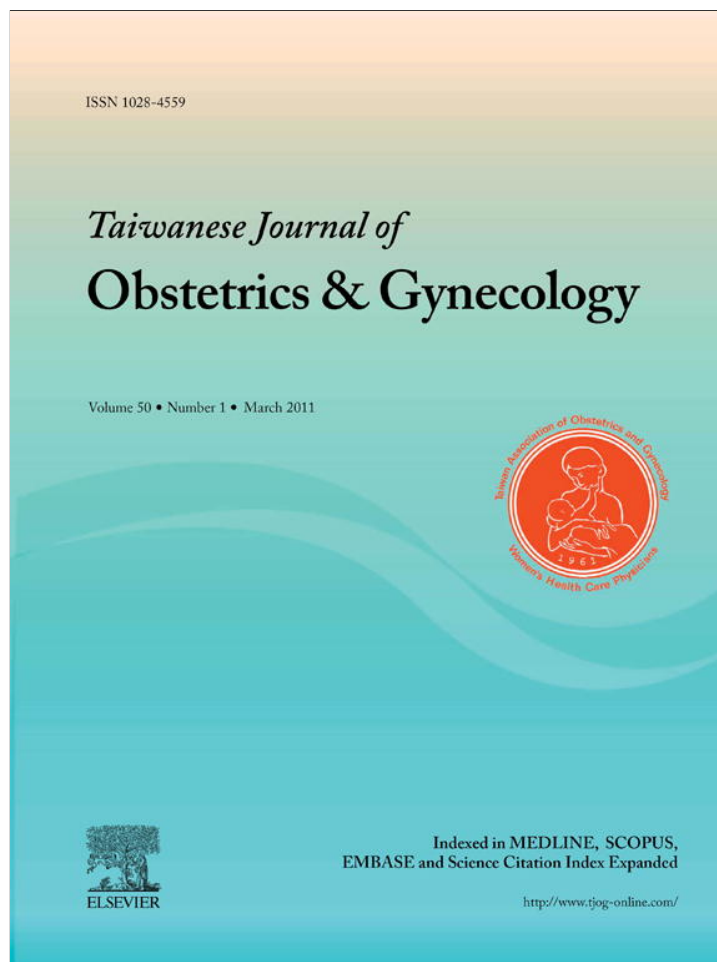


Provided for non-commercial research and education use.
Not for reproduction, distribution or commercial use.



This article appeared in a journal published by Elsevier. The attached copy is furnished to the author for internal non-commercial research and education use, including for instruction at the authors institution and sharing with colleagues.

Other uses, including reproduction and distribution, or selling or licensing copies, or posting to personal, institutional or third party websites are prohibited.

In most cases authors are permitted to post their version of the article (e.g. in Word or Tex form) to their personal website or institutional repository. Authors requiring further information regarding Elsevier's archiving and manuscript policies are encouraged to visit:

<http://www.elsevier.com/copyright>



Short Communication

Inv dup del(9p): Prenatal diagnosis and molecular cytogenetic characterization by fluorescence *in situ* hybridization and array comparative genomic hybridization

Chih-Ping Chen^{a,b,c,d,e,f,*}, Yi-Ning Su^g, Schu-Rern Chern^b, Chin-Yuan Hsu^a, Fuu-Jen Tsai^{d,h},
Pei-Chen Wu^a, Chen-Chi Lee^a, Yu-Ting Chen^b, Meng-Shan Lee^a, Wayseen Wang^{b,i}

^a Department of Obstetrics and Gynecology, Mackay Memorial Hospital, Taipei, Taiwan

^b Department of Medical Research, Mackay Memorial Hospital, Taipei, Taiwan

^c Department of Biotechnology, Asia University, Taichung, Taiwan

^d School of Chinese Medicine, College of Chinese Medicine, China Medical University, Taichung, Taiwan

^e Institute of Clinical and Community Health Nursing, National Yang-Ming University, Taipei, Taiwan

^f Department of Obstetrics and Gynecology, School of Medicine, National Yang-Ming University, Taipei, Taiwan

^g Department of Medical Genetics, National Taiwan University Hospital, Taipei, Taiwan

^h Department of Medical Research and Medical Genetics, China Medical University Hospital, Taichung, Taiwan

ⁱ Department of Bioengineering, Tatung University, Taipei, Taiwan

Accepted 11 November 2010

Abstract

Objective: To present molecular cytogenetic characterization of prenatally detected inverted duplication and deletion of 9p, or inv dup del(9p). **Materials, Methods, and Results:** A 35-year-old primigravid woman underwent amniocentesis at 16 weeks of gestation because of advanced maternal age. Amniocentesis revealed a derivative chromosome 9, or der(9) with additional material at the end of the short arm of one chromosome 9. Parental karyotypes were normal. Level II ultrasound showed ventriculomegaly and normal male external genitalia. Repeated amniocentesis was performed at 20 weeks of gestation. Array comparative genomic hybridization revealed a 0.70-Mb deletion at 9p24.3 and an 18.36-Mb duplication from 9p24.3 to 9p22.1. The distal 9p deletion encompassed the genes of *DOCK8*, *ANKRD15*, *FOXD4*, *DMRT1*, and *DMRT3*. Fluorescence *in situ* hybridization analysis using bacterial artificial chromosome clone probes specific for 9p confirmed that the der(9) was derived from the inv dup del(9p). The karyotype of the fetus was 46,XY,inv dup del(9)(:p22.1→p24.3::p24.3→qter)dn or 46,XY,der(9) del(9)(p24.3) inv dup(9)(p22.1p24.3) dn. Polymorphic DNA marker analysis determined a maternal origin of the inv dup del(9p). A 512-g male fetus was subsequently terminated at 22 weeks of gestation with facial dysmorphism. The fetus had normal male external genitalia without sex reversal.

