Squadinorlignoside: A Novel 7,9'-Dinorlignan from the Stems of *Annona squamosa*

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Two new polar lignans, *i.e.*, squadinorlignoside (=4-[(1E)-1-(hydroxymethyl)-3-(4-hydroxyphenyl)prop-1-en-1-yl]phenyl β -D-glucopyranoside; **1**) and (6R,7R,8S)-7a-[(β -D-glucopyranosyl)oxy]-1-methoxyisolariciresinol (**2**) were isolated from the stems of *Annona squamosa*, together with eight known lignans and five known neolignans (compounds **3–15**; *Fig. 1*). All of these constituents are reported for the first time from the genus *Annona*. The structures, absolute configurations, and selected conformational aspects of the new compounds were elucidated spectroscopically. Compound **1** is the first example of a 7,9'-dinorlignan natural product.

Introduction. – In previous studies, a number of bioactive phytochemicals, including ent-kaurane diterpenoids, alkaloids, annonaceous acetogenins, cyclic peptides, etc., were isolated from Annona squamosa [1-3]. In the present work, we report a series of constituents isolated from the polar fractions and the aqueous layer of the MeOH-soluble extracts of A. squamosa. The following 15 lignans and/or neolignans were isolated (Fig. 1): squadinorlignoside (1)¹), (6R,7S,8S)-7a-[$(\beta$ -D-glucopyranosyl)oxy]-1-methoxyisolariciresinol (2), (6R,7R,8S)-1-methoxyisolariciresinol (3) [4], (6S,7S,8R)-7a- $[\beta$ -D-glucopyranosyl)oxylisolariciresinol (4) [5], (6R,7R,8S)-isolariciresi $nol(5)[6], (6R,7S,8S)-7a-[(\beta-D-glucopyranosyl)oxyl]yoniresinol(6)[7], (6R,7R,8R)-7a [(\beta-D-glucopryanosyl)oxy]$ lyoniresinol (7) [7], $[(2R^*,2'R^*)-secoisolariciresin-4-yl]$ $\beta-D$ glucoside (8) [8], $(2R^*,2'R^*)$ -secoisolariciresinol (9) [9], (7S,8R,8'R)-5,5'-dimethoxylariciresinol (10) [10], (7S,8R)-7,9,9'-trihydroxy-3,3'-dimethoxy-8-O-4'-neolignan-4-O- β -D-glucopyranoside (11) [11], (75,8R)-4,7,9,9'-tetrahydroxy-3,3'-dimethoxy-8-O-4'-neolignan (12) [11], (7S,8R)-urolignoside (13) [12], (7S,8R)-dihydrodehydrodiconiferylalcohol (14) [12], and (7S,8R)-5-methoxydihydrodehydrodiconiferylalcohol (15) [13]. All of these compounds were obtained from *Annona* species for the first time, lignans 1 and 2 being new compounds.

Results and Discussion. – Compound **1**, obtained as syrup, had the molecular formula $C_{22}H_{26}O_8$ based on its HR-FAB-MS data. In the ¹H-NMR spectrum, the resonances of two 1,4-disubstituted Ph groups were observed (δ (H) 6.67, 6.93 (2d, J=8.6 Hz each, 2×2 H); 7.11, 7.19 (2d, J=8.8 Hz each, 2×2 H)). By analyzing the chemical shifts and coupling constants, one olefinic H-atom at δ (H) 5.83 (td, J=7.6, 1.2), and two sets of CH₂ resonances at 3.21 (d, J=7.6) and 4.24 (d, J=1.2 Hz) indicated a trisubstituted

¹⁾ For systematic names of the new compounds 1 and 2, see the Exper. Part.

