

Gene-expression Profiling in a Patient with Esophageal Squamous Cell Carcinoma

Nan-Yung Hsu, I-Ping Chiang¹, Chih-Shiung Shih, Pin-Ru Chen, Heng-Chien Ho²

Division of Chest Surgery, ¹Department of Pathology, ²Department of Medical Research, China Medical University Hospital, Taichung, Taiwan, R.O.C.

A 32-year-old man who has been an active tobacco smoker, alcohol drinker and betel nut chewer for more than 10 years was diagnosed as having esophageal squamous cell carcinoma (ESCC) after undergoing transhiatal esophagectomy and reconstruction with gastric graft. The pathology report indicated that the tumor had invaded the superficial muscle layer but had not metastasized to the adjacent lymph nodes. Gene-expression in the ESCC tissue was profiled by the cDNA microarray method. A total of 8354 genes were screened. Among them, 26 genes whose expression levels (fluorescence intensity) in esophageal carcinoma were more than 5 times (up regulated) those expressed in normal esophageal epithelium were selected; in addition, 28 genes whose expression levels in esophageal carcinoma were less than 0.2 times (down regulated) those expressed in the normal esophageal epithelium were selected. A search on MEDLINE of papers published from 1965 to the present revealed that only 7 of the 26 up-regulated genes and only 8 of the 28 down-regulated genes had been reported. We suggest that these characteristic genes will provide useful information for understanding the malignant nature of ESCC occurring in a young patient who has actively consumed tobacco, alcohol, and betel nut. (**Mid Taiwan J Med 2005;10:43-8**)

Key words

esophageal squamous cell carcinoma, microarray

INTRODUCTION

The cDNA microarray technique has been used to profile gene-expression in esophageal squamous cell carcinoma (ESCC); the results have helped uncover the molecular mechanisms of carcinogenesis [1-3], tumor progression [4,5], and chemotherapy sensitivity [6] in ESCC patients. We encountered ESCC in a young patient who actively chewed betel nut, drank alcohol, and smoked tobacco. The purpose of this study was to profile the genes expressed in this young patient with ESCC by cDNA microarray.

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Address reprint requests to : Heng-Chien Ho, Department of Medical Research, China Medical University Hospital, 2 Yuh-Der Road, Taichung 404, Taiwan, R.O.C.

CASE REPORT

A 32-year-old man with a 10-year history of smoking, alcohol drinking, and betel nut chewing presented with acute abdomen. At admission, his serum amylase was 431 U/L and his r-GT was 102 IU/L. Alcoholic pancreatitis was suspected. Subsequent endoscopy revealed an esophageal carcinoma 1 cm above the esophagogastric junction. The patient underwent transhiatal esophagectomy and the esophagus was reconstructed by gastric tube. The postoperative course was uneventful. Subsequent histopathological studies showed a poorly differentiated squamous cell carcinoma without adjacent lymph node metastasis. The tumor, measuring 0.7 cm × 0.5 cm × 0.5 cm, had

Table 1. Gene-expression profiling (fluorescence intensity) in esophageal carcinoma compared with the normal esophageal epithelium