Conclusion: Fluorescence *in situ* hybridization and array comparative genomic hybridization are useful to determine the nature of a prenatally detected aberrant chromosome derived from the inv dup del. Male fetuses with inv dup del(9p) and haploinsufficiency of *DMRT1* and *DMRT3* may present normal male external genitalia without sex reversal.

Copyright © 2011, Taiwan Association of Obstetrics & Gynecology. Published by Elsevier Taiwan LLC. All rights reserved.

Keywords: 9p; Array comparative genomic hybridization; Deletion; FISH; Inverted duplication; inv dup del(9p)

Introduction

An inverted duplication with a terminal deletion (inv dup del) is a rare complex chromosomal rearrangement that involves an inverted duplication of a part of a chromosome in association with a deletion distal to the site of duplication. The

* Corresponding author. Department of Obstetrics and Gynecology, Mackay Memorial Hospital, 92, Section 2, Chung-Shan North Road, Taipei, Taiwan.
E-mail address: cpc_mmh@yahoo.com (C.-P. Chen).

inv dup del has been reported in several chromosomes, such as 1p [1,2], 1q [3,4], 2p [5,6], 2q [7–10], 3p [11,12], 4p [13–15], 4q [16], 5p [17,18], 6p [19], 7q [20,21], 8p [22–29], 9p [30–35], 10p and 10q [20], 11p [36], 14q [37,38], 15q [39], 20p [40], 21q [41], and Xp [42,43].

Proposed mechanisms of the inv dup del include the U-type exchange model in which an intrachromosomal recombination and an end-to-end fusion occur in two homologous chromosomes resulting in a dicentric chromosome; and following division, the dicentric chromosome can break and result in a recombinant chromosome with an inverted duplication and a loss of chromosomal material distal to the site of recombination [44]; the nonallelic homologous recombination model mediated by an unequal crossover between inverted low-copy repeats [24]; and the premeiotic nonhomologous end joining model in which a terminal deleted chromosome is generated in the germline and passes through at least one breakage-fusion-bridge cycle leading to a sister chromatid fusion by a nonhomologous end joining and the production of gametes with terminal deletions and interrupted inverted duplications [1].

Prenatal diagnosis of the inv dup del(9p) has not been previously described. Here, we report prenatal diagnosis, molecular cytogenetic analysis, and perinatal findings of a male fetus with inv dup del(9p).

Materials, methods, and results

A 35-year-old primigravid woman underwent amniocentesis at 16 weeks of gestation because of advanced maternal age. Cytogenetic analysis then revealed a derivative chromosome 9, or der(9), with additional material at the end of the short arm of one chromosome 9. The parental karyotypes were normal. The woman requested repeated amniocentesis at 20 weeks of gestation, which revealed an inv dup del(9p) (Fig. 1). Using uncultured amniocytes, bacterial artificial chromosome (BAC)-based array comparative genomic hybridization (aCGH) (CMDX BAC-based aCGH CA3000 chips) (CMDX, Irvine, CA, USA) demonstrated a 9p deletion encompassing about 0.37 Mb from clone RP11-1112G24 to RP11-130C19

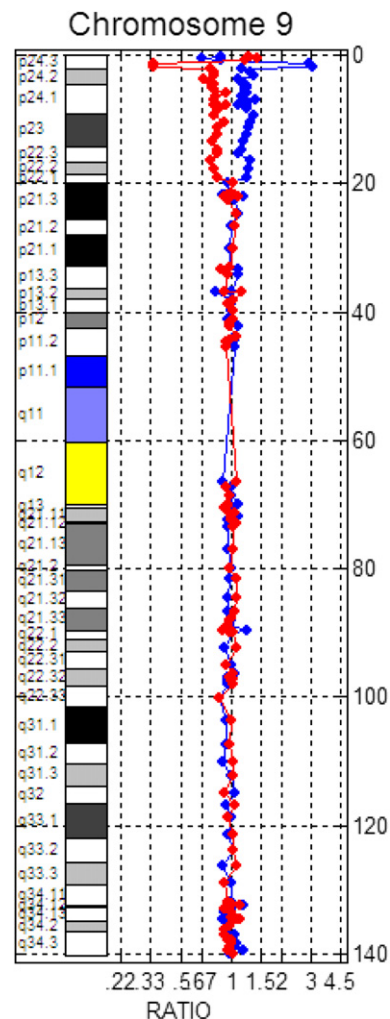


Fig. 2. BAC based-aCGH shows a deletion of terminal 9p [arr cgh 9p24.3p24.3 (RP11-1112G24→RP11-130C19)×1] and a duplication of distal 9p [arr 9p24.3p22.1 (RP11-690N7→RP11-322J7)×3]. aCGH = array comparative genomic hybridization; BAC = bacterial artificial chromosome.

and a 9p duplication encompassing about 17.87 Mb from clone RP11-690N7 to RP11-322J7. The result of BAC-aCGH was arr cgh 9p24.3p24.3 (RP11-1112G24→RP11-130C19)×1, 9p24.3p22.1 (RP11-690N7→RP11-322J7)×3 (Fig. 2).