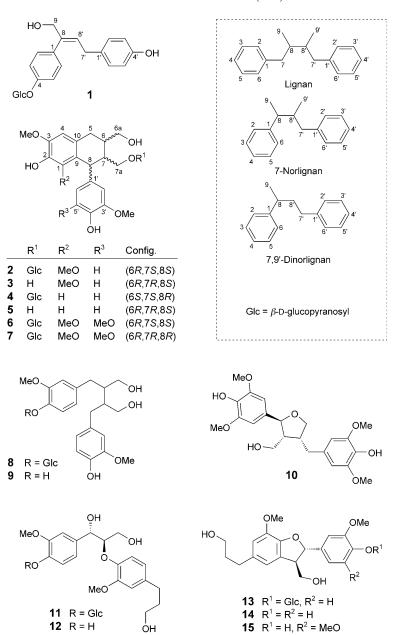


Fig. 1. Structures of compounds 1-15. Basic lignan frameworks and IUPAC atom numbering [14] are shown in the box.

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olefinic group flanked by two CH₂ groups, one of which was oxygenated ($\delta(H)$ 4.24, $\delta(C)$ 67.9). The structure of the aglycone was fully established by 2D-NMR experiments, i.e., ¹H, ¹H-COSY, TOCSY, HMQC, and HMBC spectra, and the configuration

of the olefinic group was assigned by NOESY (Fig. 2). The key NOE correlations of H-C(9)/H-C(8') and H-C(6)/H-C(7') established the (E)-configuration.

Fig. 2. Selected NOESY, HMBC, and ¹H, ¹H-COSY correlations of **1**

The sugar moiety of **1** was found to correspond to a β -D-glucopyranosyloxy (GlcO) residue attached at C(4) of the 7,9'-norlignan skeleton (see *Fig. 1*), as deduced from the HMBC spectrum and from the corresponding EI-MS fragments (*Fig. 3*). Thus, from the above data, the structure of compound **1** was identified as 4-[(1*E*)-1-(hydroxymethyl)-3-(4-hydroxyphenyl)prop-1-en-1-yl]phenyl β -D-glucopyranoside, and the compound was named *squadinorlignoside*. According to the *IUPAC* nomenclature of norlignans [14], this compound has an unprecedented 7,9'-dinorlignan skeleton (*Fig. 1*).

Fig. 3. EI-MS Fragments of 1

Compound **2** was optically active, $[\alpha]_D^{22} = +130.8$ (c=0.026, MeCN), and had the molecular formula $C_{27}H_{36}O_{12}$, as determined by HR-FAB-MS. The ¹³C-NMR spectrum of **2** (see the *Table* in the *Exper. Part*) was very similar to those of the known isolariciresinol-type lignan glycosides **4**, **6**, and **7** [5][7]. The ¹H-NMR spectrum of **2** exhibited signals for one set of *ABX*-type aromatic H-atoms, indicating 1,3,4-trisubstitution (δ (H) 6.76 (d, J=2.0, 1 H); 6.64 (d, J=8.4, 1 H); 6.50 (dd, J=8.4, 2.0 Hz, 1 H)), as well as an aromatic *singlet* at δ (H) 6.57 (1 H). In the ¹H, ¹H-COSY and TOCSY spectra, the partial structure **A** was revealed (*Fig. 4*), and three MeO groups (δ (H) 3.31, 3.77, 3.85) at C(1), C(3), and C(3'), respectively, were identified form the NOE crosspeaks of 1-MeO/H–C(8), 3-MeO/H–C(4), and 3'-MeO/H–C(2') (*Fig. 5*).

Fig. 4. ¹H, ¹H-COSY and TOCSY Correlations for the partial structure A of 2

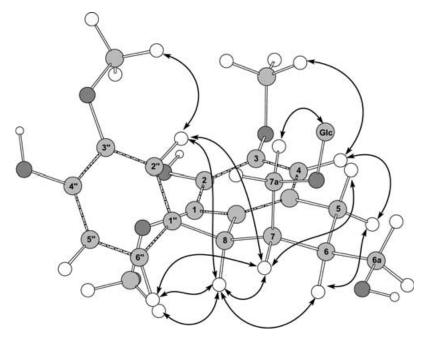


Fig. 5. Selected NOESY correlations of 2. The cross-peak between H-C(7a) and Glc refers to the anomeric H-atom H-C(1'').