| Gene name | Accession* | Ratio of mean [†] | Genetic locus |
|--|------------|----------------------------|---------------|
| Up regulation | | | |
| Metabolism/Redox enzymes | | | |
| Ornithine decarboxylase 1 | AA460115 | 13.420 | 2p25 |
| Aldehyde dehydrogenase | AA664101 | 7.680 | 9q21.13 |
| Glutathione peroxidase | AA135152 | 5.721 | 14q24.1 |
| Argininosuccinate synthetase | AA676466 | 5.707 | 9q34 |
| Oligoadenylate synthetase | AA146772 | 5.675 | 12q24.1 |
| Arylsulfatase B | AA128825 | 5.140 | 5p11 |
| ESTs | AA143342 | 12.037 | 21q22.1 |
| Parathyroid hormone-like hormone | AA845432 | 5.166 | 12p12.1 |
| Lactotransferrin | AA677706 | 5.132 | 3q21 |
| Cell Proliferation/Inflammation | | | |
| Gamma-aminobutyric acid A receptor-epsilon | H63934 | 6.250 | Xq28 |
| Tryptophanyl-tRNA synthetase | AA664040 | 8.003 | 14q32.31 |
| Replication factor C | H54751 | 6.414 | 7q11.23 |
| Chromosome 10 open reading frame 3 | AA131908 | 5.979 | 10q23.33 |
| Cell adhesion protein | | | |
| Integrin-alpha 6 | R17993 | 5.469 | 2q31 |
| Pleckstrin homology-Sec7 | AA480859 | 5.054 | 17q25 |
| Cell Cycle regulator | | | |
| Insulin-like growth factor binding protein 3 | AA598601 | 5.347 | 7p13 |
| Retinol binding protein 1-cellular | AA700832 | 5.225 | 3q23 |
| Fas-associated via death domain | AA430751 | 13.114 | 11q13.3 |
| Jun D proto-oncogene | AA131585 | 5.688 | 19p13.2 |
| 2'-5'-oligoadenylate synthetase 2-69/71kDa | AF074990 | 5.227 | 12q24.2 |
| Intracellular signalling | | | |
| Myelin basic protein | N62730 | 18.838 | 18q23 |
| Interferon-alpha-inducible protein | AA406019 | 8.548 | 11p13 |
| Cell membrane receptor | | | |
| Hypothetical protein FLJ10261 | AA400086 | 18.097 | 11q13.2 |
| Others | | | |
| Collagen-type I-alpha 1 | R48843 | 6.645 | 17q21.3 |
| Cytoskeleton-associated protein 1 | AA504477 | 5.244 | 19q13.11 |
| Hypothetical protein LOC90850 | AA620404 | 18.996 | 16p13.3 |
| Down regulation | | | |
| Metabolism/Redox enzymes | | | |
| Hydroxyprostaglandin dehydrogenase 15-(NAD) | AA775223 | 0.175 | 4q34 |
| Transglutaminase | N90882 | 0.186 | 20q11.2 |
| Myosin, light polypeptide 9, regulatory | AA877166 | 0.137 | 20q11.23 |
| DKFZP566F084 protein | AA773458 | 0.139 | 10q24.32 |
| Deiodinase-iodothyronine-type II | AA864322 | 0.174 | 14q24.2 |
| Arachidonate 15-lipoxygenase | AI474406 | 0.198 | 17p13 |
| Cell Proliferation | | | |
| Eukaryotic translation initiation factor 4 gamma-1 | R12349 | 0.161 | 3q27 |
| Hypothetical protein LOC253827 | N91811 | 0.138 | 12q14.1 |
| Hypothetical protein MGC2668 | AA424545 | 0.174 | 9q34.13 |
| Cell Cycle regulator | | | |
| Synaptopodin 2 | W52186 | 0.034 | 4q27 |
| Retinoic acid binding protein 2 | AA598508 | 0.197 | 1q21.3 |
| Mitogen-activated protein kinase | H01340 | 0.148 | 19q13.2 |
| Four and a half LIM domains 1 | AA456394 | 0.149 | Xq26 |
| Intracellular signaling | | | |
| Mal-T-cell differentiation protein | AA227594 | 0.172 | 2cen-q13 |
| p55-GAMMA | AI394701 | 0.181 | 1p34.1 |
| BMP-2 inducible kinase | R98008 | 0.184 | 4q21.23 |

*Genbank ID. [†]Wavelength intensity of tumor (635 nm) and normal tissue (532 nm).