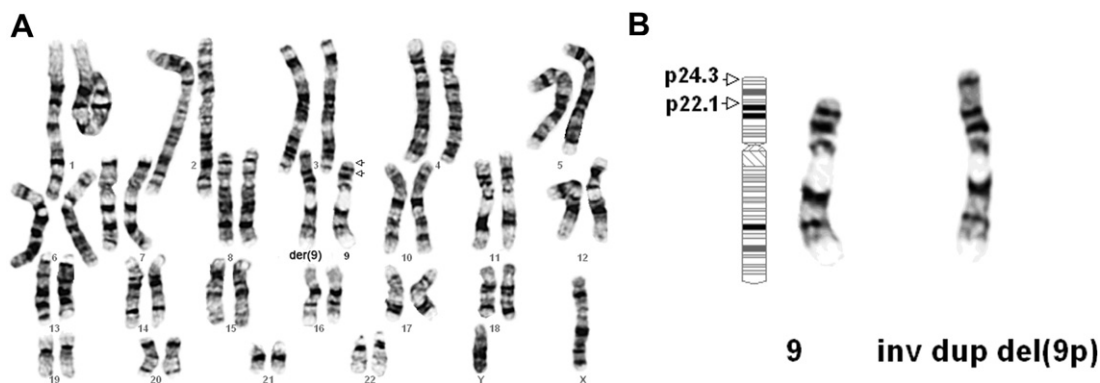


Fig. 1. (A) A karyotype 46,XY,inv dup del(9)(:p22.1→p24.3:p24.3→qter) or 46,XY,der(9)del(9)(p24.3) inv dup(9)(p22.1p24.3) in the fetus; (B) Partial G-banded karyotype of the fetus showing one normal chromosome 9 and one derivative chromosome 9, or der(9), with inv dup del(9p). The arrows indicate the breakpoints. inv dup del = inverted duplication with a terminal deletion.

For fluorescence *in situ* hybridization determination of the inv dup del in the der(9), the BAC clone probe mapping the genomic region of the distal chromosome 9p, and the telomeric region of 9p were used. The BAC clone probes RP11-32F11 (3,104,722–3,267,008) (spectrum green) at 9p24.2 and RP11-340N12 (17,136,369–17,298,494) (spectrum red) at 9p22.2 were used to determine the inverted duplication. The BAC clone probe RP11-31F19 (537,217–682,143) (spectrum green) at 9p24.3 and the Telomere 9q probe (TelVysion 9q; Vysis, Downers, Grove, IL, USA) (spectrum red) (control) were used to determine the terminal 9p deletion. Fluorescence *in situ* hybridization analysis showed an inverted duplication of distal 9p with an inverted duplication orientation of red-green-green-red (Fig. 3) and a terminal deletion with absence of a green signal on der(9) (Fig. 4). The karyotype of the fetus was 46,XY,inv dup del(9):(p22.1→p24.3::p24.3→qter)dn or 46,XY,der(9) del(9)(p24.3) inv dup(9)(p22.1p24.3)dn.

Level II ultrasound showed ventriculomegaly and normal male external genitalia at 21 weeks of gestation. The parents opted to terminate the pregnancy at 22 weeks of gestation. A 518-g male fetus was delivered with dysmorphism of hypertelorism, a prominent nose, and low-set ears (Fig. 5). The male external genitalia were normal (Fig. 6). Cytogenetic analysis of the cord blood confirmed the prenatal diagnosis. Using fetal blood, oligonucleotide-based (oligo) aCGH (SurePrint G3 Human CGH Microarray kit 60K; Agilent Technologies, Santa Clara, CA, USA) demonstrated a 0.70-Mb deletion at 9p24.3 (271,057–974,003) (NCBI build 36 March 2006) and an 18.36-Mb duplication from 9p24.3 to 9p22.1 (1,036,210–19,396,808) (Fig. 7). The result of oligo-aCGH was arr cgh 9p24.3p24.3 (271,057–974,003)×1, 9p24.3p22.1 (1,036,210–19,396,808)×3. Microsatellite analysis using the fetal and parental DNA demonstrated that the deletion and duplication of 9p were maternal in origin. In the duplicated segment of 9p,

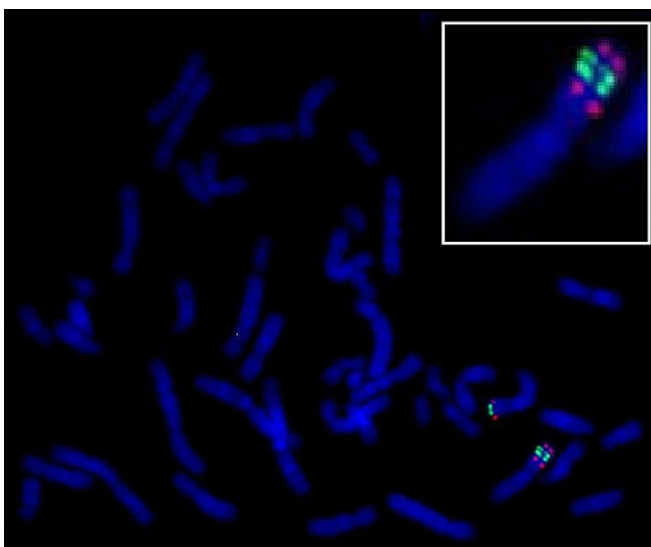


Fig. 3. FISH using BAC RP11-340N12 (red) at 9p22.2 and RP11-32F11 (green) at 9p24.2. The der(9) shows four signals in the order of red-green-green-red indicating an inverted duplication of 9p. BAC = bacterial artificial chromosome; FISH = fluorescence *in situ* hybridization.