The presence of a Glc group in **2** was inferred from its 1 H- and 13 C-NMR spectra. The sugar moiety was attached at C(7a), as deduced from the NOE cross-peaks between the anomeric Glc H-atom (δ (H) 4.27 (d, J=7.6 Hz)) and CH₂(7a) (δ (H) 3.45 (dd, J=9.8, 4.0), 3.89 (dd, J=9.8, 5.6 Hz)).

The above results, in combination with a detailed analysis of the EI-MS fragments of **2** (*Fig.* 6), indicated that the compound was a β -D-glucoside of 1-methoxyisolariciresinol. The relative configurations at C(6) to C(8) were determined by a NOESY experiment (*Fig.* 5), and corroborated by inspection of ${}^{1}H$, ${}^{1}H$ -coupling constants. The NOE correlations between H–C(2',6') and H–C(7), together with a J(7,8) value of 6.4 Hz, indicated that H–C(7) and H–C(8) are in an axial/equatorial (ax/eq) relation. The coupling constants for H–C(5) [H_{ax}–C(5) at δ (H) 2.59 (dd, J=14.8, 11.6 Hz); H_{eq}–C(5) at δ (H) 2.71 (dd, J=14.8, 4.8 Hz)] evidenced that H–C(6) is in an axial orientation. The NOESY cross-peaks of H–C(7) and H–C(8) with H–C(6), and of H_{ax}–C(5) with H–C(7), and the absence of a cross-peak between H–C(5) and H–C(8), and H–C(5) and

Fig. 6. EI-MS Fragments of 2

H-C(2',6'), indicated that the cyclohexane ring of **2** is in a half-envelope conformation (*Fig.* 5), with the relative (6*R**,7*S**,8*S**)-configuration. From circular-dichroism (CD) experiments, the absolute (6*R*,7*S*,8*S*)-configuration was established, based on Δε values of -0.06 and +0.11 at 299 and 275 nm, respectively [15]. From all these data, compound **2** was identified as [(1*S*,2*S*,3*R*)-1,2,3,4-tetrahydro-7-hydroxy-1-(4-hydroxy-3-methoxyphenyl)-3-(hydroxymethyl)-6,8-dimethoxynaphthalen-2-yl]methyl β-D-glucopyranoside.

The other isolated lignanoids 3-15 were structurally elucidated by spectroscopic analysis and comparison with literature data. In previous studies, only furofuran lignans with a 7,9':7',9-diepoxylignan skeleton have been reported from *Annona* species [16][17]. In the present study, three different lignan skeletons were identified: the 2,7'-cyclolignans 2-7, the diarylbutanelignans 8 and 9, and the 7',9'-epoxylignan 10, all of which are biogenetically derived from the furofuran lignans. In addition, five neolignans, 11-15, were isolated for the first time from *Annona*. So far, only one neolignan analogue, named grossamide, has been reported from *Annona* [18].

Experimental Part

General. Silica gel 60 (230–400 mesh; Merck) was used for column chromatography (CC). Prep. HPLC: Develosil ODS and C30-UG-5 columns (250×20 mm) on a JASCO PU-1580 apparatus with a UV-1575 detector.

TLC: Spots were detected by spraying with 50% $\rm H_2SO_4$, and then heated on a hot plate. UV Spectra: $\it JASCOV_530$ spectrophotometer; $\it \lambda_{max}$ in nm. Optical rotations: $\it JASCOP_1020$ digital polarimeter. CD Spectra: $\it JASCOV_530$ spectrophotometer; $\it \lambda(\Delta\epsilon)$ in nm. IR Spectra: a $\it Mattson Genesis-II$ spectrophotometer; in cm $^{-1}$. 1 H-NMR: at 400 or 500 MHz in (D $_6$) acetone or CD $_3$ OD; $\it \delta$ in ppm, $\it J$ in Hz. 13 C-NMR, DEPT, 1 H, 1 H-COSY, TOCSY, HMBC, HMQC, and NOESY Spectra: $\it Varian Unity Plus-400$ and $\it Unity INOVA-500$. EI-MS: $\it Finnigan POLARISQ$ mass spectrometer, with direct-insert probe; HR-FAB-MS: $\it Jeol JMS-HX-110$ mass spectrometer; in $\it m/z$ (rel. %).