Table 1. Continued

| Gene name | Accession* | Ratio of mean [†] | Genetic locus |
|--|------------|----------------------------|---------------|
| Cell adhesion protein | | | |
| Sorbin and SH3 domain containing 1 | AA459944 | 0.055 | 10q23.3 |
| Phosphoglucomutase 5 | AA706788 | 0.064 | 9p12 |
| Desmuslin | AA877815 | 0.076 | 15q26.3 |
| Alpha-actinin-2-associated LIM protein | AA972352 | 0.177 | 4q35 |
| Cell membrane receptor | | | |
| Brain cell membrane protein 1 | N52772 | 0.183 | Xp11.4 |
| Endothelin receptor type B | H28710 | 0.189 | 13q22 |
| Others | | | |
| Cytokeratin 4 | AA629189 | 0.140 | 12q12 |
| Collagen-type VIII | AA872420 | 0.170 | 3q12.3 |
| Cytokeratin 1 | AA706022 | 0.174 | 12q12 |
| Serine protease inhibitor | Hs.331555 | 0.148 | 5q32 |
| Caldesmon 1 | AA402898 | 0.173 | 7q33 |
| Alpha-fetoprotein | T56880 | 0.174 | 4q11 |

Table 2. A search of MEDLINE for papers published from 1965 to the present revealed that 7 of the 26 up-regulated genes and 8 of the 28 down-regulated genes had been reported

| Number | Up-regulated genes | Down-regulated genes |
|--------|----------------------------------|----------------------------------|
| 1 | Ornithine decarboxylase | Transglutaminase |
| 2 | Aldehyde dehydrogenase | Serine protease inhibitor |
| 3 | Glutathione peroxidase | Retinoic acid binding protein |
| 4 | Parathyroid hormone-like hormone | Cytokeratin 4 |
| 5 | Lactotransferrin | Mitogen-activated protein kinase |
| 6 | Integrin-alpha 6 | Arachidonate 15-lipoxygenase |
| 7 | Fas-associated via death domain | Cytokeratin 1 |
| 8 | | Alpha-fetoprotein |

infiltrated the superficial muscle layer. The pathological stage was T2N0M0 (IIA). The patient has been well for one year after the surgery without any sign of recurrence.

Gene expression in the ESCC tissue was profiled by the cDNA microarray method. In brief, total RNA was extracted from the normal and tumor tissues by TRIzol reagent. The isolated RNA was qualified by gel analysis and the ratio of absorbance was determined at 260 nm and 280 nm (> 1.7). Thirty micrograms of total RNA from normal and tumor tissues were Cy3- and Cy5-labeled, respectively. PCR products were arrayed by a Microgrid II pro arrayer (Biorobotics Ltd., Cambridgeshire, United Kingdom). The labeled RNA was then hybridized at 62°C for 15.5 h. Arrays were scanned and each spot was defined by automatic positioning of a grid using image-analysis spot-tracking software (patent US 10/173,672 June 19, 2002; CA 2,389,901 June 20, 2002). We focused first on a set of 1416 genes

(about 17.0% of the 8354 genes analyzed, which comprised 922 up-regulated genes and 494 down-regulated genes), whose transcripts varied in abundance by at least twofold expression levels (fluorescence intensity) from their mean abundance in esophageal carcinoma compared with the normal esophageal epithelium. Among the 922 up-regulated genes, 26 (2.8%) genes whose expression levels in esophageal carcinoma were more than 5 times the levels expressed in normal esophageal epithelium were selected. In addition, 28 (5.7%) of the 494 down-regulated genes whose expression levels in esophageal carcinoma were less than 0.2 times the levels expressed in normal esophageal epithelium were selected. These genes were classified according to their biologic roles, such as metabolism, cell proliferation, cell adhesion protein, cell cycle regulator, intracellular signaling and transmembrane protein (Table 1). A search of MEDLINE for papers published from 1965 to

the present revealed that only 7 of the 26 up-regulated genes and only 8 of the 28 down-regulated genes have been reported to be associated with gene expression in ESCC (Table 2).