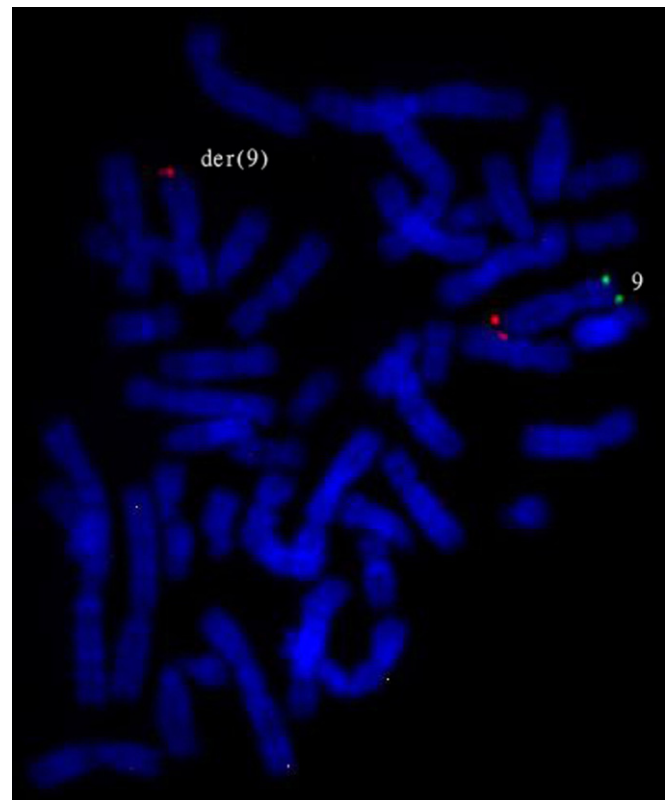


Fig. 4. FISH using BAC RP11-31F19 (green) at 9p24.3 and the Telomere 9q probe (TelVysion 9q) (red) shows absence of a green signal on der(9) indicating a terminal deletion of 9p. BAC = bacterial artificial chromosome; FISH = fluorescence *in situ* hybridization.

all the informative microsatellites were homozygous indicating an intrachromosomal event.

Discussion

We have reported an inverted duplication of the distal portion of the short arm of chromosome 9 (9p22.1→p24.3) and a deletion of the distal portion (9p24.3→pter) in a male fetus who manifested ventriculomegaly on prenatal ultrasound and facial dysmorphism at birth. The inv dup del(9p) was maternal in origin and intrachromosomal. The homozygosity throughout the duplicated segment implicates a possible U-type exchange mechanism, although other mechanisms cannot be completely excluded. To date, at least six cases with 9p duplication/deletion have previously been reported, and all were females. Teebi et al [30] first reported a 20-month-old female with an inverted duplication of proximal 9p and a deletion of distal 9p, or inv dup del(9):(p13→p22::p22→qter), prenatal intrauterine growth restriction, psychomotor developmental delay, a small umbilical hernia, and craniofacial dysmorphisms, including a narrow forehead with metopic ridging, a small anterior fontanelle, a small nose, midface hypoplasia, mild upward slant of palpebral fissures, mild epicanthic folds, thin and long eyebrows with mild synophrys, thin lips, and a short neck. The girl had an inverted duplication of proximal 9p (9p13→p22) and a deletion of distal 9p (p22→pter). In this case, during pregnancy, intrauterine



Fig. 5. The craniofacial appearance of the fetus at birth.

growth restriction was noted but a chromosome analysis of amniocytes was reportedly normal. Krepisch-Santos and Vianna-Morgante [31] reported a 16-year-2-month-old female with terminal deletion and an inverted duplication involving at least 9p23-p25.1. The duplication was paternal in origin. The girl manifested microcephaly, frontal bossing, a low-set frontal hairline, facial hirsutism, hypertelorism, deep-set and down-slanting eyes, epicanthus, a broad-based nose with a bulbous tip, a short and well-defined philtrum, downturned corners of the mouth, retrognathia, low-set large and protruding ears, hypoplastic nails, and bilateral single transverse creases. Her menstrual cycles were normal. Chabchoub et al [32] reported a 10-year-old female with mosaicism for inv dup del(9p) and a karyotype of 46,XX,del(9)(p22.1)/46,XX,der(9)t(5;9)

(p13.3;p22.1), del(9)(p22.1),dup(9) (p13.3→p22.1::p22.1→qter). The girl manifested psychomotor developmental delay, mild synophrys, hypoplastic alae nasi, long and smooth philtrum, a thin upper lip, small and dysmorphic ears, camptodactyly of the fifth fingers, upslanting palpebral fissures, prominent metopic suture, depressed and broad nasal root, plagiocephaly, a hypoplastic ectopic right kidney, and vesiculoureteral reflux. The inv dup del(9p) was paternal in origin. Swinkels et al [33] in 2008 reported a 2-year-old female (Case 7) with a deletion of 9p (p22.3→pter) and an inv dup(9)(p22.1p22.3). The girl manifested developmental speech and motor delay, hypotonia, short stature, trigonocephaly, upward slant short palpebral fissures, epicanthal folds, low-set posteriorly angulated ears, a short flat nose with anteverted nostrils, a flat philtrum, a thin upper lip, a high/narrow palate, broad internipple distance, tapering fingers, hyperconvex nails, and cardiac defects. Hulick et al [34] reported a 4-month-old female with a deletion of 9p (p24.2→pter) and an inv dup(9)(p21.3p24.2). The girl manifested hypotonia, growth and developmental delay, cleft palate, absent uvula, clinodactyly, large palpebral fissures, hypertelorism, a bulbous nose, abnormal ears, and dystrophic nails. Mosca et al [35] reported a 12-year-old female with 22q11.2 microdeletion and inv dup del(9p) consisting of a deletion region spanning 0.4–0.6 Mb and a duplication region spanning 1.6–11.8 Mb. The girl manifested mental retardation and asymmetric polymicrogyria predominantly affecting the right occipital lobe.

The present case was associated with partial trisomy 9p and ventriculomegaly. Gene dosage effect on chromosome 9p is responsible for normal development of the central nervous system. Partial trisomy 9p has been reported to be associated with abnormal neural migration, subcortical band heterotopia,



Fig. 6. The male external genitalia of the fetus at birth.