Plant Material. Fresh stems of *A. squamosa* were collected from Shueimen, Pingtung County, Taiwan, in May 2000. The plant was identified by Dr. *Hsin-Fu Yen*, National Museum of Natural Science, Taichung, Taiwan. A voucher specimen (Annona 6) was deposited at the Graduate Institute of Natural Products, Kaohsiung Medical University, Kaohsiung, Taiwan.

Extraction and Isolation. Fresh stems of A. squamosa (15 kg) were extracted repeatedly with MeOH at r.t. The combined extracts were evaporated under reduced pressure to yield a dark-brown syrup (550 g), which was partitioned between CHCl₃ and H₂O. Both layers were further processed separately. a) The CHCl₃ layer was extracted with 3% aq. HCl to remove alkaloids. The 'neutral' CHCl₃ soln. was dried and evaporated to leave

Table. NMR Data of the 2,7'-Cyclolignans **2–4**, **6**, and **7**. At 500/125 MHz, resp., in CD₃OD; δ in ppm, J in Hz.

Position	2		3	4	6	7
	$\delta(H)$	δ(C)	$\delta(C)$	$\delta(C)$	$\delta(C)$	$\delta(C)$
1	_	147.5	147.6	117.4	147.6	147.6
2	_	138.9	138.9	145.9	138.9	138.9
3	_	148.6	148.5	147.3	148.6	148.8
4	6.57 (s)	107.9	107.8	112.3	107.9	107.9
5	2.59 $(dd, J=14.8, 11.6)$ 2.71 $(dd, J=14.8, 4.8)$	33.9	33.6	33.6	33.8	33.8
6	$1.68-1.74 \ (m)$	40.6	40.9	41.1	40.6	41.2
6a	3.52 (dd, J=10.8, 6.4) 3.60-3.66 (m)	66.3	66.8	65.5	66.2	66.2
7	$2.04-2.10 \ (m)$	46.8	49.6	45.3	46.7	46.6
7a	3.45 (dd, J=9.8, 4.0) 3.89 (dd, J=9.8, 5.6)	71.5	64.1	70.7	71.5	71.6
8	4.40 (d, J=6.4)	42.4	42.0	48.3a)	42.7	43.2
9	_	126.6	126.4	129.3	126.4	126.2
10		130.2	130.1	138.8	130.2	130.2
1'	_	140.1	140.1	133.7	139.3	139.4
2'	6.76 (d, J=2.0)	113.6	113.4	113.9	106.9	107.1
3′	_	148.7	148.6	149.0	149.0	149.0
4'	_	145.3	145.3	145.2	134.5	134.5
5'	6.64 (d, J = 8.4)	115.7	115.7	116.0	149.0	149.0
6'	6.50 (dd, J = 8.4, 2.0)	121.7	121.7	123.5	106.9	107.1
1"	4.27 (d, J=7.6)	104.8	_	103.8	104.8	104.2
2"	3.20-3.70	75.2	_	75.0	75.2	75.1
3"	3.20-3.70	78.2	_	78.2	78.2	78.2
4''	3.20-3.70	71.7	_	71.4	71.7	72.0
5''	3.20-3.70	77.9	_	77.8	77.9	78.0
6"	3.63-3.69 (<i>m</i>) 3.81 (<i>dd</i> , <i>J</i> =10.0, 2.0)	62.8	-	62.4	62.8	62.7
1-MeO	3.31 (s)	60.1	60.1	_	60.2	60.1
3-MeO	3.85(s)	56.6	56.6	56.5	56.6	56.6
3'-MeO	3.77 (s)	56.5	56.3	56.4	56.8	56.8
5'-MeO	-	-	-	-	56.8	56.8

