2):373–90. 321

Beindl W, Mitterauer T, Hohenegger M, Ijzerman AP, Nanoff C, Freissmuth M. Inhibition 322 of receptor/G protein coupling by suramin analogues. Mol Pharmacol 1996;50(2): 323 415–23. 324

- Braun K, Rettinger J, Ganso M, Kassack M, Hildebrandt C, Ullmann H, et al. NF449: a 325 subnanomolar potency antagonist at recombinant rat P2X1 receptors. Naunyn 326 Schmiedebergs Arch Pharmacol 2001;364(3):285–90. 327
- Damer S, Niebel B, Czeche S, Nickel P, Ardanuy U, Schmalzing G, et al. NF279: a novel 328 potent and selective antagonist of P2X receptor-mediated responses. Eur J 329 Pharmacol 1998;350(1):R5–6. 330
- Dhar S, Gullbo J, Csoka K, Eriksson E, Nilsson K, Nickel P, et al. Antitumour activity of 331

 suramin analogues in human tumour cell lines and primary cultures of tumour cells

 from patients. Eur J Cancer 2000;36(6):803–9.
- Dunn PM, Blakeley AG. Suramin: a reversible P2-purinoceptor antagonist in the mouse 334 vas deferens. Br J Pharmacol 1988;93(2):243–5. 335
- Fagerlund MJ, Eriksson LI. Current concepts in neuromuscular transmission. Br J 336 Anaesth 2009;103(1):108–14. 337
- Gill JS, Connolly DC, McManus MJ, Maihle NJ, Windebank AJ. Suramin induces 338 phosphorylation of the high-affinity nerve growth factor receptor in PC12 cells 339 and dorsal root ganglion neurons. J Neurochem 1996;66(3):963–72. 340
- Ginsborg BL, Warriner J. The isolated chick biventer cervicis nerve_muscle preparation. 341 Br J Pharmacol Chemother 1960;15:410–1. 342
- Gunosewoyo H, Kassiou M. P2X purinergic receptor ligands: recently patented 343 compounds. Expert Opin Ther Pat 2010;20(5):625–46. 344
- Henning RH, Nelemans A, Houwertjes M, Agoston S. Reversal by suramin of 345 neuromuscular block produced by pancuronium in the anaesthetized rat. Br J 346 Pharmacol 1993;108(3):717-20. 347
- Henning RH, Rowan EG, Braga MF, Nelemans A, Harvey AL. The prejunctional inhibitory 348 effect of suramin on neuromuscular transmission in vitro. Eur J Pharmacol 349 1996;301(1–3):91–7. 350
- Hohenegger M, Waldhoer M, Beindl W, Boing B, Kreimeyer A, Nickel P, et al. 351 Gsalpha-selective G protein antagonists. Proc Natl Acad Sci USA 1998;95(1): 352 346–51. 353
- Horti AG, Gao Y, Kuwabara H, Dannals RF. Development of radioligands with optimized 354 imaging properties for quantification of nicotinic acetylcholine receptors by 355 positron emission tomography. Life Sci 2010;86(15–16):575–84. 356
- Huh KH, Fuhrer C. Clustering of nicotinic acetylcholine receptors: from the 357 neuromuscular junction to interneuronal synapses. Mol Neurobiol 2002;25(1): 358 79-112. 359
- Hülsmann M, Nickel P, Kassack M, Schmalzing G, Lambrecht G, Markwardt F. NF449, a 360 novel picomolar potency antagonist at human P2X1 receptors. Eur J Pharmacol 361 2003;470(1–2):1–7. 362
- Inoue K, Nakazawa K, Ohara-Imaizumi M, Obama T, Fujimori K, Takanaka A. Selective 363 and competitive antagonism by suramin of ATP-stimulated catecholamine- 364 secretion from PC12 phaeochromocytoma cells. Br J Pharmacol 1991;102(3): 365 581–4. 366
- Jentsch KD, Hunsmann G, Hartmann H, Nickel P. Inhibition of human immunodeficiency virus type I reverse transcriptase by suramin-related compounds. J Gen Virol 368 1987;68(Pt 8):2183–92. 369