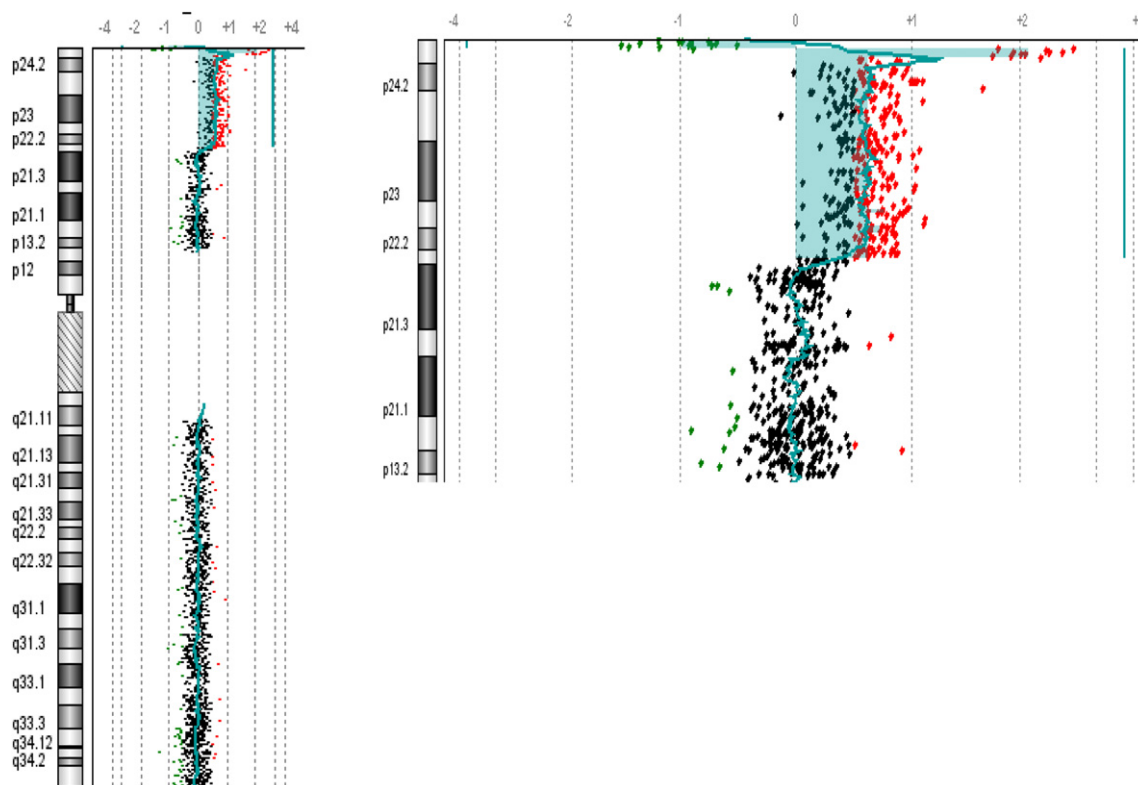


Fig. 7. Oligonucleotide-based aCGH shows a 0.70-Mb deletion at terminal 9p [arr cgh 9p24.3p24.3 (271,057–974,003)×1] and an 18.36-Mb duplication of 9p22.1→p24.3 [arr cgh 9p24.3p22.1 (1,036,210–19,396,808)×3]. aCGH = array comparative genomic hybridization.

Dandy-Walker malformation, ventriculomegaly, corpus callosum hypogenesis or agenesis, and polymicrogyria [35,45–49].

The present case was also associated with a distal 9p deletion encompassing the genes of *DOCK8*, *ANKRD15*, *FOXD4*, *DMRT1*, and *DMRT3*. Genetic aberrations in *DOCK8*, *ANKRD15*, and *FOXD4* may result in neurological and psychiatric disorders. *DOCK8* [Online Mendelian Inheritance in Man (OMIM) 611432] is a gene associated with autosomal dominant mental retardation 2 [50]. Heterozygous disruption of the *DOCK8* gene by deletion or by translocation breakpoints has been reported to cause mental retardation and developmental disability. *ANKRD15* (OMIM 607704) is a maternally imprinted gene that is expressed only from the paternal allele [51]. Deletion of the *ANKRD15* gene causes parent-of-origin-dependent inheritance of familial cerebral palsy that occurs only in individuals inheriting the deletion from the father [51]. *FOXD4* (OMIM 601092) encodes forkhead box D4 protein, which is a transcription factor. *FOXD4* is expressed in the heart, skeletal muscles, and brain [52]. Mutations in the human *FOXD4* gene can cause a complex phenotype consisting of dilated cardiomyopathy, obsessive-compulsive disorders, and suicidality [53].

Distal 9p deletion can be associated with 46,XY gonadal dysgenesis and sex reversal [54–56]. There are three *DMRT* genes, namely *DMRT1*, *DMRT2*, and *DMRT3* genes, all at 9p24.3. *DMRT1* and *DMRT2* are well-known sex-determining genes [54,56], but the role of *DMRT3* on sex determination is not clear at the present time. *DMRT1* (OMIM 602424)

encodes doublesex- and MAB3-related transcription factor 1, which is a male-specific transcriptional regulator involved in sex determination and differentiation [57,58]. *DMRT2* (OMIM 604935) encodes DMRT2 protein, which is also associated with gonadal dysgenesis and XY sex reversal [54,59]. In the present case, only *DMRT1* and *DMRT3* were deleted, and *DMRT2* was intact. Our fetus showed normal male external genitalia without sex reversal. Although *DMRT1* gene is involved in sex development, incomplete penetrance with differences in the phenotype of abnormal male sex development still exists [60]. Patients with a 9p24 deletion have been reported to manifest normal male external genitalia and/or mild gonadal abnormalities [33,61–63]. Barbaro et al [60] suggested that a more complicated mechanism should be hypothesized to explain the variable penetrance. In addition to *DMRT*, other sex-determining genes, such as *SF1*, *DAX1*, *WT1*, *WNT4*, and *SOX9* are associated with male gonadal development. Investigations of the interaction among those sex-determining genes and the gene dosage threshold effect are required for further elucidation of the complicated mechanism of the variable penetrance.