^a) Overlapping with solvent peak.

a brownish, viscous residue (160 g), which was subjected to CC (SiO₂; CHCl₃/MeOH mixtures of increasing polarity): 22 fractions (Fr.) on the basis of TLC. Fr. 20 was subjected to HPLC to afford 15 subfractions: Fr. 20.1–20.15. Compounds 12 (13 mg), 15 (19 mg), and 14 (18 mg) were isolated by PR-HPLC (C18; H₂O/MeCN 80:20) from Fr. 20.6, Fr. 20.14, and Fr. 20.15, resp. Compounds 3 (5 mg), 5 (6 mg), and 10 (7 mg) were obtained by RP-HPLC (C30; H₂O/MeCN 80:20) from Fr. 20.7, Fr. 20.8, and Fr. 20.13, resp. Further purification of Fr. 21 by RP-HPLC (C30; H₂O/MeCN 80:20) yielded compound 9 (10 mg).

b) The original aq. extract (see above) was subjected to CC (Diaion HP-20; H₂O/MeOH): Fr. A1-A5. Fr. A3 (eluted with H₂O/MeOH 1:1) was partitioned between CHCl₃ and H₂O, and the aq. layer was re-extracted with AcOEt. The resulting AcOEt layer was subjected to CC (SiO₂; AcOEt/MeOH 10:1): Fr. A3.1-A3.14. Fr. A3.10 was further separated into nine subfractions: Fr. A3.10-1-A3.10-9. Recyclic RP-HPLC (C30, MeCN/H₂O 30:70) of Fr. A3.10-4 afforded 6 (4 mg). Compounds 1 (3 mg), 2 (4 mg), 4 (4 mg), 7 (5 mg), 8 (4 mg), 11 (5 mg), and 13 (16 mg) were obtained by recyclic RP-HPLC (C30; MeCN/H₂O 15:85) from the subfractions -2, -3, -5, -6, -7, -8, and -9, resp., of Fr. A3.10.

Squadinorlignoside (=4-[(1E)-1-(Hydroxymethyl)-3-(4-hydroxyphenyl)prop-1-en-1-yl]phenyl β-D-glucopyranoside; 1). Syrup. UV (MeCN): 195, 225 (sh), 274. IR (neat): 3415, 1618, 1520. 1 H-NMR (400 MHz; CD₃OD)²): 3.21 (d, J=7.6, CH₂(7')); 3.40 (m, H-C(4")); 3.43 (m, H-C(5")); 3.47 (m, H-C(2",3")); 3.70 (dd, J=12.0, 5.6, H_a-C(6")); 3.90 (dd, J=12.0, 2.4, H_b-C(6")); 4.24 (d, J=1.2, CH₂(9)); 4.90 (overlapping, H-C(1")); 5.83 (dd, J=7.6, 1.2, H-C(8")); 6.67 (d, J=8.6, H-C(3',5")); 6.93 (d, J=8.6, H-C(2',6")); 7.11 (d, J=8.8, H-C(3,5)); 7.19 (d, J=8.8, H-C(2,6)). 13 C-NMR (125 MHz, CD₃OD): 34.8 (C(7')); 62.5 (C(6")); 67.9 (C(9)); 71.4 (C(4")); 74.9 (C(2")); 78.0 (C(3")); 78.1 (C(5")); 102.3 (C(1")); 116.2 (C(3',5")); 117.5 (C(3,5)); 127.9 (C(8')); 130.2 (C(2',6')); 130.9 (C(2,6)); 133.1 (C(1')); 134.1 (C(1)); 141.7 (C(8)); 156.5 (C(4')); 158.2 (C(4)). EI-MS: 256 (10), 239 (57), 238 (100), 225 (40), 145 (22), 131 (47), 107 (73). HR-ESI-MS: 441.1540 ([M+Na] $^+$, C₂₂H₂₆NaO $_8^+$; calc. 441.1525).