DISCUSSION

The major methods for screening and identifying specific gene(s) involved in carcinogenesis, disease progression and resistance of treatment of ESCC comprise microarray, cDNA subtraction, serial analysis of gene expression (SAGE), differential display and proteomics [6-8]. Although cDNA microarray technology has limitations, such as chip to chip variation, unanswered biological relevance of fold change, cross-hybridization of sequences with high identity, and difficulty of mRNA levels reflecting protein levels [9], it has enabled the expression profiles of thousands of genes to be analyzed simultaneously. In addition, although the correlation between clinicopathological phenotypes and gene expression status has not been completely proven, this technique can provide a powerful means to stratify tumors into classes with distinct molecular pathophysiologies [5].

Several of the genes expressed in ESCC tissue identified in the literature were also found in this study. For example, ornithine decarboxylase was the most highly (13.4-fold) expressed up regulated gene in this study [10]. In addition, ornithine decarboxylase has also been reported to be one of the transcriptional genes expressed in esophageal adenocarcinoma [11]. The Fas-associated via death domain (FADD), which was abundantly expressed in our study, has been reported to be associated with an increased risk of developing ESCC [12,13]. Interestingly, it has been reported that both transglutaminase and cytokeratin were down-regulated genes in ESCC in Chinese [14]. The clinical relevance of those two genes needs to be clarified.

It is still unclear whether the disease process in young patients with ESCC is more aggressive and has a worse long-term outcome than in older patients [15,16]. It has been reported

that esophageal carcinogenesis differs between the young and the elderly [17]. Whether there is any significant difference in gene-expression profiling between young patients and older patients with ESCC needs to be explored in the future.

In this study, tobacco smoking was defined as one pack or more per month; alcohol consumption was defined as consumption of at least one alcoholic beverage (equivalent to 125 mL of normal strength beer) per week; betel nut chewing was defined as consumption of at least 10 betel nuts per week [18]. The patient in this study was defined as an active tobacco smoker, alcohol drinker and betel nut chewer. A study of Taiwanese who smoke, drink alcohol and chew betel nut revealed that 89% of betel nut chewers also were tobacco smokers, and that 66% of male and 61% of female betel nut chewers also drank alcohol [18]. Clinically, it has been reported that the synergetic effect of betel nut and tobacco induces a higher risk of cancer of the oral cavity, pharynx, and esophagus [19,20]; however, relatively limited information is available on gene-expression profiling regarding the synergetic effect of these three habits on ESCC.

We were able to profile the genes expressed in ESCC tissue occurring in one patient. In the near future, we will try to confirm the validity of the microarray results by qualitative reverse transcription-PCR and immunohistochemical evaluation of the gene products whose levels were assessed to be differentially expressed in normal and tumor cells in this patient.

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發生在一位年輕食道癌病人的基因表現資料

許南榮 江宜平¹ 施志勳 陳品儒 何恆堅²

中國醫藥大學附設醫院 胸腔外科 病理部¹ 醫學研究部²

一位32歲男性病人有抽煙，喝酒和嚼檳榔10年以上的病史，因診斷在食道下段有鱗狀細胞癌而接受經食道裂孔切除和胃重建。根據病理檢查，腫瘤侵犯至表皮肌肉層，而無周圍淋巴腺轉移。於是，食道鱗狀細胞癌組織以cDNA微陣列方法，進行基因表現資料分析。一共篩選8354個基因，其中比正常組織多5倍螢光強度的基因表現有26個基因，而少5倍的有28個基因。經由資料搜查，發現26個基因中，僅有7個；而28個基因中，僅有8個已發表在文獻中。我們建議這些基因的特質可以針對具有抽煙，喝酒和嚼檳榔病史的年輕食道癌病人，提供食道鱗狀細胞癌惡化本質有用的資料。(中台灣醫誌 2005;10:43-8)

關鍵詞

食道鱗狀細胞癌，微陣列

聯絡作者：何恆堅

地址：404台中市北區育德路2號

中國醫藥大學附設醫院 醫學研究部

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