Acknowledgments

This work was supported by research grants NSC-96-2314-B-195-008-MY3 and NSC-97-2314-B-195-006-MY3 from the National Science Council and MMH-E-99004 from Mackay Memorial Hospital, Taipei, Taiwan.

References

- [1] Ballif BC, Yu W, Shaw CA, Kashork CD, Shaffer LG. Monosomy 1p36 breakpoint junctions suggest pre-meiotic breakage-fusion-bridge cycles are involved in generating terminal deletions. *Hum Mol Genet* 2003;12: 2153–65.
- [2] Tonk VS, Wilson GN, Yatsenko SA, Stankiewicz P, Lupski JR, Schutt RC, et al. Molecular cytogenetic characterization of a familial der (1)del(1)(p36.33)dup(p36.33p36.22) with variable phenotype. *Am J Med Genet* 2005;139A:136–40.
- [3] Mewar R, Harrison W, Weaver DD, Palmer C, Davee MA, Overhauser J. Molecular cytogenetic determination of a deletion/duplication of 1q that results in a trisomy 18 syndrome-like phenotype. *Am J Med Genet* 1994; 52:178–83.
- [4] de Brasi D, Rossi E, Giglio S, D'Agostino A, Titomanlio L, Farina V, et al. Inv dup del(1)(pter→q44::q44→q42:) with the classical phenotype of trisomy 1q42-qter. *Am J Med Genet* 2001;104:127–30.
- [5] Gruchy N, Jacquemont ML, Lyonnet S, Labrune P, El Kamel I, Siffroi JP, et al. Recurrent inverted duplication of 2p with terminal deletion in a patient with the classical phenotype of trisomy 2p23-pter. *Am J Med Genet* 2007;143A:2417–22.
- [6] Bonaglia MC, Giorda R, Massagli A, Galluzzi R, Ciccone R, Zuffardi O. A familial inverted duplication/deletion of 2p25.1-25.3 provides new clues on the genesis of inverted duplications. *Eur J Hum Genet* 2009;17: 179–86.
- [7] Bonaglia MC, Giorda R, Poggi G, Raggi ME, Rossi E, Baroncini A, et al. Inverted duplications are recurrent rearrangements always associated with a distal deletion: Description of a new case involving 2q. *Eur J Hum Genet* 2000;8:597–603.
- [8] Aldred MA, Sanford ROC, Thomas NS, Barrow MA, Wilson LC, Brueton LA, et al. Molecular analysis of 20 patients with 2q37.3 monosomy: definition of minimum deletion intervals for key phenotypes. *J Med Genet* 2004;41:433–9.
- [9] Cuscó I, del Campo M, Vilardell M, González E, Gener B, Galán E, et al. Array-CGH in patients with Kabuki-like phenotype: Identification of two patients with complex rearrangements including 2q37 deletions and no other recurrent aberration. *BMC Med Genet* 2008;9:27.
- [10] Vera-Carbonell A, López-Expósito I, Bafalliu JA, Ballesta-Martínez M, Glóver G, Llópis C, et al. Molecular characterization of a new patient with a non-recurrent inv dup del 2q and review of the mechanisms for this rearrangement. *Am J Med Genet* 2010;152A:2670–80.
- [11] Jenderny J, Poetsch M, Hoeltzenbein M, Friedrich U, Jauch A. Detection of a concomitant distal deletion in an inverted duplication of chromosome 3. Is there an overall mechanism for the origin of such duplications/deficiencies? *Eur J Hum Genet* 1998;6:439–44.
- [12] Kennedy D, Silver MM, Winsor EJT, Toi A, Provias J, Macha M, et al. Inverted duplication of the distal short arm of chromosome 3 associated with lobar holoprosencephaly and lumbosacral meningomyelocele. *Am J Med Genet* 2000;91:167–70.
- [13] Cotter PD, Kaffe S, Li L, Gershin IF, Hirschhorn K. Loss of subtelomeric sequence associated with a terminal inversion duplication of the short arm of chromosome 4. *Am J Med Genet* 2001;102:76–80.
- [14] Kondoh Y, Toma T, Ohashi H, Harada N, Yoshiura K, Ohta T, et al. Inv dup del (4)(:p14→p16.3::p16.3→qter) with manifestations of partial duplication 4p and Wolf-Hirschhorn syndrome. *Am J Med Genet* 2003; 120A:123–6.
- [15] Paskulin GA, Riegel M, Cotter PD, Kiss A, Rosa RF, Zen PR, et al. Inv dup del (4)(:p13→p16.3::p16.3→qter) in a girl without typical manifestations of Wolf-Hirschhorn syndrome. *Am J Med Genet* 2009;149A: 1302–7.
- [16] Van Buggenhout G, Maas NMC, Fryns JP, Vermeesch JR. A dysmorphic boy with 4qter deletion and 4q32.3-34.3 duplication: Clinical, cytogenetic, and molecular findings. *Am J Med Genet* 2004;131A:186–9.
- [17] Sreekantaiah C, Kronn D, Marinescu RC, Goldin B, Overhauser J. Characterization of a complex chromosomal rearrangement in a patient with a typical catlike cry and no other clinical findings of cri-du-chat syndrome. *Am J Med Genet* 1999;86:264–8.