- Kassack MU, Braun K, Ganso M, Ullmann H, Nickel P, Böing B, et al. Structureactivity relationships of analogues of NF449 confirm NF449 as the most potent and selective known P2X1 receptor antagonist. Eur J Med Chem 2004;39(4): 372 345–57. 373
- Krejci P, Murakami S, Prochazkova J, Trantirek L, Chlebova K, Ouyang Z, et al. NF449 374
 is a novel inhibitor of fibroblast growth factor receptor 3 (FGFR3) signaling 375
 active in chondrocytes and multiple myeloma cells. J Biol Chem 2010;285(27): 376
 20644–53. 377
- Lin-Shiau SY, Lin MJ. Studies on curare-like action of the tripeptide carbobenzoxy-Gly- 378 Gly-Arg-beta-naphthylamide in mouse diaphragm. Eur J Pharmacol 1998;343(1): 379 51–6. 380
- Mallart A. Electric current flow inside perineurial sheaths of mouse motor nerves. J 381 Physiol 1985;368:565–75. 382
- McArdle JJ, Angaut-Petit D, Mallart A, Bournaud R, Faille L, Brigant JL. Advantages of the 383 triangularis sterni muscle of the mouse for investigations of synaptic phenomena. J 384 Neurosci Methods 1981;4(2):109–15. 385
- Miledi R, Potter LT. Acetylcholine receptors in muscle fibres. Nature 1971;233(5322): 386 599–603. 387
- Ord JJ, Streeter E, Jones A, Le Monnier K, Cranston D, Crew J, et al. Phase I trial of 388 intravesical Suramin in recurrent superficial transitional cell bladder carcinoma. Br 389 J Cancer 2005;92(12):2140–7. 390
- Rettinger J, Schmalzing G, Damer S, Müller G, Nickel P, Lambrecht G. The suramin 391 analogue NF279 is a novel and potent antagonist selective for the P2X1 receptor. 392 Neuropharmacology 2000;39(11):2044–53. 393
- Schneider J. Treatment of African trypanosomiasis (sleeping sickness). Presse Med 394 1960;68:881–3. 395
- Schulz-Key H, Karam M, Prost A. Suramin in the treatment of onchocerciasis: the 396 efficacy of low doses on the parasite in an area with vector control. Trop Med 397 Parasitol 1985;36(4):244–8. 398
- Soto F, Lambrecht G, Nickel P, Stühmer W, Busch AE. Antagonistic properties of the 399 suramin analogue NF023 at heterologously expressed P2X receptors. Neurophar-400 macology 1999;38(1):141–9. 401
- Su TR, Chen CH, Huang SJ, Lee CY, Su MC, Chen GH, et al. Functional study of the effect of 402 phosphatase inhibitors on KCNQ4 channels expressed in Xenopus oocytes. Acta 403 Pharmacol Sin 2009;30(9):1220–6. 404

T.-R. Su et al. / Life Sciences xxx (2011) xxx-xxx

 Ullmann H, Meis S, Hongwiset D, Marzian C, Wiese M, Nickel P, et al. Synthesis and structure_activity relationships of suramin-derived P2Y11 receptor antagonists with nanomolar potency. J Med Chem 2005;48(22):7040–8.
 Van Rhee AM, van der Heijden MPA, Beukers MW, Ijzerman AP, Soudijn W, Nickel P. Novel competitive antagonists for P2 purinoceptors. European J Pharmacology Molecular Pharmacology 1994;268(1):1–7. 405406 407

Villalona-Calero MA, Otterson GA, Wientjes MG, Weber F, Bekaii-Saab T, Young D, et al. 411 Noncytotoxic suramin as a chemosensitizer in patients with advanced non-small-

- 408 409 410
- cell lung cancer: a phase II study. Ann Oncol 2008;19(11):1903–9. 413 Zouridakis M, Zisimopoulou P, Poulas K, Tzartos SJ. Recent advances in understanding 414 the structure of nicotinic acetylcholine receptors. IUBMB Life 2009;61(4):407–23. 415

416