(6R, 7R, 8S)-7a-[(β-D-Glucopyranosyl)oxy]-1-methoxyisolariciresinol (=[(1S, 2S, 3R)-1,2,3,4-Tetrahydro-7-hydroxy-1-(4-hydroxy-3-methoxyphenyl)-3-(hydroxymethyl)-6,8-dimethoxynaphthalen-2-yl]methyl β-D-glucopyranoside; **2**). Syrup. UV (MeCN): 201, 232 (sh), 282. [α]_D²² = +130.8 (c=0.026, MeCN). CD (MeCN): 299 (-0.06), 275 (+0.11), 249 (+0.28). IR (neat): 3400, 1620, 1515. 1 H- and 13 C-NMR: see the *Table*. EI-MS: 552 (5, M⁺), 390 (35), 389 (29), 372 (91), 371 (100), 340 (50), 218 (21), 210 (23). HR-FAB-MS: 575.2108 ([M+Na] $^{+}$; C₂₇H₃₆NaO $^{+}$ ₁₂; calc. 575.2105).

REFERENCES

- [1] Y. L. Yang, F. R. Chang, C. C. Wu, W. Y. Wang, Y. C. Wu, J. Nat. Prod. 2002, 65, 1462.
- [2] M. C. Zafra-Polo, B. Figadère, T. Gallardo, J. R. Tormo, D. Cortes, Phytochemistry 1998, 48, 1087.
- [3] H. Morita, Y. Sato, J. Kobayashi, Tetrahedron 1999, 55, 7509.
- [4] C. Rajendiran, B. R. Pai, P. S. Subramanian, *Indian J. Chem.*, Sect. B 1991, 30, 681.
- [5] M. Wang, J. Li, M. Rangarajan, Y. Shao, E. J. LaVoie, T. C. Huang, C. T. Ho, J. Agric. Food Chem. 1998, 46, 4869
- [6] E. Okuyama, K. Suzumura, M. Yamazaki, Chem. Pharm. Bull. 1995, 43, 2200.
- [7] K. Ohashi, H. Watanabe, Y. Okumura, T. Uji, I. Kitagawa, Chem. Pharm. Bull. 1994, 42, 1924.
- [8] S. Matsuura, M. Iinuma, Phytochemistry 1985, 24, 626.
- [9] A. Buske, J. Schmidt, A. Porzel, G. Adam, Eur. J. Org. Chem. 2001, 18, 3537.
- [10] H. Achenbach, M. Stocker, M. A. Constenla, *Phytochemistry* 1988, 27, 1835.
- [11] N. Matsuda, M. Kikuchi, Chem. Pharm. Bull. 1996, 44, 1676.
- [12] Y. C. Shen, P. W. Hsieh, Y. H. Kuo, Phytochemistry 1998, 48, 719.
- [13] A. Masakazu, S. Akira, Mokuzai Gakkaishi 1978, 24, 422.
- [14] G. P. Moss, Pure Appl. Chem. 2000, 72, 1493.
- [15] P. B. Hulbert, W. Klyne, P. H. Scopes, J. Chem. Res., Miniprint 1981, 401.
- [16] C. Y. Chen, T. Y. Wu, F. R. Chang, Y. C. Wu, J. Chin. Chem. Soc. 1998, 45, 629.
- [17] Y. C. Wu, G. Y. Chang, F. N. Ko, C. M. Teng, Planta Med. 1995, 61, 746.
- [18] L. P. Santos, M. A. D. Boaventura, A. B. Oliveira, J. M. Cassady, *Planta Med.* 1996, 72, 76.

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For atom numbering, see Fig. 1. Doubly primed atoms refer to the Glc moiety, the 1"-postion corresponding to the anomeric center.