- [18] Wang J-C, Coe BP, Lomax B, MacLeod PM, Parslow MI, Schein JE, et al. Inverted duplication with terminal deletion of 5p and no cat-like cry. *Am J Med Genet* 2008;146A:1173–9.
- [19] Martinet D, Filges I, Besuchet Schmutz N, Morris MA, Gaide AC, Dahoun S, et al. Subtelomeric 6p deletion: clinical and array-CGH characterization in two patients. *Am J Med Genet* 2008;146A:2094–102.
- [20] Hoo JJ, Chao M, Szego K, Rauer M, Echiverri SC, Harris C. Four new cases of inverted terminal duplication: a modified hypothesis of mechanism of origin. *Am J Med Genet* 1995;58:299–304.
- [21] Stetten G, Charity LL, Kasch LM, Scott AF, Berman CL, Pressman E, et al. A paternally derived inverted duplication of 7q with evidence of a telomeric deletion. *Am J Med Genet* 1997;68:76–81.
- [22] Dill FJ, Schertzer M, Sandercock J, Tischler B, Wood S. Inverted tandem duplication generates a duplication deficiency of chromosome 8p. *Clin Genet* 1987;32:109–13.
- [23] Mitchell JJ, Vekemans M, Luscombe S, Hayden M, Weber B, Richter A, et al. U-type exchange in a paracentric inversion as a possible mechanism of origin of an inverted tandem duplication of chromosome 8. *Am J Med Genet* 1994;49:384–7.
- [24] Floridia G, Piantanida M, Minelli A, Dellavecchia C, Bonaglia C, Rossi E, et al. The same molecular mechanism at the maternal meiosis I produces mono- and dicentric 8p duplications. *Am J Hum Genet* 1996;58:785–96.
- [25] Fan Y-S, Siu VM. Molecular cytogenetic characterization of a derivative chromosome 8 with an inverted duplication of 8p21.3→p23.3 and a rearranged duplication of 8q24.13→qter. *Am J Med Genet* 2001;102: 266–71.
- [26] Giglio S, Broman KW, Matsumoto N, Calvari V, Gimelli G, Neumann T, et al. Olfactory receptor-gene clusters, genomic-inversion polymorphisms, and common chromosome rearrangements. *Am J Hum Genet* 2001;68: 874–83.
- [27] Shimokawa O, Kurosawa K, Ida T, Harada N, Kondoh T, Miyake N, et al. Molecular characterization of inv dup del (8p): Analysis of five cases. *Am J Med Genet* 2004;128A:133–7.
- [28] Buysse K, Antonacci F, Callewaert B, Loeyls B, Fränkel U, Siu V, et al. Unusual 8p inverted duplication deletion with telomere capture from 8q. *Eur J Med Genet* 2009;52:31–6.
- [29] Hand M, Gray C, Glew G, Tsuchiya KD. Mild phenotype in a patient with mosaic del(8p)/inv dup del(8p). *Am J Med Genet* 2010;152A: 2827–31.
- [30] Teebi AS, Gibson L, McGrath J, Meyn MS, Breg WR, Yang-Feng TL. Molecular and cytogenetic characterization of 9p- abnormalities. *Am J Med Genet* 1993;46:288–92.
- [31] Krepisch-Santos ACV, Vianna-Morgante AM. Disclosing the mechanisms of origin of *de novo* short-arm duplications of chromosome 9. *Am J Med Genet* 2003;117A:41–6.
- [32] Chabchoub E, Rodríguez L, Galán E, Mansilla E, Martínez-Fernández ML, Martínez-Frías ML, et al. Molecular characterisation of a mosaicism with a complex chromosome rearrangement: evidence for coincident chromosome healing by telomere capture and neo-telomere formation. *J Med Genet* 2007;44:250–6.
- [33] Swinkels MEM, Simons A, Smeets DF, Vissers LE, Veltman JA, Pfundt R, et al. Clinical and cytogenetic characterization of 13 Dutch patients with deletion 9p syndrome: delineation of the critical region for a consensus phenotype. *Am J Med Genet* 2008;146A:1430–8.
- [34] Hulick PJ, Noonan KM, Kulkarni S, Donovan DJ, Listewnik M, Ihm C, et al. Cytogenetic and array-CGH characterization of a complex *de novo* rearrangement involving duplication and deletion of 9p and clinical findings in a 4-month-old female. *Cytogenet Genome Res* 2009;126:305–12.
- [35] Mosca AL, Callier P, Faivre L, Marle N, Mejean N, Thauvin-Robinet C, et al. Polymicrogyria in a child with inv dup del (9p) and 22q11.2 microduplication. *Am J Med Genet* 2009;149A:475–81.
- [36] Fisher AM, Thomas NS, Cockwell A, Stecko O, Kerr B, Temple IK, et al. Duplication of chromosome 11p15 of maternal origin result in a phenotype that includes growth retardation. *Hum Genet* 2002;111:290–6.
- [37] Chen CP, Chern SR, Lin SP, Lin CC, Li YC, Wang TH, et al. A paternally derived inverted duplication of distal 14q with a terminal 14q deletion. *Am J Med Genet* 2005;139A:146–50.

- [38] Knijnenburg J, van Haeringen A, Hansson KB, Lankester A, Smit MJ, Belfroid RD, et al. Ring chromosome formation as a novel escape mechanism in patients with inverted duplication and terminal deletion. *Eur J Hum Genet* 2007;15:548–55.
- [39] Genesio R, De Brasi D, Conti A, Borghese A, Di Micco P, Di Costanzo P, et al. Inverted duplication of 15q with terminal deletion in a multiple malformed newborn with intrauterine growth failure and lethal phenotype. *Am J Med Genet* 2004;128A:422–8.
- [40] Leclercq S, Maincent K, Baverel F, Le Tessier D, Letourneur F, Lebbar A, et al. Molecular cytogenetic characterization of the first reported case of inv dup del 20p compatible with a U-type exchange model. *Am J Med Genet* 2009;149A:437–45.
- [41] Pangalos C, Théophile D, Sinet P-M, Marks A, Stambouliéh-Abazis D, Chettouh Z, et al. No significant effect of monosomy for distal 21q22.3 on the Down syndrome phenotype in "mirror" duplications of chromosome 21. *Am J Hum Genet* 1992;51:1240–50.
- [42] Milunsky J, Huang XL, Wyandt HE, Milunsky A. Schizophrenia susceptibility gene locus at Xp22.3. *Clin Genet* 1999;55:455–60.
- [43] Dupont C, Lebbar A, Teinturier C, Baverel F, Viot G, Le Tessier D, et al. First reported case of intrachromosomal cryptic inv dup del Xp in a boy with developmental retardation. *Am J Med Genet* 2007;143A:1236–43.
- [44] Weleber RG, Verma RS, Kimberling WJ, Fieger Jr HG, Lubs HA. Duplication-deficiency of the short arm of chromosome 8 following artificial insemination. *Ann Génét* 1976;19:241–7.
- [45] Federico A, Tomasetti P, Zollino M, Diomedì M, Dotti MT, De Stefano N, et al. Association of trisomy 9p and band heterotopia. *Neurology* 1999;53:430–2.
- [46] Chen CP, Chang TY, Shih JC, et al. Prenatal diagnosis of the Dandy-Walker malformation and ventriculomegaly associated with partial trisomy 9p and distal 12p deletion. *Prenat Diagn* 2002;22:1063–6.
- [47] Chen CP, Shih JC. Association of partial trisomy 9p and the Dandy-Walker malformation. *Am J Med Genet* 2005;132A:111–2.
- [48] D'Agostino MD, Bernasconi A, Das S, Bastos A, Valerio RM, Palmi A, et al. Subcortical band heterotopia (SBH) in males: Clinical, imaging and genetic findings in comparison with females. *Brain* 2002;125:2507–22.
- [49] Temtamy SA, Kamel AK, Ismail S, Helmy NA, Aglan MS, El Gammal M, et al. Phenotypic and cytogenetic spectrum of 9p trisomy. *Genet Couns* 2007;18:29–48.
- [50] Griggs BL, Ladd S, Saul RA, DuPont BR, Srivastava AK. Deducator of cytokinesis 8 is disrupted in two patients with mental retardation and developmental disabilities. *Genomics* 2008;91:195–202.
- [51] Lerer I, Sagi M, Meiner V, Cohen T, Zlotogora J, Abeliovich D. Deletion of the *ANKRD15* gene at 9p24.3 causes parent-of-origin-dependent inheritance of familial cerebral palsy. *Hum Mol Genet* 2005;14:3911–20.
- [52] Pierrou S, Hellqvist M, Samuelsson L, Enerback S, Carlsson P. Cloning and characterization of seven human forkhead proteins: binding site specificity and DNA bending. *EMBO J* 1994;13:5002–12.
- [53] Minoretti P, Arra M, Emanuele E, Olivieri V, Aldeghi A, Politi P, et al. A W148R mutation in the human *FOXD4* gene segregating with dilated cardiomyopathy, obsessive-compulsive disorder, and suicidality. *Int J Mol Med* 2007;19:369–72.
- [54] Muroya K, Okuyama T, Goishi K, Ogiso Y, Fukuda S, Kameyama J, et al. Sex-determining gene(s) on distal 9p: clinical and molecular studies in six cases. *J Clin Endocrinol Metab* 2000;85:3094–100.
- [55] Livadas S, Mavrou A, Sofocleous C, van Vliet-Constantinidou C, Dracopoulou M, Dacou-Voutetakis C. Gonadoblastoma in a patient with del(9)(p22) and sex reversal: report of a case and review of the literature. *Cancer Genet Cytogenet* 2003;143:174–7.
- [56] Öunap K, Uibo O, Zordania R, Kiho L, Ilus T, Öiglane-Shlik E, et al. Three patients with 9p deletions including *DMRT1* and *DMRT2*: a girl with XY complement, bilateral ovotestes, and extreme growth retardation, and two XX females with normal pubertal development. *Am J Med Genet* 2004;130A:415–23.
- [57] Raymond CS, Shamu CE, Shen MM, Seifert KJ, Hirsch B, Hodgkin J, et al. Evidence for evolutionary conservation of sex-determining genes. *Nature* 1998;391:691–5.
- [58] Ying M, Chen B, Tian Y, Hou Y, Li Q, Shang X, et al. Nuclear import of human sexual regulator *DMRT1* is mediated by importin-beta. *Biochim Biophys Acta* 2007;1773:804–13.
- [59] Raymond CS, Parker ED, Kettlewell JR, Brown LG, Page DC, Kusz K, et al. A region of human chromosome 9p required for testis development contains two genes related to known sexual regulators. *Hum Mol Genet* 1999;8:989–96.
- [60] Barbaro M, Balsamo A, Anderlid BM, Myhre AG, Gennari M, Nicoletti A, et al. Characterization of deletions at 9p affecting the candidate regions for sex reversal and deletion 9p syndrome by MLPA. *Eur J Hum Genet* 2009;17:1439–47.
- [61] Hayashi S, Kurosawa K, Imoto I, Mizutani S, Inazawa J. Detection of cryptic chromosome aberrations in a patient with a balanced t(1;9)(p34.2;p24) by array-based comparative genomic hybridization. *Am J Med Genet* 2005;139A:32–6.
- [62] Vinci G, Chantot-Bastaraud S, El Houate B, Lortat-Jacob S, Brauner R, McElreavey K. Association of deletion 9p, 46, XY gonadal dysgenesis and autistic spectrum disorder. *Mol Hum Reprod* 2007;13:685–9.
- [63] Hauge X, Raca G, Cooper S, May K, Spiro R, Adam M, et al. Detailed characterization of, and clinical correlations in, 10 patients with distal deletions of chromosome 9p. *Genet Med* 2008;10:599–611.